SECRETED AND TRANSMEMBRANE POLYPEPTIDES AND NUCLEIC ACIDS ENCODING THE SAME

FIELD OF THE INVENTION

The present invention relates generally to the identification and isolation of novel DNA and to the recombinant production of novel polypeptides.

BACKGROUND OF THE INVENTION

Extracellular proteins play important roles in, among other things, the formation, differentiation and maintenance of multicellular organisms. The fate of many individual cells, e.g., proliferation, migration, differentiation, or interaction with other cells, is typically governed by information received from other cells and/or the immediate environment. This information is often transmitted by secreted polypeptides (for instance, mitogenic factors, survival factors, cytotoxic factors, differentiation factors, neuropeptides, and hormones) which are, in turn, received and interpreted by diverse cell receptors or membrane-bound proteins. These secreted polypeptides or signaling molecules normally pass through the cellular secretory pathway to reach their site of action in the extracellular environment.

Secreted proteins have various industrial applications, including as pharmaceuticals, diagnostics, biosensors and bioreactors. Most protein drugs available at present, such as thrombolytic agents, interferons, interleukins, erythropoietins, colony stimulating factors, and various other cytokines, are secretory proteins. Their receptors, which are membrane proteins, also have potential as therapeutic or diagnostic agents. Efforts are being undertaken by both industry and academia to identify new, native secreted proteins. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci. 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

Membrane-bound proteins and receptors can play important roles in, among other things, the formation, differentiation and maintenance of multicellular organisms. The fate of many individual cells, e.g., proliferation, migration, differentiation, or interaction with other cells, is typically governed by information received from other cells and/or the immediate environment. This information is often transmitted by secreted polypeptides (for instance, mitogenic factors, survival factors, cytotoxic factors, differentiation factors, neuropeptides, and hormones) which are, in turn, received and interpreted by diverse cell receptors or membrane-bound proteins. Such membrane-bound proteins and cell receptors include, but are not limited to, cytokine receptors, receptor kinases, receptor phosphatases, receptors involved in cell-cell interactions, and cellular adhesin molecules like selectins and integrins. For instance, transduction of signals that regulate cell growth and differentiation is regulated in part by phosphorylation of various cellular proteins. Protein tyrosine kinases, enzymes that catalyze that process, can also act as growth factor receptors. Examples include fibroblast growth factor receptor and nerve growth factor receptor.

Membrane-bound proteins and receptor molecules have various industrial applications, including as pharmaceutical and diagnostic agents. Receptor immunoadhesins, for instance, can be employed as therapeutic agents to block receptor-ligand interactions. The membrane-bound proteins can also be employed for screening of potential peptide or small molecule inhibitors of the relevant receptor/ligand interaction.

Efforts are being undertaken by both industry and academia to identify new, native receptor or membrane-bound proteins. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel receptor or membrane-bound proteins.

1. **PRO211 and PRO217**

Epidermal growth factor (EGF) is a conventional mitogenic factor that stimulates the proliferation of various types of cells including epithelial cells and fibroblasts. EGF binds to and activates the EGF receptor (EGFR), which initiates intracellular signaling and subsequent effects. The EGFR is expressed in neurons of the cerebral cortex, cerebellum, and hippocampus in addition to other regions of the central nervous system (CNS). In addition, EGF is also expressed in various regions of the CNS. Therefore, EGF acts not only on mitotic cells, but also on postmitotic neurons. In fact, many studies have indicated that EGF has neurotrophic or neuromodulatory effects on various types of neurons in the CNS. For example, EGF acts directly on cultured cerebral cortical and cerebellar neurons, enhancing neurite outgrowth and survival. On the other hand, EGF also acts on other cell types, including septal cholinergic and mesencephalic dopaminergic neurons, indirectly through glial cells. Evidence of the effects of EGF on neurons in the CNS is accumulating, but the mechanisms of action remain essentially unknown. EGF-induced signaling in mitotic cells is better understood than in postmitotic neurons. Studies of cloned pheochromocytoma PC12 cells and cultured cerebral cortical neurons have suggested that the EGF-induced neurotrophic actions are mediated by sustained activation of the EGFR and mitogen-activated protein kinase (MAPK) in response to EGF. The sustained intracellular signaling correlates with the decreased rate of EGFR down-regulation, which might determine the response of neuronal cells to EGF. It is likely that EGF is a multi-potent growth factor that acts upon various types of cells including mitotic cells and postmitotic neurons.

EGF is produced by the salivary and Brunner's glands of the gastrointestinal system, kidney, pancreas, thyroid gland, pituitary gland, and the nervous system, and is found in body fluids such as saliva, blood, cerebrospinal fluid (CSF), urine, amniotic fluid, prostatic fluid, pancreatic juice, and breast milk, Plata-Salaman, *Peptides* 12: 653-663 (1991).

EGF is mediated by its membrane specific receptor, which contains an intrinsic tyrosine kinase. Stoscheck *et al., J. Cell Biochem.* 31: 135-152 (1986). EGF is believed to function by binding to the extracellular portion of its receptor which induces a transmembrane signal that activates the intrinsic tyrosine kinase.

Purification and sequence analysis of the EGF-like domain has revealed the presence of six conserved cysteine residues which cross-bind to create three peptide loops, Savage *et al., J. Biol. Chem.* 248: 7669-7672 (1979). It is now generally known that several other peptides can react with the EGF receptor which share the same generalized motif $X_nCX_7CX_{4/5}CX_{10}CXCX_5GX_2CX_n$, where X represents any non-cysteine amino acid, and n is a variable repeat number. Non isolated peptides having this motif include TGF-α, amphiregulin,

schwannoma-derived growth factor (SDGF), heparin-binding EGF-like growth factors and certain virally encoded peptides (e.g., Vaccinia virus, Reisner, *Nature* 313: 801-803 (1985), Shope fibroma virus, Chang et al., Mol Cell Biol. 7: 535-540 (1987), Molluscum contagiosum, Porter and Archard, *J. Gen. Virol.* 68: 673-682 (1987), and Myxoma virus, Upton *et al.*, *J. Virol.* 61: 1271-1275 (1987), Prigent and Lemoine, *Prog. Growth Factor Res.* 4: 1-24 (1992).

EGF-like domains are not confined to growth factors but have been observed in a variety of cell-surface and extracellular proteins which have interesting properties in cell adhesion, protein-protein interaction and development, Laurence and Gusterson, *Tumor Biol.* 11: 229-261 (1990). These proteins include blood coagulation factors (factors VI, IX, X, XII, protein C, protein S, protein Z, tissue plasminogen activator, urokinase), extracellular matrix components (laminin, cytotactin, entactin), cell surface receptors (LDL receptor, thrombomodulin receptor) and immunity-related proteins (complement C1r, uromodulin).

Even more interesting, the general structure pattern of EGF-like precursors is preserved through lower organisms as well as in mammalian cells. A number of genes with developmental significance have been identified in invertebrates with EGF-like repeats. For example, the *notch* gene of Drosophila encodes 36 tandemly arranged 40 amino acid repeats which show homology to EGF, Wharton *et al.*, *Cell* 43: 557-581 (1985). Hydropathy plots indicate a putative membrane spanning domain, with the EGF-related sequences being located on the extracellular side of the membrane. Other homeotic genes with EGF-like repeats include Delta, 95F and 5ZD which were identified using probes based on Notch, and the nematode gene *Lin-12* which encodes a putative receptor for a developmental signal transmitted between two specified cells.

Specifically, EGF has been shown to have potential in the preservation and maintenance of gastrointestinal mucosa and the repair of acute and chronic mucosal lesions, Konturek *et al.*, *Eur. J. Gastroenterol Hepatol.* 7 (10), 933-37 (1995), including the treatment of necrotizing enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration gastrointestinal ulcerations and congenital microvillus atrophy, Guglietta and Sullivan, *Eur. J. Gastroenterol Hepatol*, 7(10), 945-50 (1995). Additionally, EGF has been implicated in hair follicle differentiation; du Cros, *J. Invest. Dermatol.* 101 (1 Suppl.), 106S-113S (1993), Hillier, *Clin. Endocrinol.* 33(4), 427-28 (1990); kidney function, Hamm *et al.*, *Semin. Nephrol.* 13 (1): 109-15 (1993), Harris, *Am. J. Kidney Dis.* 17(6): 627-30 (1991); tear fluid, van Setten *et al.*, *Int. Ophthalmol* 15(6); 359-62 (1991); vitamin K mediated blood coagulation, Stenflo *et al.*, *Blood* 78(7): 1637-51 (1991). EGF is also implicated various skin disease characterized by abnormal keratinocyte differentiation, e.g., psoriasis, epithelial cancers such as squamous cell carcinomas of the lung, epidermoid carcinoma of the vulva and gliomas. King *et al.*, *Am. J. Med. Sci.* 296: 154-158 (1988).

Of great interest is mounting evidence that genetic alterations in growth factors signaling pathways are closely linked to developmental abnormalities and to chronic diseases including cancer. Aaronson, *Science* 254: 1146-1153 (1991). For example, c-erb-2 (also known as HER-2), a proto-oncogene with close structural similarity to EGF receptor protein, is overexpressed in human breast cancer. King *et al.*, *Science* 229: 974-976 (1985); Gullick, *Hormones and their actions*, Cooke *et al.*, eds, Amsterdam, Elsevier, pp 349-360 (1986).

We herein describe the identification and characterization of novel polypeptides having homology to EGF, wherein those polypeptides are herein designated PRO211 and PRO217.

2. PRO230

Nephritis is a condition characterized by inflammation of the kidney affecting the structure and normal function of the kidney. This condition can be chronic or acute and is generally caused by infection, degenerative process or vascular disease. In all cases, early detection is desirable so that the patient with nephritis can begin treatment of the condition.

An approach to detecting nephritis is to determine the antigens associated with nephritis and antibodies thereto. In rabbit, a tubulointerstitial nephritis antigen (TIN-ag) has been reported in Nelson, T. R., et al., J. Biol. Chem., 270(27):16265-70 (July 1995) (GENBANK/U24270). This study reports that the rabbit TIN-ag is a basement membrane glycoprotein having a predicted amino acid sequence which has a carboxyl-terminal region exhibiting 30% homology with human preprocathepsin B, a member of the cystein proteinase family of proteins. It is also reported that the rabbit TIN-ag has a domain in the amino-terminal region containing an epidermal growth factor-like motif that shares homology with laminin A and S chains, alpha 1 chain of type I collagen, von Willebrand's factor and mucin, indicating structural and functional similarities. Studies have also been conducted in mice. However, it is desirable to identify tubulointerstitial nephritis antigens in humans to aid in the development of early detection methods and treatment of nephritis.

Proteins which have homology to tubulointerstitial nephritis antigens are of particular interest to the medical and industrial communities. Often, proteins having homology to each other have similar function. It is also of interest when proteins having homology do not have similar functions, indicating that certain structural motifs identify information other than function, such as locality of function. We herein describe the identification and characterization of a novel polypeptide, designated hgerein as PRO230, which has homology to tubulointerstitial nephritis antigens.

3. PRO232

Stem cells are undifferentiated cells capable of (a) proliferation, (b) self maintenance, (c) the production of a large number of differentiated functional progeny, (d) regeneration of tissue after injury and/or (e) a flexibility in the use of these options. Stem cells often express cell surface antigens which are capable of serving as cell specific markers that can be exploited to identify stem cells, thereby providing a means for identifying and isolating specific stem cell populations.

Having possession of different stem cell populations will allow for a number of important applications. For example, possessing a specific stem cell population will allow for the identification of growth factors and other proteins which are involved in their proliferation and differentiation. In addition, there may be as yet undiscovered proteins which are associated with (1) the early steps of dedication of the stem cell to a particular lineage, (2) prevention of such dedication, and (3) negative control of stem cell proliferation, all of which may be identified if one has possession of the stem cell population. Moreover, stem cells are important and ideal targets for gene therapy where the inserted genes promote the health of the individual into whom the stem cells are transplanted. Finally, stem cells may play important roles in transplantation of organs or tissues, for example liver regeneration and skin grafting.

Given the importance of stem cells in various different applications, efforts are currently being undertaken by both industry and academia to identify new, native stem cell antigen proteins so as to provide

specific cell surface markers for identifying stem cell populations as well as for providing insight into the functional roles played by stem cell antigens in cell proliferation and differentiation. We herein describe the identification and characterization of novel polypeptides having homology to a stem cell antigen, wherein those polypeptides are herein designated as PRO232 polypeptides..

4. PRO187

Growth factors are molecular signals or mediators that enhance cell growth or proliferation, alone or in concert, by binding to specific cell surface receptors. However, there are other cellular reactions than only growth upon expression to growth factors. As a result, growth factors are better characterized as multifunctional and potent cellular regulators. Their biological effects include proliferation, chemotaxis and stimulation of extracellular matrix production. Growth factors can have both stimulatory and inhibitory effects. For example, transforming growth factor (TGF-β) is highly pleiotropic and can stimulate proliferation in some cells, especially connective tissue, while being a potent inhibitor of proliferation in others, such as lymphocytes and epithelial cells.

The physiological effect of growth stimulation or inhibition by growth factors depends upon the state of development and differentiation of the target tissue. The mechanism of local cellular regulation by classical endocrine molecules involves comprehends autocrine (same cell), juxtacrine (neighbor cell), and paracrine (adjacent cells) pathways. Peptide growth factors are elements of a complex biological language, providing the basis for intercellular communication. They permit cells to convey information between each other, mediate interaction between cells and change gene expression. The effect of these multifunctional and pluripotent factors is dependent on the presence or absence of other peptides.

FGF-8 is a member of the fibroblast growth factors (FGFs) which are a family of heparin-binding, potent mitogens for both normal diploid fibroblasts and established cell lines, Gospodarowicz *et al.* (1984), *Proc. Natl. Acad. Sci. USA* 81:6963. The FGF family comprises acidic FGF (FGF-1), basic FGF (FGF-2), INT-2 (FGF-3), K-FGF/HST (FGF-4), FGF-5, FGF-6, KGF (FGF-7), AIGF (FGF-8) among others. All FGFs have two conserved cysteine residues and share 30-50% sequence homology at the amino acid level. These factors are mitogenic for a wide variety of normal diploid mesoderm-derived and neural crest-derived cells, including granulosa cells, adrenal cortical cells, chondrocytes, myoblasts, corneal and vascular endothelial cells (bovine or human), vascular smooth muscle cells, lens, retina and prostatic epithelial cells, oligodendrocytes, astrocytes, chrondocytes, myoblasts and osteoblasts.

Fibroblast growth factors can also stimulate a large number of cell types in a non-mitogenic manner. These activities include promotion of cell migration into wound area (chemotaxis), initiation of new blood vessel formulation (angiogenesis), modulation of nerve regeneration and survival (neurotrophism), modulation of endocrine functions, and stimulation or suppression of specific cellular protein expression, extracellular matrix production and cell survival. Baird & Bohlen, Handbook of Exp. Pharmacol. 95(1): 369-418, Springer, (1990). These properties provide a basis for using fibroblast growth factors in therapeutic approaches to accelerate wound healing, nerve repair, collateral blood vessel formation, and the like. For example, fibroblast growth factors have been suggested to minimize myocardium damage in heart disease and surgery (U.S.P. 4,378,347).

FGF-8, also known as androgen-induced growth factor (AIGF), is a 215 amino acid protein which

shares 30-40% sequence homology with the other members of the FGF family. FGF-8 has been proposed to be under androgenic regulation and induction in the mouse mammary carcinoma cell line SC3. Tanaka *et al., Proc. Natl. Acad. Sci. USA* 89: 8928-8932 (1992); Sato *et al., J. Steroid Biochem. Molec. Biol.* 47: 91-98 (1993). As a result, FGF-8 may have a local role in the prostate, which is known to be an androgen-responsive organ. FGF-8 can also be oncogenic, as it displays transforming activity when transfected into NIH-3T3 fibroblasts. Kouhara *et al., Oncogene* 9 455-462 (1994). While FGF-8 has been detected in heart, brain, lung, kidney, testis, prostate and ovary, expression was also detected in the absence of exogenous androgens. Schmitt *et al., J. Steroid Biochem. Mol. Biol.* 57 (3-4): 173-78 (1996).

FGF-8 shares the property with several other FGFs of being expressed at a variety of stages of murine embryogenesis, which supports the theory that the various FGFs have multiple and perhaps coordinated roles in differentiation and embryogenesis. Moreover, FGF-8 has also been identified as a protooncogene that cooperates with Wnt-1 in the process of mammary tumorigenesis (Shackleford *et al., Proc. Natl. Acad. Sci. USA* 90, 740-744 (1993); Heikinheimo *et al., Mech. Dev.* 48: 129-138 (1994)).

In contrast to the other FGFs, FGF-8 exists as three protein isoforms, as a result of alternative splicing of the primary transcript. Tanaka et al., *supra*. Normal adult expression of FGF-8 is weak and confined to gonadal tissue, however northern blot analysis has indicated that FGF-8 mRNA is present from day 10 through day 12 or murine gestation, which suggests that FGF-8 is important to normal development. Heikinheimo *et al., Mech Dev.* 48(2): 129-38 (1994). Further *in situ* hybridization assays between day 8 and 16 of gestation indicated initial expression in the surface ectoderm of the first bronchial arches, the frontonasal process, the forebrain and the midbrain-hindbrain junction. At days 10-12, FGF-8 was expressed in the surface ectoderm of the forelimb and hindlimb buds, the nasal its and nasopharynx, the infundibulum and in the telencephalon, diencephalon and metencephalon. Expression continues in the developing hindlimbs through day 13 of gestation, but is undetectable thereafter. The results suggest that FGF-8 has a unique temporal and spatial pattern in embryogenesis and suggests a role for this growth factor in multiple regions of ectodermal differentiation in the post-gastrulation embryo.

We herein describe the identification of novel poypeptides having homology to FGF-8, wherein those polypeptides are heein designated PRO187 polypeptides.

5. PRO265

Protein-protein interactions include receptor and antigen complexes and signaling mechanisms. As more is known about the structural and functional mechanisms underlying protein-protein interactions, protein-protein interactions can be more easily manipulated to regulate the particular result of the protein-protein interaction. Thus, the underlying mechanisms of protein-protein interactions are of interest to the scientific and medical community.

All proteins containing leucine-rich repeats are thought to be involved in protein-protein interactions. Leucine-rich repeats are short sequence motifs present in a number of proteins with diverse functions and cellular locations. The crystal structure of ribonuclease inhibitor protein has revealed that leucine-rich repeats correspond to beta-alpha structural units. These units are arranged so that they form a parallel beta-sheet with one surface exposed to solvent, so that the protein acquires an unusual, nonglubular shape. These two features

have been indicated as responsible for the protein-binding functions of proteins containing leucine-rich repeats. See, Kobe and Deisenhofer, Trends Biochem. Sci., 19(10):415-421 (Oct. 1994).

A study has been reported on leucine-rich proteoglycans which serve as tissue organizers, orienting and ordering collagen fibrils during ontogeny and are involved in pathological processes such as wound healing, tissue repair, and tumor stroma formation. Iozzo, R. V., Crit. Rev. Biochem. Mol. Biol., 32(2):141-174 (1997). Others studies implicating leucine rich proteins in wound healing and tissue repair are De La Salle, C., et al., Vouv. Rev. Fr. Hematol. (Germany), 37(4):215-222 (1995), reporting mutations in the leucine rich motif in a complex associated with the bleeding disorder Bernard-Soulier syndrome and Chlemetson, K. J., Thromb. Haemost. (Germany), 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats. Another protein of particular interest which has been reported to have leucine-rich repeats is the SLIT protein which has been reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanistsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Other studies reporting on the biological functions of proteins having leucine-rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996) (gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996) (apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement); and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation (decorin binding to transforming growth factor-β involvement for treatment for cancer, wound healing and scarring). Also of particular interest is fibromodulin and its use to prevent or reduce dermal scarring. A study of fibromodulin is found in U.S. Patent No. 5,654,270 to Ruoslahti, et al.

Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions. Of particular interest are those proteins having leucine rich repeats and homology to known proteins having leucine rich repeats such as fibromodulin, the SLIT protein and platelet glycoprotein V. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound proteins having leucine rich repeats. We herein describe the identification and characterization of novel polypeptides having homology to fibromodulin, herein designated as PRO265 polypeptides.

6. PRO219

Human matrilin-2 polypeptide is a member of the von Willebrand factor type A-like module superfamily. von Willebrand factor is a protein which plays an important role in the maintenence of hemostasis. More specifically, von Willebrand factor is a protein which is known to participate in platelet-vessel wall interactions at the site of vascular injury via its ability to interact and form a complex with Factor VIII. The absence of von Willebrand factor in the blood causes an abnormality with the blood platelets that prevents platelet adhesion to the vascular wall at the site of the vascular injury. The result is the propensity for brusing, nose bleeds, intestinal bleeding, and the like comprising von Willebrand's disease.

Given the physiological importance of the blood clotting factors, efforts are currently being undertaken by both industry and academia to identify new, native proteins which may be involved in the coagulation process. We herein describe the identification of a novel full-length polypeptide which possesses homology to the human

matrilin-2 precursor polypeptide.

7. PRO246

The cell surface protein HCAR is a membrane-bound protein that acts as a receptor for subgroup C of the adenoviruses and subgroup B of the coxsackieviruses. Thus, HCAR may provide a means for mediating viral infection of cells in that the presence of the HCAR receptor on the cellular surface provides a binding site for viral particles, thereby facilitating viral infection.

In light of the physiological importance of membrane-bound proteins and specficially those which serve a cell surface receptor for viruses, efforts are currently being undertaken by both industry and academia to identify new, native membrane-bound receptor proteins. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel receptor proteins. We herein describe a novel membrane-bound polypeptide (designated herein as PRO246) having homology to the cell surface protein HCAR and to various tumor antigens including A33 and carcinoembryonic antigen, wherein this polypeptide may be a novel cell surface virus receptor or tumor antigen.

8. PRO228

There are a number of known seven transmembrane proteins and within this family is a group which includes CD97 and EMR1. CD97 is a seven-span transmembrane receptor which has a cellular ligand, CD55, DAF. Hamann, et al., J. Exp. Med. (U.S.), 184(3):1189 (1996). Additionally, CD97 has been reported as being a dedifferentiation marker in human thyroid carcinomas and as associated with inflammation. Aust, et al., Cancer Res. (U.S.), 57(9):1798 (1997); Gray, et al., J. Immunol. (U.S.), 157(12):5438 (1996). CD97 has also been reported as being related to the secretin receptor superfamily, but unlike known members of that family, CD97 and EMR1 have extended extracellular regions that possess several EGF domains at the N-terminus. Hamann, et al., Genomics, 32(1):144 (1996); Harmann, et al., J. Immunol., 155(4):1942 (1995). EMR1 is further described in Lin, et al., Genomics, 41(3):301 (1997) and Baud, et al., Genomics, 26(2):334 (1995). While CD97 and EMR1 appear to be related to the secretin receptors, a known member of the secretin family of G protein-coupled receptors includes the alpha-latroxin receptor, latrophilin, which has been described as calcium independent and abundant among neuronal tissues. Lelianova, et al., J. Biol. Chem., 272(34), 21504 (1997); Davletov, et al., J. Biol. Chem. (U.S.), 271(38):23239 (1996). Both members of the secretin receptor superfamily and non-members which are related to the secretin receptor superfamily, or CRF and calcitonin receptors are of interest. In particular, new members of these families, identified by their homology to known proteins, are of interest.

Efforts are being undertaken by both industry and academia to identify new membrane-bound receptor proteins, particularly transmembrane proteins with EGF repeats and large N-terminuses which may belong to the family of seven-transmembrane proteins of which CD97 and EMR1 are members. We herein describe the identification and charactization of novel polypeptides having homology to CD97 and EMR1, designated herein as PRO228 polypeptides.

9. PRO533

Growth factors are molecular signals or mediators that enhance cell growth or proliferation, alone or in concert, by binding to specific cell surface receptors. however, there are other cellular reactions than only growth upon expression to growth factors. As a result, growth factors are better characterized as multifunctional and potent cellular regulators. Their biological effects include proliferation, chemotaxis and stimulation of extracellular matrix production. Growth factors can have both stimulatory and inhibitory effects. For example, transforming growth factors (TGF-β) is highly pleiotropic and can stimulate proliferation in some cells, especially connective tissues, while being a potent inhibitor of proliferation in others, such as lymphocytes and epithelial cells.

The physiological effect of growth stimulation or inhibition by growth factors depends upon the state of development and differentiation of the target tissue. The mechanism of local cellular regulation by classical endocrine molecules comprehends autocrine (same cell), juxtacrine (neighbor cell), and paracrine (adjacent cell) pathways. Peptide growth factors are elements of a complex biological language, providing the basis for intercellular communication. They permit cells to convey information between each other, mediate interaction between cells and change gene expression. the effect of these multifunctional and pluripotent factors is dependent on the presence or absence of other peptides.

Fibroblast growth factors (FGFs) are a family of heparin-binding, potent mitogens for both normal diploid fibroblasts and established cell lines, Godpodarowicz, D. et al. (1984), Proc. Natl. Acad. Sci. USA 81: 6983. the FGF family comprises acidic FGF (FGF-1), basic FGF (FGF-2), INT-2 (FGF-3), K-FGF/HST (FGF-4), FGF-5, FGF-6, KGF (FGF-7), AIGF (FGF-8) among others. All FGFs have two conserved cysteine residues and share 30-50% sequence homology at the amino acid level. These factors are mitogenic for a wide variety of normal diploid mesoderm-derived and neural crest-derived cells, inducing granulosa cells, adrenal cortical cells, chrondocytes, myoblasts, corneal and vascular endothelial cells (bovine or human), vascular smooth muscle cells, lens, retina and prostatic epithelial cells, oligodendrocytes, astrocytes, chrondocytes, myoblasts and osteoblasts.

Fibroblast growth factors can also stimulate a large number of cell types in a non-mitogenic manner. These activities include promotion of cell migration into a wound area (chemotaxis), initiation of new blood vessel formulation (angiogenesis), modulation of nerve regeneration and survival (neurotrophism), modulation of endocrine functions, and stimulation or suppression of specific cellular protein expression, extracellular matrix production and cell survival. Baird, A. & Bohlen, P., *Handbook of Exp. Phrmacol.* 95(1): 369-418 (1990). These properties provide a basis for using fibroblast growth factors in therapeutic approaches to accelerate wound healing, nerve repair, collateral blood vessel formation, and the like. For example, fibroblast growth factors, have been suggested to minimize myocardium damage in heart disease and surgery (U.S.P. 4,378,437).

We herein describe the identification and characterization of novel polypeptides having homology to FGF, herein designated PRO533 polypeptides.

10. PRO245

Some of the most important proteins involved in the above described regulation and modulation of cellular processes are the enzymes which regulate levels of protein phosphorylation in the cell. For example, it is known that the transduction of signals that regulate cell growth and differentiation is regulated at least in

part by phosphorylation and dephosphorylation of various cellular proteins. The enzymes that catalyze these processes include the protein kinases, which function to phosphorylate various cellular proteins, and the protein phosphatases, which function to remove phosphate residues from various cellular proteins. The balance of the level of protein phosphorylation in the cell is thus mediated by the relative activities of these two types of enzymes.

Although many protein kinase enzymes have been identified, the physiological role played by many of these catalytic proteins has yet to be elucidated. It is well known, however, that a number of the known protein kinases function to phosphorylate tyrosine residues in proteins, thereby leading to a variety of different effects. Perhaps most importantly, there has been a great deal of interest in the protein tyrosine kinases since the discovery that many oncogene products and growth factors possess intrinsic protein tyrosine kinase activity. There is, therefore, a desire to identify new members of the protein tyrosine kinase family.

Given the physiological importance of the protein kinases, efforts are being undertaken by both industry and academia to identify new, native kinase proteins. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel kinase proteins. We herein describe the identification and characterization of novel polypeptides having homology to tyrosine kinase proteins, designated herein as PRO245 polypeptides.

11. PRO220, PRO221 and PRO227

Protein-protein interactions include receptor and antigen complexes and signaling mechanisms. As more is known about the structural and functional mechanisms underlying protein-protein interactions, protein-protein interactions can be more easily manipulated to regulate the particular result of the protein-protein interaction. Thus, the underlying mechanisms of protein-protein interactions are of interest to the scientific and medical community.

All proteins containing leucine-rich repeats are thought to be involved in protein-protein interactions. Leucine-rich repeats are short sequence motifs present in a number of proteins with diverse functions and cellular locations. The crystal structure of ribonuclease inhibitor protein has revealed that leucine-rich repeats correspond to beta-alpha structural units. These units are arranged so that they form a parallel beta-sheet with one surface exposed to solvent, so that the protein acquires an unusual, nonglubular shape. These two features have been indicated as responsible for the protein-binding functions of proteins containing leucine-rich repeats. See, Kobe and Deisenhofer, Trends Biochem. Sci., 19(10):415-421 (Oct. 1994).

A study has been reported on leucine-rich proteoglycans which serve as tissue organizers, orienting and ordering collagen fibrils during ontogeny and are involved in pathological processes such as wound healing, tissue repair, and tumor stroma formation. Iozzo, R. V., Crit. Rev. Biochem. Mol. Biol., 32(2):141-174 (1997). Others studies implicating leucine rich proteins in wound healing and tissue repair are De La Salle, C., et al., Vouv. Rev. Fr. Hematol. (Germany), 37(4):215-222 (1995), reporting mutations in the leucine rich motif in a complex associated with the bleeding disorder Bernard-Soulier syndrome and Chlemetson, K. J., Thromb. Haemost. (Germany), 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats. Another protein of particular interest which has been reported to have leucine-rich repeats is the SLIT protein which has been reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage

such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanistsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Other studies reporting on the biological functions of proteins having leucine-rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996) (gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996) (apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement); and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation (decorin binding to transforming growth factorβ involvement for treatment for cancer, wound healing and scarring).

Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions. Of particular interest are those proteins having leucine rich repeats and homology to known proteins having leucine rich repeats such as the SLIT protein and platelet glycoprotein V.

12. PRO258

Immunoglobulins are antibody molecules, the proteins that function both as receptors for antigen on the B-cell membrane and as the secreted products of the plasma cell. Like all antibody molecules, immunoglobulins perform two major functions: they bind specifically to an antigen and they participate in a limited number of biological effector functions. Therefore, new members of the Ig superfamily are always of interest. Molecules which act as receptors by various viruses and those which act to regulate immune function are of particular interest. Also of particular interest are those molecules which have homology to known Ig family members which act as virus receptors or regulate immune function. Thus, molecules having homology to poliovirus receptors, CRTAM and CD166 (a ligand for lymphocyte antigen CD6) are of particular interest.

Extracellular and membrane-bound proteins play important roles in the formation, differentiation and maintenance of multicellular organisms. The fate of many individual cells, e.g., proliferation, migration, differentiation, or interaction with other cells, is typically governed by information received from other cells and/or the immediate environment. This information is often transmitted by secreted polypeptides (for instance, mitogenic factors, survival factors, cytotoxic factors, differentiation factors, neuropeptides, and hormones) which are, in turn, received and interpreted by diverse cell receptors or membrane-bound proteins. These secreted polypeptides or signaling molecules normally pass through the cellular secretory pathway to reach their site of action in the extracellular environment, usually at a membrane-bound receptor protein.

We herein describe the identification and characterization of novel polypeptides having homology to CRTAM, designated herein as PRO258 polypeptides.

13. PRO266

Protein-protein interactions include receptor and antigen complexes and signaling mechanisms. As more is known about the structural and functional mechanisms underlying protein-protein interactions, protein-protein interactions can be more easily manipulated to regulate the particular result of the protein-protein interaction. Thus, the underlying mechanisms of protein-protein interactions are of interest to the scientific and medical community.

All proteins containing leucine-rich repeats are thought to be involved in protein-protein interactions. Leucine-rich repeats are short sequence motifs present in a number of proteins with diverse functions and cellular locations. The crystal structure of ribonuclease inhibitor protein has revealed that leucine-rich repeats correspond to beta-alpha structural units. These units are arranged so that they form a parallel beta-sheet with one surface exposed to solvent, so that the protein acquires an unusual, nonglobular shape. These two features have been indicated as responsible for the protein-binding functions of proteins containing leucine-rich repeats. See, Kobe and Deisenhofer, Trends Biochem. Sci., 19(10):415-421 (Oct. 1994).

A study has been reported on leucine-rich proteoglycans which serve as tissue organizers, orienting and ordering collagen fibrils during ontogeny and are involved in pathological processes such as wound healing, tissue repair, and tumor stroma formation. Iozzo, R. V., Crit. Rev. Biochem. Mol. Biol., 32(2):141-174 (1997). Others studies implicating leucine rich proteins in wound healing and tissue repair are De La Salle, C., et al., Vouv. Rev. Fr. Hematol. (Germany), 37(4):215-222 (1995), reporting mutations in the leucine rich motif in a complex associated with the bleeding disorder Bernard-Soulier syndrome and Chlemetson, K. J., Thromb. Haemost. (Germany), 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats. Another protein of particular interest which has been reported to have leucine-rich repeats is the SLIT protein which has been reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanistsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Other studies reporting on the biological functions of proteins having leucine-rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996) (gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996) (apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement); and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation (decorin binding to transforming growth factorβ involvement for treatment for cancer, wound healing and scarring).

Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions, neuronal development and adhesin molecules. Of particular interest are those proteins having leucine rich repeats and homology to known proteins having leucine rich repeats such as the SLIT protein. We herein describe novel polypeptides having homology to SLIT, designated herein as PRO266 polypeptides.

14. PRO269

Thrombomodulin binds to and regulates the activity of thrombin. It is important in the control of blood coagulation. Thrombomodulin functions as a natural anticoagulant by accelerating the activation of protein C by thrombin. Soluble thrombomodulin may have therapeutic use as an antithrombotic agent with reduced risk for hemorrhage as compared with heparin. Thrombomodulin is a cell surface trans-membrane glycoprotein, present on endothelial cells and platelets. A smaller, functionally active form of thrombomodulin circulates in the plasma and is also found in urine. (*In* Haeberli, A., Human Protein Data, VCH Oub., N.Y., 1992). Peptides having homology to thrombomodulin are particularly desirable.

We herein describe the identification and characterization of novel polypeptides having homology to

thrombomodulin, designated herein as PRO269 polypeptides.

15. **PRO287**

Procollagen C-proteinase enhancer protein binds to and enhances the activity of bone morphogenic protein "BMP1"/procollagen C-proteinase (PCP). It plays a role in extracellular matrix deposition. BMP1 proteins may be used to induce bone and/or cartilage formation and in wound healing and tissue repair. Therefore, procollagen C-proteinase enhancer protein, BMP1 and proteins having homology thereto, are of interest to the scientific and medical communities.

We herein describe the identification and characterization of novel polypeptides having homology to procollagen C-proteinase enhancer protein precursor and procollagen C-proteinase enhancer protein, designated herein as PRO287 polypeptides.

16. **PRO214**

Growth factors are molecular signals or mediators that enhances cell growth or proliferation, alone or in concert, by binding to specific cell surface receptors. However, there are other cellular reactions than only growth upon expression to growth factors. As a result, growth factors are better characterized as multifunctional and potent cellular regulators. Their biological effects include proliferation, chemotaxis and stimulation of extracellular matrix production. Growth factors can have both stimulatory and inhibitory effects. For example, transforming growth factor β (TGF-β) is highly pleiotropic and can stimulate proliferation in some cells, especially connective tissue, while being a potent inhibitor of proliferation in others, such as lymphocytes and epithelial cells.

The physiological effect of growth stimulation or inhibition by growth factors depends upon the state of development and differentiation of the target tissue. The mechanism of local cellular regulation by classical endocrine molecules involves comprehends autocrine (same cell), juxtacrine (neighbor cell), and paracrine (adjacent cells) pathways. Peptide growth factors are elements of a complex biological language, providing the basis for intercellular communication. They permit cells to convey information between each other, mediate interaction between cells and change gene expression. The effect of these multifunctional and pluripotent factors is dependent on the presence or absence of other peptides.

Epidermal growth factor (EGF) is a conventional mitogenic factor that stimulates the proliferation of various types of cells including epithelial cells and fibroblasts. EGF binds to and activates the EGF receptor (EGFR), which initiates intracellular signaling and subsequent effects. The EGFR is expressed in neurons of the cerebral cortex, cerebellum, and hippocampus in addition to other regions of the central nervous system (CNS). In addition, EGF is also expressed in various regions of the CNS. Therefore, EGF acts not only on mitotic cells, but also on postmitotic neurons. In fact, many studies have indicated that EGF has neurotrophic or neuromodulatory effects on various types of neurons in the CNS. For example, EGF acts directly on cultured cerebral cortical and cerebellar neurons, enhancing neurite outgrowth and survival. On the other hand, EGF also acts on other cell types, including septal cholinergic and mesencephalic dopaminergic neurons, indirectly through glial cells. Evidence of the effects of EGF on neurons in the CNS is accumulating, but the mechanisms of action remain essentially unknown. EGF-induced signaling in mitotic cells is better understood than in

postmitotic neurons. Studies of cloned pheochromocytoma PC12 cells and cultured cerebral cortical neurons have suggested that the EGF-induced neurotrophic actions are mediated by sustained activation of the EGFR and mitogen-activated protein kinase (MAPK) in response to EGF. The sustained intracellular signaling correlates with the decreased rate of EGFR down-regulation, which might determine the response of neuronal cells to EGF. It is likely that EGF is a multi-potent growth factor that acts upon various types of cells including mitotic cells and postmitotic neurons.

EGF is produced by the salivary and Brunner's glands of the gastrointestinal system, kidney, pancreas, thyroid gland, pituitary gland, and the nervous system, and is found in body fluids such as saliva, blood, cerebrospinal fluid (CSF), urine, amniotic fluid, prostatic fluid, pancreatic juice, and breast milk, Plata-Salaman, CR *Peptides* 12: 653-663 (1991).

EGF is mediated by its membrane specific receptor, which contains an intrinsic tyrosine kinase. Stoscheck CM *et al., J. Cell Biochem.* 31: 135-152 (1986). EGF is believed to function by binding to the extracellular portion of its receptor which induces a transmembrane signal that activates the intrinsic tyrosine kinase.

Purification and sequence analysis of the EGF-like domain has revealed the presence of six conserved cysteine residues which cross-bind to create three peptide loops, Savage *CR et al., J. Biol. Chem.* 248: 7669-7672 (1979). It is now generally known that several other peptides can react with the EGF receptor which share the same generalized motif $X_aCX_7CX_{4/5}CX_{10}CXCX_5GX_2CX_n$, where X represents any non-cysteine amino acid, and n is a variable repeat number. Non isolated peptides having this motif include TGF-a, amphiregulin, schwannoma-derived growth factor (SDGF), heparin-binding EGF-like growth factors and certain virally encoded peptides (e.g., Vaccinia virus, Reisner AH, *Nature* 313: 801-803 (1985), Shope fibroma virus, Chang W., et al., Mol Cell Biol. 7: 535-540 (1987), Molluscum contagiosum, Porter CD & Archard LC, *J. Gen. Virol.* 68: 673-682 (1987), and Myxoma virus, Upton C *et al., J. Virol.* 61: 1271-1275 (1987). Prigent SA & Lemoine N.R., *Prog. Growth Factor Res.* 4: 1-24 (1992).

EGF-like domains are not confined to growth factors but have been observed in a variety of cell-surface and extracellular proteins which have interesting properties in cell adhesion, protein-protein interaction and development, Laurence DJR & Gusterson BA, *Tumor Biol.* 11: 229-261 (1990). These proteins include blood coagulation factors (factors VI, IX, X, XII, protein C, protein S, protein Z, tissue plasminogen activator, urokinase), extracellular matrix components (laminin, cytotactin, entactin), cell surface receptors (LDL receptor, thrombomodulin receptor) and immunity-related proteins (complement C1r, uromodulin).

Even more interesting, the general structure pattern of EGF-like precursors is preserved through lower organisms as well as in mammalian cells. A number of genes with developmental significance have been identified in invertebrates with EGF-like repeats. For example, the *notch* gene of Drosophila encodes 36 tandemly arranged 40 amino acid repeats which show homology to EGF, Wharton W *et al., Cell* 43: 557-581 (1985). Hydropathy plots indicate a putative membrane spanning domain, with the EGF-related sequences being located on the extracellular side of the membrane. Other homeotic genes with EGF-like repeats include Delta, 95F and 5ZD which were identified using probes based on Notch, and the nematode gene *Lin-12* which encodes a putative receptor for a developmental signal transmitted between two specified cells.

Specifically, EGF has been shown to have potential in the preservation and maintenance of gastrointestinal mucosa and the repair of acute and chronic mucosal lesions, Konturek, *PC et al., Eur. J. Gastroenterol Hepatol.* 7 (10), 933-37 (1995), including the treatment of necrotizing enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration gastrointestinal ulcerations and congenital microvillus atrophy, A. Guglietta & PB Sullivan, *Eur. J. Gastroenterol Hepatol*, 7(10), 945-50 (1995). Additionally, EGF has been implicated in hair follicle differentiation; C.L. du Cros, *J. Invest. Dermatol.* 101 (1 Suppl.), 106S-113S (1993), SG Hillier, *Clin. Endocrinol.* 33(4), 427-28 (1990); kidney function, L.L. Hamm *et al., Semin. Nephrol.* 13 (1): 109-15 (1993), RC Harris, *Am. J. Kidney Dis.* 17(6): 627-30 (1991); tear fluid, GB van Setten *et al., Int. Ophthalmol* 15(6); 359-62 (1991); vitamin K mediated blood coagulation, J. Stenflo *et al., Blood* 78(7): 1637-51 (1991). EGF is also implicated various skin disease characterized by abnormal keratinocyte differentiation, e.g., psoriasis, epithelial cancers such as squamous cell carcinomas of the lung, epidermoid carcinoma of the vulva and gliomas. King, LE *et al., Am. J. Med. Sci.* 296: 154-158 (1988).

Of great interest is mounting evidence that genetic alterations in growth factors signaling pathways are closely linked to developmental abnormalities and to chronic diseases including cancer. Aaronson SA, *Science* 254: 1146-1153 (1991). For example, c-erb-2 (also known as HER-2), a proto-oncogene with close structural similarity to EGF receptor protein, is overexpressed in human breast cancer. King *et al.*, *Science* 229: 974-976 (1985); Gullick, WJ, *Hormones and their actions*, Cooke BA *et al.*, eds, Amsterdam, Elsevier, pp 349-360 (1986).

17. PRO317

The TGF-β supergene family, or simply TGF-β superfamily, a group of secreted proteins, includes a large number of related growth and differentiation factors expressed in virtually all phyla. Superfamily members bind to specific cell surface receptors that activate signal transduction mechanisms to elicit their multifunctional cytokine effects. Kolodziejczyk and Hall, Biochem. Cell. Biol., 74: 299-314 (1996); Attisano and Wrana, Cytokine Growth Factor Rev., 7: 327-339 (1996); and Hill, Cellular Signaling, 8: 533-544 (1996).

Members of this family include five distinct forms of TGF-β (Sporn and Roberts, in Peptide Growth Factors and Their Receptors, Sporn and Roberts, eds. (Springer-Verlag: Berlin, 1990) pp. 419-472), as well as the differentiation factors vg1 (Weeks and Melton, Cell, 51: 861-867 (1987)) and DPP-C polypeptide (Padgett *et al.*, Nature, 325: 81-84 (1987)), the hormones activin and inhibin (Mason *et al.*, Nature, 318: 659-663 (1985); Mason *et al.*, Growth Factors, 1: 77-88 (1987)), the Mullerian-inhibiting substance (MIS) (Cate *et al.*, Cell, 45: 685-698 (1986)), the bone morphogenetic proteins (BMPs) (Wozney *et al.*, Science, 242: 1528-1534 (1988); PCT WO 88/00205 published January 14, 1988; U.S. 4,877,864 issued October 31, 1989), the developmentally regulated proteins Vgr-1 (Lyons *et al.*, Proc. Natl. Acad. Sci. USA. 86: 4554-4558 (1989)) and Vgr-2 (Jones *et al.*, Molec. Endocrinol., 6: 1961-1968 (1992)), the mouse growth differentiation factor (GDF), such as GDF-3 and GDF-9 (Kingsley, Genes Dev., 8: 133-146 (1994); McPherron and Lee, J. Biol. Chem., 268: 3444-3449 (1993)), the mouse lefty/Stra1 (Meno *et al.*, Nature, 381: 151-155 (1996); Bouillet *et al.*, Dev. Biol., 170: 420-433 (1995)), glial cell line-derived neurotrophic factor (GDNF) (Lin *et al.*, Science, 260: 1130-1132 (1993), neurturin (Kotzbauer *et al.*, Nature, 384: 467-470 (1996)), and endometrial bleeding-associated factor (EBAF) (Kothapalli *et al.*, J. Clin. Invest., 99: 2342-2350 (1997)). The subset BMP-2A and BMP-2B is approximately

75% homologous in sequence to DPP-C and may represent the mammalian equivalent of that protein.

The proteins of the TGF-β superfamily are disulfide-linked homo- or heterodimers encoded by larger precursor polypeptide chains containing a hydrophobic signal sequence, a long and relatively poorly conserved N-terminal pro region of several hundred amino acids, a cleavage site (usually polybasic), and a shorter and more highly conserved C-terminal region. This C-terminal region corresponds to the processed mature protein and contains approximately 100 amino acids with a characteristic cysteine motif, *i.e.*, the conservation of seven of the nine cysteine residues of TGF-β among all known family members. Although the position of the cleavage site between the mature and pro regions varies among the family members, the C-terminus of all of the proteins is in the identical position, ending in the sequence Cys-X-Cys-X, but differing in every case from the TGF-β consensus C-terminus of Cys-Lys-Cys-Ser. Sporn and Roberts, 1990, *supra*.

There are at least five forms of TGF-β currently identified, TGF-β1, TGF-β2, TGF-β3, TGF-β4, and TGF-β5. The activated form of TGF-β1 is a homodimer formed by dimerization of the carboxy-terminal 112 amino acids of a 390 amino acid precursor. Recombinant TGF-β1 has been cloned (Derynck *et al.*, Nature, 316:701-705 (1985)) and expressed in Chinese hamster ovary cells (Gentry *et al.*, Mol. Cell. Biol., 7: 3418-3427 (1987)). Additionally, recombinant human TGF-β2 (deMartin *et al.*, EMBO J., 6: 3673 (1987)), as well as human and porcine TGF-β3 (Derynck *et al.*, EMBO J., 7: 3737-3743 (1988); ten Dijke *et al.*, Proc. Natl. Acad. Sci. USA, 85: 4715 (1988)) have been cloned. TGF-β2 has a precursor form of 414 amino acids and is also processed to a homodimer from the carboxy-terminal 112 amino acids that shares approximately 70% homology with the active form of TGF-β1 (Marquardt *et al.*, J. Biol. Chem., 262: 12127 (1987)). See also EP 200,341; 169,016; 268,561; and 267,463; U.S. Pat. No. 4,774,322; Cheifetz *et al.*, Cell, 48: 409-415 (1987); Jakowlew *et al.*, Molecular Endocrin., 2: 747-755 (1988); Derynck *et al.*, J. Biol. Chem., 261: 4377-4379 (1986); Sharples *et al.*, DNA, 6: 239-244 (1987); Derynck *et al.*, Nucl. Acids. Res., 15: 3188-3189 (1987); Derynck *et al.*, Nucl. Acids. Res., 15: 3187 (1987); Seyedin *et al.*, J. Biol. Chem., 261: 5693-5695 (1986); Madisen *et al.*, DNA, 7: 1-8 (1988); and Hanks *et al.*, Proc. Natl. Acad. Sci. (U.S.A.), 85: 79-82 (1988).

TGF-β4 and TGF-β5 were cloned from a chicken chondrocyte cDNA library (Jakowlew *et al.*, Molec. Endocrinol., 2: 1186-1195 (1988)) and from a frog oocyte cDNA library, respectively.

The pro region of TGF-β associates non-covalently with the mature TGF-β dimer (Wakefield *et al.*, J. Biol. Chem., 263: 7646-7654 (1988); Wakefield *et al.*, Growth Factors, 1: 203-218 (1989)), and the pro regions are found to be necessary for proper folding and secretion of the active mature dimers of both TGF-β and activin (Gray and Mason, Science, 247: 1328-1330 (1990)). The association between the mature and pro regions of TGF-β masks the biological activity of the mature dimer, resulting in formation of an inactive latent form. Latency is not a constant of the TGF-β superfamily, since the presence of the pro region has no effect on activin or inhibin biological activity.

A unifying feature of the biology of the proteins from the TGF-β superfamily is their ability to regulate developmental processes. TGF-β has been shown to have numerous regulatory actions on a wide variety of both normal and neoplastic cells. TGF-β is multifunctional, as it can either stimulate or inhibit cell proliferation, differentiation, and other critical processes in cell function (Sporn and Roberts, *supra)*.

One member of the TGF-β superfamily, EBAF, is expressed in endometrium only in the late secretory phase and during abnormal endometrial bleeding. Kothapalli *et al.*, J. Clin. Invest., 99: 2342-2350 (1997).

Human endometrium is unique in that it is the only tissue in the body that bleeds at regular intervals. In addition, abnormal endometrial bleeding is one of the most common manifestations of gynecological diseases, and is a prime indication for hysterectomy. *In situ* hybridization showed that the mRNA of EBAF was expressed in the stroma without any significant mRNA expression in the endometrial glands or endothelial cells.

The predicted protein sequence of EBAF showed a strong homology to the protein encoded by mouse *lefty/stra3* of the TGF-β superfamily. A motif search revealed that the predicted EBAF protein contains most of the cysteine residues which are conserved among the TGF-β-related proteins and which are necessary for the formation of the cysteine knot structure. The EBAF sequence contains an additional cysteine residue, 12 amino acids upstream from the first conserved cysteine residue. The only other family members known to contain an additional cysteine residue are TGF-βs, inhibins, and GDF-3. EBAF, similar to LEFTY, GDF-3/Vgr2, and GDF-9, lacks the cysteine residue that is known to form the intermolecular disulfide bond. Therefore, EBAF appears to be an additional member of the TGF-β superfamily with an unpaired cysteine residue that may not exist as a dimer. However, hydrophobic contacts between the two monomer subunits may promote dimer formation. Fluorescence *in situ* hybridization showed that the *ebaf* gene is located on human chromosome 1 at band q42.1.

Additional members of the TGF-β superfamily, such as those related to EBAF, are being searched for by industry and academics. We herein describe the identification and characterization of novel polypeptides having homology to EBAF, designated herein as PRO317 polypeptides.

18. PRO301

The widespread occurrence of cancer has prompted the devotion of considerable resources and discovering new treatments of treatment. One particular method involves the creation of tumor or cancer specific monoclonal antibodies (mAbs) which are specific to tumor antigens. Such mAbs, which can distinguish between normal and cancerous cells are useful in the diagnosis, prognosis and treatment of the disease. Particular antigens are known to be associated with neoplastic diseases, such as colorectal cancer.

One particular antigen, the A33 antigen is expressed in more than 90% of primary or metastatic colon cancers as well as normal colon epithelium. Since colon cancer is a widespread disease, early diagnosis and treatment is an important medical goal. Diagnosis and treatment of colon cancer can be implemented using monoclonal antibodies (mAbs) specific therefore having fluorescent, nuclear magnetic or radioactive tags. Radioactive gene, toxins and/or drug tagged mAbs can be used for treatment *in situ* with minimal patient description. mAbs can also be used to diagnose during the diagnosis and treatment of colon cancers. For example, when the serum levels of the A33 antigen are elevated in a patient, a drop of the levels after surgery would indicate the tumor resection was successful. On the other hand, a subsequent rise in serum A33 antigen levels after surgery would indicate that metastases of the original tumor may have formed or that new primary tumors may have appeared. Such monoclonal antibodies can be used in lieu of, or in conjunction with surgery and/or other chemotherapies. For example, U.S.P. 4,579,827 and U.S.S.N. 424,991 (E.P. 199,141) are directed to therapeutic administration of monoclonal antibodies, the latter of which relates to the application of anti-A33 mAb.

Many cancers of epithelial origin have adenovirus receptors. In fact, adenovirus-derived vectors have

been proposed as a means of inserting antisense nucleic acids into tumors (U.S.P. 5,518,885). Thus, the association of viral receptors with neoplastic tumors is not unexpected.

We herein describe the identification and characterization of novel polypeptides having homology to certain cancer-associated antigens, designated herein as PRO301 polypeptides.

19. PRO224

Cholesterol uptake can have serious implications on one's health. Cholesterol uptake provides cells with most of the cholesterol they require for membrane synthesis. If this uptake is blocked, cholesterol accumulates in the blood and can contribute to the formation of atherosclerotic plaques in blood vessel walls. Most cholesterol is transported in the blood bound to protein in the form of complexes known as low-density lipoproteins (LDLs). LDLs are endocytosed into cells via LDL receptor proteins. Therefore, LDL receptor proteins, and proteins having homology thereto, are of interest to the scientific and medical communities.

Membrane-bound proteins and receptors can play an important role in the formation, differentiation and maintenance of multicellular organisms. The LDL receptors are an example of membrane-bound proteins which are involved in the synthesis and formation of cell membranes, wherein the health of an individual is affected directly and indirectly by its function. Many membrane-bound proteins act as receptors such as the LDL receptor. These receptors can function to endocytose substrates or they can function as a receptor for a channel. Other membrane-bound proteins function as signals or antigens.

Membrane-bound proteins and receptor molecules have various industrial applications, including as pharmaceutical and diagnostic agents. The membrane-bound proteins can also be employed for screening of potential peptide or small molecule regulators of the relevant receptor/ligand interaction. In the case of the LDL receptor, it is desirable to find molecules which enhance endocytosis so as to lower blood cholesterol levels and plaque formation. It is also desirable to identify molecules which inhibit endocytosis so that these molecules can be avoided or regulated by individuals having high blood cholesterol. Polypeptides which are homologous to lipoprotein receptors but which do not function as lipoprotein receptors are also of interest in the determination of the function of the fragments which show homology.

The following studies report on previously known low density lipoprotein receptors and related proteins including apolipoproteins: Sawamura, et al., Nippon Chemiphar Co, Japan patent application J09098787; Novak, S., et al., J. Biol. Chem., 271:(20)11732-6 (1996); Blaas, D., J. Virol., 69(11)7244-7 (Nov. 1995); Scott, J., J. Inherit. Metab. Dis. (UK), 9/Supp. 1 (3-16) (1986); Yamamoto, et al., Cell, 39:27-38 (1984); Rebece, et al., Neurobiol. Aging, 15:5117 (1994); Novak, S., et al., J. Biol. Chemistry, 271:11732-11736 (1996); and Sestavel and Fruchart, Cell Mol. Biol., 40(4):461-81 (June 1994). These publications and others published prior to the filing of this application provide further background to peptides already known in the art.

Efforts are being undertaken by both industry and academia to identify new, native membrane-bound receptor proteins, particularly those having homology to lipoprotein receptors. We herein describe the identification and characterization of novel polypeptides having homology to lipoprotein receptors, designated herein as PRO224 polypeptides.

20. PRO222

Complement is a group of proteins found in the blood that are important in humoral immunity and inflammation. Complement proteins are sequentially activated by antigen-antibody complexes or by proteolytic enzymes. When activated, complement proteins kill bacteria and other microorganisms, affect vascular permeability, release histamine and attract white blood cells. Complement also enhances phagocytosis when bound to target cells. In order to prevent harm to autologous cells, the complement activation pathway is tightly regulated.

Deficiencies in the regulation of complement activation or in the complement proteins themselves may lead to immune-complex diseases, such as systemic lupus erythematosus, and may result in increased susceptibility to bacterial infection. In all cases, early detection of complement deficiency is desirable so that the patient can begin treatment. Thus, research efforts are currently directed toward identification of soluble and membrane proteins that regulate complement activation.

Proteins known to be important in regulating complement activation in humans include Factor H and Complement receptor type 1 (CR1). Factor H is a 150 kD soluble serum protein that interacts with complement protein C3b to accelerate the decay of C3 convertase and acts as a cofactor for Factor I-mediated cleavage of complement protein C4b. Complement receptor type 1 is a 190-280 kD membrane bound protein found in mast cells and most blood cells. CR1 interacts with complement proteins C3b, C4b, and iC3b to accelerate dissociation of C3 convertases, acts as a cofactor for Factor I-mediated cleavage of C3b and C4b, and binds immune complexes and promotes their dissolution and phagocytosis.

Proteins which have homology to complement proteins are of particular interest to the medical and industrial communities. Often, proteins having homology to each other have similar function. It is also of interest when proteins having homology do not have similar functions, indicating that certain structural motifs identify information other than function, such as locality of function.

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound proteins, particularly those having homology to known proteins involved in the complement pathway. Proteins involved in the complement pathway were reviewed in Birmingham DJ (1995), Critical Reviews in Immunology, 15(2):133-154 and in Abbas AK, et al. (1994) Cellular and Molecular Immunology, 2nd Ed. W.B. Saunders Company, Philadelphia, pp 295-315.

We herein describe the identification and characterization of novel polypeptides having homology to complement receptors, designated herein as PRO222 polypeptides.

21. PRO234

The successful function of many systems within multicellular organisms is dependent on cell-cell interactions. Such interactions are affected by the alignment of particular ligands with particular receptors in a manner which allows for ligand-receptor binding and thus a cell-cell adhesion. While protein-protein interactions in cell recognition have been recognized for some time, only recently has the role of carbohydrates in physiologically relevant recognition been widely considered (see B.K. Brandley *et al., J. Leuk. Biol.* 40: 97 (1986) and N. Sharon *et al., Science* 246: 227 (1989). Oligosaccharides are well positioned to act as recognition novel lectins due to their cell surface location and structural diversity. Many oligosaccharide structures can be created through the differential activities of a smaller number of glycosyltransferases. The diverse structures

of oligosaccharides can be generated by transcription of relatively few gene products, which suggests that the oligosaccharides are a plausible mechanism by which is directed a wide range of cell-cell interactions. Examples of differential expression of cell surface carbohydrates and putative carbohydrate binding proteins (lectins) on interacting cells have been described (J. Dodd & T.M. Jessel, *J. Neurosci.* 5: 3278 (1985); L.J. Regan *et al., Proc. Natl. Acad. Sci. USA* 83: 2248 (1986); M. Constantine-Paton *et al., Nature* 324: 459 (1986); and M. Tiemeyer *et al., J. Biol. Chem.* 263: 1671 (1989). One interesting member of the lectin family are selectins.

The migration of leukocytes to sites of acute or chronic inflammation involves adhesive interactions between these cells and the endothelium. This specific adhesion is the initial event in the cascade that is initiated by inflammatory insults, and it is, therefore, of paramount importance to the regulated defense of the organism.

The types of cell adhesion molecules that are involved in the interaction between leukocytes and the endothelium during an inflammatory response currently stands at four: (1) selectins; (2) (carbohydrate and glycoprotein) ligands for selectins; (3) integrins; and (4) integrin ligands, which are members of the immunoglobulin gene superfamily.

The selectins are cell adhesion molecules that are unified both structurally and functionally. Structurally, selectins are characterized by the inclusion of a domain with homology to a calcium-dependent lectin (C-lectins), an epidermal growth factor (egf)-like domain and several complement binding-like domains, Bevilacqua, M.P. *et al., Science* 243: 1160-1165 (1989); Johnston *et al., Cell* 56: 1033-1044 (1989); Lasky *et al, Cell* 56: 1045-1055 (1989); Siegalman, M. *et al., Science* 243: 1165-1172 (1989); Stoolman, L.M., *Cell* 56: 907-910 (1989). Functionally, selectins share the common property of their ability to mediate cell binding through interactions between their lectin domains and cell surface carbohydrate ligands (Brandley, B, *et al., Cell* 63, 861-863 (1990); Springer, T. and Lasky, L.A., *Nature* 349, 19-197 (1991); Bevilacqua, M.P. and Nelson, R.M., *J. Clin. Invest.* 91 379-387 (1993) and Tedder *et al., J. Exp. Med.* 170: 123-133 (1989).

There are three members identified so far in the selectin family of cell adhesion molecules: L-selectin (also called peripheral lymph node homing receptor (pnHR), LEC-CAM-1, LAM-1, gp90^{MEL}, gp100^{MEL}, gp110^{MEL}, MEL-14 antigen, Leu-8 antigen, TQ-1 antigen, DREG antigen), E-selectin (LEC-CAM-2, LECAM-2, ELAM-1) and P-selectin (LEC-CAM-3, LECAM-3, GMP-140, PADGEM).

The identification of the C-lectin domain has led to an intense effort to define carbohydrate binding ligands for proteins containing such domains. E-selectin is believed to recognize the carbohydrate sequence NeuNAcα2-3Galβ1-4(Fucα1-3)GlcNAc (sialyl-Lewis x, or sLe^x) and related oligosaccharides, Berg *et al., J. Biol. Chem.* 265: 14869-14872 (1991); Lowe *et al., Cell* 63: 475-484 (1990); Phillips *et al., Science* 250: 1130-1132 (1990); *Tiemeyer et al., Proc. Natl. Acad. Sci. USA* 88: 1138-1142 (1991).

L-selectin, which comprises a lectin domain, performs its adhesive function by recognizing carbohydrate-containing ligands on endothelial cells. L-selectin is expressed on the surface of leukocytes, such as lymphocytes, neutrophils, monocytes and eosinophils, and is involved with the trafficking of lymphocytes to peripheral lymphoid tissues (*Gallatin et al., Nature* 303: 30-34 (1983)) and with acute neutrophil-medicated inflammatory responses (Watson, S.R., *Nature* 349: 164-167 (1991)). The amino acid sequence of L-selectin and the encoding nucleic acid sequence are, for example, disclosed in U.S. patent No. 5,098,833 issued 24 March 1992.

L-selectin (LECAM-1) is particularly interesting because of its ability to block neutrophil influx (Watson

et al., Nature 349: 164-167 (1991). It is expressed in chronic lymphocytic leukemia cells which bind to HEV (Spertini *et al., Nature* 349: 691-694 (1991). It is also believed that HEV structures at sites of chronic inflammation are associated with the symptoms of diseases such as rheumatoid arthritis, psoriasis and multiple sclerosis.

E-selectin (ELAM-1), is particularly interesting because of its transient expression on endothelial cells in response to IL-1 or TNF. Bevilacqua et al., *Science* 243: 1160 (1989). The time course of this induced expression (2-8 h) suggests a role for this receptor in initial neutrophil induced extravasation in response to infection and injury. It has further been reported that anti-ELAM-1 antibody blocks the influx of neutrophils in a primate asthma model and thus is beneficial for preventing airway obstruction resulting from the inflammatory response. Gundel *et al., J. Clin. Invest.* 88: 1407 (1991).

The adhesion of circulating neutrophils to stimulated vascular endothelium is a primary event of the inflammatory response. P-selectin has been reported to recognize the Lewis x structure (Galβ1-4(Fucα1-3) GlcNAc), Larsen *et al., Cell* 63: 467-474(1990). Others report that an additional terminal linked sialic acid is required for high affinity binding, Moore *et al., J. Cell. Biol.* 112: 491-499 (1991). P-selectin has been shown to be significant in acute lung injury. Anti-P-selectin antibody has been shown to have strong protective effects in a rodent lung injury model. M.S. Mulligan *et al., J. Clin. Invest.* 90: 1600 (1991).

We herein describe the identification and characterization of novel polypeptides having homology to lectin proteins, herein designated as PRO234 polypeptides.

22. PRO231

Some of the most important proteins involved in the above described regulation and modulation of cellular processes are the enzymes which regulate levels of protein phosphorylation in the cell. For example, it is known that the transduction of signals that regulate cell growth and differentiation is regulated at least in part by phosphorylation and dephosphorylation of various cellular proteins. The enzymes that catalyze these processes include the protein kinases, which function to phosphorylate various cellular proteins, and the protein phosphatases, which function to remove phosphate residues from various cellular proteins. The balance of the level of protein phosphorylation in the cell is thus mediated by the relative activities of these two types of enzymes.

Protein phosphatases represent a growing family of enzymes that are found in many diverse forms, including both membrane-bound and soluble forms. While many protein phosphatases have been described, the functions of only a very few are beginning to be understood (Tonks, *Semin. Cell Biol.* 4:373-453 (1993) and Dixon, *Recent Prog. Horm. Res.* 51:405-414 (1996)). However, in general, it appears that many of the protein phosphatases function to modulate the positive or negative signals induced by various protein kinases. Therefore, it is likely that protein phosphatases play critical roles in numerous and diverse cellular processes.

Given the physiological importance of the protein phosphatases, efforts are being undertaken by both industry and academia to identify new, native phosphatase proteins. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel phosphatase proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to acid phosphatases, designated herein as PRO231 polypeptides.

23. PRO229

Scavenger receptors are known to protect IgG molecules from catabolic degradation. Riechmann and Hollinger, Nature Biotechnology, 15:617 (1997). In particular, studies of the CH2 and CH3 domains have shown that specific sequences of these domains are important in determining the half-lives of antibodies. Ellerson, et al., J. Immunol., 116: 510 (1976); Yasmeen, et al., J. Immunol. 116: 518 (1976; Pollock, et al., Eur. J. Immunol., 20: 2021 (1990). Scavenger receptor proteins and antibodies thereto are further reported in U.S. Patent No. 5,510,466 to Krieger, et al. Due to the ability of scavenger receptors to increase the half-life of polypeptides and their involvement in immune function, molecules having homology to scavenger receptors are of importance to the scientific and medical community.

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins, particularly those having homology to scavenger receptors. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to scavenger receptors, designated herein as PRO229 polypeptides.

24. PRO238

Oxygen free radicals and antioxidants appear to play an important role in the central nervous system after cerebral ischemia and reperfusion. Moreover, cardiac injury, related to ischaemia and reperfusion has been reported to be caused by the action of free radicals. Additionally, studies have reported that the redox state of the cell is a pivotal determinant of the fate of the cells. Furthermore, reactive oxygen species have been reported to be cytotoxic, causing inflammatory disease, including tissue necrosis, organ failure, atherosclerosis, infertility, birth defects, premature aging, mutations and malignancy. Thus, the control of oxidation and reduction is important for a number of reasons including for control and prevention of strokes, heart attacks, oxidative stress and hypertension. In this regard, reductases, and particularly, oxidoreductases, are of interest. Publications further describing this subject matter include Kelsey, et al., Br. J. Cancer, 76(7):852-4 (1997); Friedrich and Weiss, J. Theor. Biol., 187(4):529-40 (1997) and Pieulle, et al., J. Bacteriol., 179(18):5684-92 (1997).

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins, particularly secreted proteins which have homology to reductase. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to

reductase, designated herein as PRO238 polypeptides.

25. PRO233

Studies have reported that the redox state of the cell is an important determinant of the fate of the cell. Furthermore, reactive oxygen species have been reported to be cytotoxic, causing inflammatory disease, including tissue necrosis, organ failure, atherosclerosis, infertility, birth defects, premature aging, mutations and malignancy. Thus, the control of oxidation and reduction is important for a number of reasons, including the control and prevention of strokes, heart attacks, oxidative stress and hypertension. Oxygen free radicals and antioxidants appear to play an important role in the central nervous system after cerebral ischemia and reperfusion. Moreover, cardiac injury, related to ischaemia and reperfusion has been reported to be caused by the action of free radicals. In this regard, reductases, and particularly, oxidoreductases, are of interest. In addition, the transcription factors, NF-kappa B and AP-1, are known to be regulated by redox state and to affect the expression of a large variety of genes thought to be involved in the pathogenesis of AIDS, cancer, atherosclerosis and diabetic complications. Publications further describing this subject matter include Kelsey, et al., Br. J. Cancer, 76(7):852-4 (1997); Friedrich and Weiss, J. Theor. Biol., 187(4):529-40 (1997) and Pieulle, et al., J. Bacteriol., 179(18):5684-92 (1997). Given the physiological importance of redox reactions *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in redox reactions. We describe herein the identification of novel polypeptides which have homology to reductase, designated herein as PRO233 polypeptides.

26. PRO223

The carboxypeptidase family of exopeptidases constitutes a diverse group of enzymes that hydrolyze carboxyl-terminal amide bonds in polypeptides, wherein a large number of mammalian tissues produce these enzymes. Many of the carboxypeptidase enzymes that have been identified to date exhibit rather strong cleavage specificities for certain amino acids in polypeptides. For example, carboxypeptidase enzymes have been identified which prefer lysine, arginine, serine or amino acids with either aromatic or branched aliphatic side chains as substrates at the carboxyl terminus of the polypeptide.

With regard to the serine carboxypeptidases, such amino acid specific enzymes have been identified from a variety of different mammalian and non-mammalian organisms. The mammalian serine carboxypeptidase enzymes play important roles in many different biological processes including, for example, protein digestion, activation, inactivation, or modulation of peptide hormone activity, and alteration of the physical properties of proteins and enzymes.

In light of the physiological importance of the serine carboxypeptidases, efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins and specifically novel carboxypeptidases. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. We describe herein novel polypeptides having homology to one or more serine carboxypeptidase polypeptides, designated herein as PRO223 polypeptides.

27. **PRO235**

Plexin was first identified in Xenopus tadpole nervous system as a membrane glycoprotein which was shown to mediate cell adhesion via a homophilic binding mechanism in the presence of calcium ions. Strong evolutionary conservation between Xenopus, mouse and human homologs of plexin has been observed. [Kaneyama et al., Biochem. And Biophys. Res. Comm. 226: 524-529 (1996)]. Given the physiological importance of cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in cell adhesion. We describe herein the identification of a novel polypeptide which has homology to plexin, designated herein as PRO235.

28. **PRO236 and PRO262**

β-galactosidase is a well known enzymatic protein which functions to hydrolyze β-galactoside molecules. β-galactosidase has been employed for a variety of different applications, both *in vitro* and *in vivo* and has proven to be an extremely useful research tool. As such, there is an interest in obtaining novel polypeptides which exhibit homology to the β-galactosidase polypeptide.

Given the strong interest in obtaining novel polypeptides having homology to β-galactosidase, efforts are currently being undertaken by both industry and academia to identify new, native β-galactosidase homolog proteins. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel β-galactosidase-like proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)]. We herein describe novel poylpeptides having siginificant homology to the β-galactosidase enzyme, designated herein as PRO236 and PRO262 polypeptides.

29. **PRO239**

Densin is a glycoprotein which has been isolated from the brain which has all the hallmarks of an adhesion molecule. It is highly concentrated at synaptic sites in the brain and is expressed prominently in dendritic processes in developing neurons. Densin has been characterized as a member of the O-linked sialoglycoproteins. Densin has relevance to medically important processes such as regeneration. Given the physiological importance of synaptic processes and cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in synaptic machinery and cell adhesion. We describe herein the identification of novel polypeptides which have homology to densin, designated herein as PRO239 polypeptides.

30. **PRO257**

Ebnerin is a cell surface protein associated with von Ebner glands in mammals. Efforts are being undertaken by both industry and academia to identify new, native cell surface receptor proteins and specifically those which possess sequence homology to cell surface proteins such as ebnerin. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel receptor proteins. We herein describe the identification of novel polypeptides having significant homology to the von Ebner's gland-associated protein ebnerin, designated herein as PRO257 polypeptides.

31. **PRO260**

Fucosidases are enzymes that remove fucose residues from fucose containing proteoglycans. In some pathological conditions, such as cancer, rheumatoid arthritis, and diabetes, there is an abnormal fucosylation of serum proteins. Therefore, fucosidases, and proteins having homology to fucosidase, are of importance to the study and abrogation of these conditions. In particular, proteins having homology to the alpha-l-fucosidase precursor are of interest. Fucosidases and fucosidase inhibitors are further described in U.S. Patent Nos. 5,637,490, 5,382,709, 5,240,707, 5,153,325, 5,100,797, 5,096,909 and 5,017,704. Studies are also reported in Valk, et al., J. Virol., 71(9):6796 (1997), Aktogu, et al., Monaldi. Arch. Chest Dis. (Italy), 52(2):118 (1997) and Focarelli, et al., Biochem. Biophys. Res. Commun. (U.S.), 234(1):54 (1997).

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins. Of particular interest are proteins having homology to the alpha-l-fucosidase precursor. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to fucosidases, designated herein as PRO260 polypeptides.

32. **PRO263**

CD44 is a cell surface adhesion molecule involved in cell-cell and cell-matrix interactions. Hyaluronic acid, a component of the extracellular matrix is a major ligand. Other ligands include collagen, fibronectin, laminin, chrondroitin sulfate, mucosal addressin, serglycin and osteoponin. CD44 is also important in regulating cell traffic, lymph node homing, transmission of growth signals, and presentation of chemokines and growth factors to traveling cells. CD44 surface proteins are associated with metastatic tumors and CD44 has been used as a marker for HIV infection. Certain splice variants are associated with metastasis and poor prognosis of cancer patients. Therefore, molecules having homology with CD44 are of particular interest, as their homology indicates that they may have functions related to those functions of CD44. CD44 is further described in U.S. Patent Nos. 5,506,119, 5,504,194 and 5,108,904; Gerberick, et al., Toxicol. Appl. Pharmacol., 146(1):1 (1997); Wittig, et al., Immunol. Letters (Netherlands), 57(1-3):217 (1997); and Oliveira and Odell, Oral Oncol. (England), 33(4):260 (1997).

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins, particularly transmembrane proteins with homology to CD44 antigen. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to CD44 antigen, designated herein as PRO263 polypeptides.

33. **PRO270**

Thioredoxins effect reduction-oxidation (redox) state. Many diseases are potentially related to redox state and reactive oxygen species may play a role in many important biological processes. The transcription factors, NF-kappa B and AP-1, are regulated by redox state and are known to affect the expression of a large variety of genes thought to be involved in the pathogenesis of AIDS, cancer, atherosclerosis and diabetic complications. Such proteins may also play a role in cellular antioxidant defense, and in pathological conditions involving oxidative stress such as stroke and inflammation in addition to having a role in apoptosis. Therefore, thioredoxins, and proteins having homology thereto, are of interest to the scientific and medical communities.

We herein describe the identification and characterization of novel polypeptides having homology to thioredoxin, designated herein as PRO270 polypeptides.

34. **PRO271**

The proteoglycan link protein is a protein which is intimately associated with various extracellular matrix proteins and more specifically with proteins such as collagen. For example, one primary component of collagen is a large proteoglycan called aggrecan. This molecule is retained by binding to the glycosaminoglycan hyaluronan through the amino terminal G1 globular domain of the core protein. This binding is stabilized by the proteoglycan link protein which is a protein that is also associated with other tissues containing hyaluronan binding proteoglycans such as versican.

Link protein has been identified as a potential target for autoimmune antibodies in individuals who suffer from juvenile rheumatoid arthritis (see Guerassimov et al., *J. Rheumatology* 24(5):959-964 (1997)). As such, there is strong interest in identifying novel proteins having homology to link protein. We herein describe the identification and characterization of novel polypeptides having such homology, designated herein as PRO271 polypeptides.

35. **PRO272**

Reticulocalbin is an endoplasmic reticular protein which may be involved in protein transport and luminal protein processing. Reticulocalbin resides in the lumen of the endopladsmic rerticulum, is known to bind calcium, and may be involved in a luminal retention mechanism of the endoplasmic reticulum. It contains six domains of the EF-hand motif associated with high affinity calcium binding. We describe herein the identification and characterization of a novel polypeptide which has homology to the reticulocalbin protein, designated herein as PRO272.

36. **PRO294**

Collagen, a naturally occurring protein, finds wide application in industry. Chemically hydrolyzed natural collagen can be denatured and renatured by heating and cooling to produce gelatin, which is used in photographic and medical, among other applications. Collagen has important properties such as the ability to form interchain aggregates having a conformation designated as a triple helix. We herein describe the identification and characterization of a novel polypeptide which has homology to portions of the collagen molecule, designated herein as PRO294.

37. PRO295

The integrins comprise a supergene family of cell-surface glycoprotein receptors that promote cellular adhesion. Each cell has numerous receptors that define its cell adhesive capabilities. Integrins are involved in a wide variety of interaction between cells and other cells or matrix components. The integrins are of particular importance in regulating movement and function of immune system cells The platelet IIb/IIIA integrin complex is of particular importance in regulating platelet aggregation. A member of the integrin family, integrin β-6, is expressed on epithelial cells and modulates epithelial inflammation. Another integrin, leucocyte-associated antigen-1 (LFA-1) is important in the adhesion of lymphocytes during an immune response. The integrins are expressed as heterodimers of non-covalently associated alpha and beta subunits. Given the physiological importance of cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in cell adhesion. We describe herein the identification and characterization of a novel polypeptide which has homology to integrin, designated herein as PRO295.

38. PRO293

Protein-protein interactions include receptor and antigen complexes and signaling mechanisms. As more is known about the structural and functional mechanisms underlying protein-protein interactions, protein-protein interactions can be more easily manipulated to regulate the particular result of the protein-protein interaction. Thus, the underlying mechanisms of protein-protein interactions are of interest to the scientific and medical community.

All proteins containing leucine-rich repeats are thought to be involved in protein-protein interactions. Leucine-rich repeats are short sequence motifs present in a number of proteins with diverse functions and cellular locations. The crystal structure of ribonuclease inhibitor protein has revealed that leucine-rich repeats correspond to beta-alpha structural units. These units are arranged so that they form a parallel beta-sheet with one surface exposed to solvent, so that the protein acquires an unusual, nonglubular shape. These two features have been indicated as responsible for the protein-binding functions of proteins containing leucine-rich repeats. See, Kobe and Deisenhofer, Trends Biochem. Sci., 19(10):415-421 (Oct. 1994).

A study has been reported on leucine-rich proteoglycans which serve as tissue organizers, orienting and ordering collagen fibrils during ontogeny and are involved in pathological processes such as wound healing, tissue repair, and tumor stroma formation. Iozzo, R. V., Crit. Rev. Biochem. Mol. Biol., 32(2):141-174 (1997). Others studies implicating leucine rich proteins in wound healing and tissue repair are De La Salle, C., et al., Vouv. Rev. Fr. Hematol. (Germany), 37(4):215-222 (1995), reporting mutations in the leucine rich motif in a complex associated with the bleeding disorder Bernard-Soulier syndrome and Chlemetson, K. J., Thromb. Haemost. (Germany), 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats. Another protein of particular interest which has been reported to have leucine-rich repeats is the SLIT protein which has been reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanistsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Other studies reporting on the biological functions of proteins having leucine-rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996) (gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996)

(apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement); and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation (decorin binding to transforming growth factorβ involvement for treatment for cancer, wound healing and scarring).

Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions. Of particular interest are those proteins having leucine rich repeats and homology to known neuronal leucine rich repeat proteins. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound proteins having leucine rich repeats. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We describe herein the identification and characterization of a novel polypeptide which has homology to leucine rich repeat proteins, designated herein as PRO293.

39. PRO247

Protein-protein interactions include receptor and antigen complexes and signaling mechanisms. As more is known about the structural and functional mechanisms underlying protein-protein interactions, protein-protein interactions can be more easily manipulated to regulate the particular result of the protein-protein interaction. Thus, the underlying mechanisms of protein-protein interactions are of interest to the scientific and medical community.

All proteins containing leucine-rich repeats are thought to be involved in protein-protein interactions. Leucine-rich repeats are short sequence motifs present in a number of proteins with diverse functions and cellular locations. The crystal structure of ribonuclease inhibitor protein has revealed that leucine-rich repeats correspond to beta-alpha structural units. These units are arranged so that they form a parallel beta-sheet with one surface exposed to solvent, so that the protein acquires an unusual, nonglubular shape. These two features have been indicated as responsible for the protein-binding functions of proteins containing leucine-rich repeats. See, Kobe and Deisenhofer, Trends Biochem. Sci., 19(10):415-421 (Oct. 1994).

A study has been reported on leucine-rich proteoglycans which serve as tissue organizers, orienting and ordering collagen fibrils during ontogeny and are involved in pathological processes such as wound healing, tissue repair, and tumor stroma formation. Iozzo, R. V., Crit. Rev. Biochem. Mol. Biol., 32(2):141-174 (1997). Others studies implicating leucine rich proteins in wound healing and tissue repair are De La Salle, C., et al., Vouv. Rev. Fr. Hematol. (Germany), 37(4):215-222 (1995), reporting mutations in the leucine rich motif in a complex associated with the bleeding disorder Bernard-Soulier syndrome and Chlemetson, K. J., Thromb. Haemost. (Germany), 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats. Another protein of particular interest which has been reported to have leucine-rich repeats is the SLIT protein which has been reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanistsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Other studies reporting on the biological functions of proteins having leucine-rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996)

(gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996) (apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement); and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation (decorin binding to transforming growth factorβ involvement for treatment for cancer, wound healing and scarring).

Densin is a glycoprotein which has been isolated from the brain which has all the hallmarks of an adhesion molecule. It is highly concentrated at synaptic sites in the brain and is expressed prominently in dendritic processes in developing neurons. Densin has been characterized as a member of the O-linked sialoglycoproteins. Densin has relevance to medically important processes such as regeneration. Given the physiological importance of synaptic processes and cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in synaptic machinery and cell adhesion. Densin is further described in Kennedy, M.B, Trends Neurosci. (England), 20(6):264 (1997) and Apperson, et al., J. Neurosci., 16(21):6839 (1996).

Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions. Of particular interest are those proteins having leucine rich repeats and homology to known proteins having leucine rich repeats such as KIAA0231 and densin. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound proteins having leucine rich repeats. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We describe herein the identification and characterization of a novel polypeptide which has homology to leucine rich repeat proteins, designated herein as PRO247.

40. PRO302, PRO303, PRO304, PRO307 and PRO343

Proteases are enzymatic proteins which are involved in a large number of very important biological processes in mammalian and non-mammalian organisms. Numerous different protease enzymes from a variety of different mammalian and non-mammalian organisms have been both identified and characterized. The mammalian protease enzymes play important roles in many different biological processes including, for example, protein digestion, activation, inactivation, or modulation of peptide hormone activity, and alteration of the physical properties of proteins and enzymes.

In light of the important physiological roles played by protease enzymes, efforts are currently being undertaken by both industry and academia to identify new, native protease homologs. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)]. We herein describe the identification of novel polypeptides having homology to various protease enzymes, designated herein as PRO302, PRO303, PRO304, PRO307 and PRO343 polypeptides.

41. PRO328

The GLIP protein family has been characterized as comprising zinc-finger proteins which play important roles in embryogenesis. These proteins may function as transcriptional regulatory proteins and are known to be amplified in a subset of human tumors. Glioma pathogenesis protein is structurally related to a group of plant pathogenesis-related proteins. It is highly expressed in glioblastoma. See US Pat. Nos. 5,582,981 (issued Dec. 10, 1996) and 5,322,801 (issued June 21, 1996), Ellington, A.D. et al., Nature, 346:818 (1990), Grindley, J.C. et al., Dev. Biol., 188(2):337 (1997), Marine, J.C. et al., Mech. Dev., 63(2):211 (1997), The CRISP or cysteine rich secretory protein family are a group of proteins which are also structurally related to a group of plant pathogenesis proteins. [Schwidetzky, U., Biochem. J., 321:325 (1997), Pfisterer, P., Mol. Cell Biol., 16(11):6160 (1996), Kratzschmar, J., Eur. J. Biochem., 236(3):827 (1996)]. We describe herein the identification of a novel polypeptide which has homology to GLIP and CRISP, designated herein as PRO328 polypeptides.

42. PRO335, PRO331 and PRO326

Protein-protein interactions include receptor and antigen complexes and signaling mechanisms. As more is known about the structural and functional mechanisms underlying protein-protein interactions, protein-protein interactions can be more easily manipulated to regulate the particular result of the protein-protein interaction. Thus, the underlying mechanisms of protein-protein interactions are of interest to the scientific and medical community.

All proteins containing leucine-rich repeats are thought to be involved in protein-protein interactions. Leucine-rich repeats are short sequence motifs present in a number of proteins with diverse functions and cellular locations. The crystal structure of ribonuclease inhibitor protein has revealed that leucine-rich repeats correspond to beta-alpha structural units. These units are arranged so that they form a parallel beta-sheet with one surface exposed to solvent, so that the protein acquires an unusual, nonglubular shape. These two features have been indicated as responsible for the protein-binding functions of proteins containing leucine-rich repeats. See, Kobe and Deisenhofer, Trends Biochem. Sci., 19(10):415-421 (Oct. 1994).

A study has been reported on leucine-rich proteoglycans which serve as tissue organizers, orienting and ordering collagen fibrils during ontogeny and are involved in pathological processes such as wound healing, tissue repair, and tumor stroma formation. Iozzo, R. V., Crit. Rev. Biochem. Mol. Biol., 32(2):141-174 (1997). Others studies implicating leucine rich proteins in wound healing and tissue repair are De La Salle, C., et al., Vouv. Rev. Fr. Hematol. (Germany), 37(4):215-222 (1995), reporting mutations in the leucine rich motif in a complex associated with the bleeding disorder Bernard-Soulier syndrome, Chlemetson, K. J., Thromb. Haemost. (Germany), 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation reporting that decorin binding to transforming growth factorβ has involvement in a treatment for cancer, wound healing and scarring. Related by function to this group of proteins is the insulin like growth factor (IGF), in that it is useful in wound-healing and associated therapies concerned with re-growth of tissue, such as connective tissue, skin and bone; in promoting body growth in humans and animals; and in stimulating other growth-related processes. The acid labile subunit of IGF (ALS) is also of interest in that it increases the half-life of IGF and is part of the IGF complex in vivo.

Another protein which has been reported to have leucine-rich repeats is the SLIT protein which has been

reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanistsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Of particular interest is LIG-1, a membrane glycoprotein that is expressed specifically in glial cells in the mouse brain, and has leucine rich repeats and immunoglobulin-like domains. Suzuki, et al., J. Biol. Chem. (U.S.), 271(37):22522 (1996). Other studies reporting on the biological functions of proteins having leucine rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996) (gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996) (apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement).

Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions. Of particular interest are those proteins having leucine rich repeats and homology to known proteins having leucine rich repeats such as LIG-1, ALS and decorin. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound proteins having leucine rich repeats. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We describe herein the identification and characterization of novel polypeptides which have homology to proteins of the leucine rich repeat superfamily, designated herein as PRO335, PRO331 and PRO326 polypeptides.

43. PRO332

Secreted proteins comprising a repeat characterized by an arrangement of conserved leucine residues (leucine-rich repeat motif) have diverse biological roles. Certain proteoglycans, such as biglycan, fibromodulin and decorin, are, for example, characterized by the presence of a leucine-rich repeat of about 24 amino acids [Ruoslahti, Ann. Rev. Cell. Biol. 4 229-255 (1988); Oldberg *et al.*, EMBO J. 8, 2601-2604 (1989)]. In general, proteoglycans are believed to play a role in regulating extracellular matrix, cartilage or bone function. The proteoglycan decorin binds to collagen type I and II and affects the rate of fibril formation. Fibromodulin also binds collagen and delays fibril formation. Both fibromodulin and decorin inhibit the activity of transforming growth factor beta (TGF-β) (U.S. Patent No. 5,583,103 issued December 10, 1996). TGF-β is known to play a key role in the induction of extracellular matrix and has been implicated in the development of fibrotic diseases, such as cancer and glomerulonephritis. Accordingly, proteoglycans have been proposed for the treatment of fibrotic cancer, based upon their ability to inhibit TGF-β's growth stimulating activity on the cancer cell. Proteoglycans have also been described as potentially useful in the treatment of other proliferative pathologies, including rheumatoid arthritis, arteriosclerosis, adult respiratory distress syndrome, cirrhosis of the liver, fibrosis of the lungs, post-myocardial infarction, cardiac fibrosis, post-angioplasty restenosis, renal interstitial fibrosis and certain dermal fibrotic conditions, such as keloids and scarring, which might result from burn injuries, other invasive skin injuries, or cosmetic or reconstructive surgery (U.S. Patent No. 5,654,270, issued August 5, 1997).

We describe herein the identification and characterization of novel polypeptides which have homology

to proteins of the leucine rich repeat superfamily, designated herein as PRO332 polypeptides.

44. PRO334

Microfibril bundles and proteins found in association with these bundles, particularly attachment molecules, are of interest in the field of dermatology, particularly in the study of skin which has been damaged from aging, injuries or the sun. Fibrillin microfibrils define the continuous elastic network of skin, and are present in dermis as microfibril bundles devoid of measurable elastin extending from the dermal-epithelial junction and as components of the thick elastic fibres present in the deep reticular dermis. Moreover, Marfan syndrome has been linked to mutations which interfere with multimerization of fibrillin monomers or other connective tissue elements.

Fibulin-1 is a modular glycoprotein with amino-terminal anaphlatoxin-like modules followed by nine epidermal growth factor (EGF)-like modules and, depending on alternative splicing, four possible carboxyl termini. Fibulin-2 is a novel extracellular matrix protein frequently found in close association with microfibrils containing either fibronectin or fibrillin. Thus, fibrillin, fibulin, and molecules related thereto are of interest, particularly for the use of preventing skin from being damaged from aging, injuries or the sun, or for restoring skin damaged from same. Moreover, these molecules are generally of interest in the study of connective tissue and attachment molecules and related mechanisms. Fibrillin, fibulin and related molecules are further described in Adams, et al., J. Mol. Biol., 272(2):226-36 (1997); Kielty and Shuttleworth, Microsc. Res. Tech., 38(4):413-27 (1997); and Child, J. Card. Surg., 12(2Supp.):131-5 (1997).

Currently, efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins, particularly secreted proteins which have homology to fibulin and fibrillin. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to fibulin and fibrillin, designated herein as PRO334 polypeptides.

45. PRO346

The widespread occurrence of cancer has prompted the devotion of considerable resources and discovering new treatments of treatment. One particular method involves the creation of tumor or cancer specific monoclonal antibodies (mAbs) which are specific to tumor antigens. Such mAbs, which can distinguish between normal and cancerous cells are useful in the diagnosis, prognosis and treatment of the disease. Particular antigens are known to be associated with neoplastic diseases, such as colorectal and breast cancer. Since colon cancer is a widespread disease, early diagnosis and treatment is an important medical goal. Diagnosis and treatment of cancer can be implemented using monoclonal antibodies (mAbs) specific therefore having fluorescent, nuclear magnetic or radioactive tags. Radioactive genes, toxins and/or drug tagged mAbs can be used for treatment *in situ* with minimal patient description.

Carcinoembryonic antigen (CEA) is a glycoprotein found in human colon cancer and the digestive

organs of a 2-6 month human embryos. CEA is a known human tumor marker and is widely used in the diagnosis of neoplastic diseases, such as colon cancer. For example, when the serum levels of CEA are elevated in a patient, a drop of CEA levels after surgery would indicate the tumor resection was successful. On the other hand, a subsequent rise in serum CEA levels after surgery would indicate that metastases of the original tumor may have formed or that new primary tumors may have appeared. CEA may also be a target for mAb, antisense nucleotides

46. PRO268

Protein disulfide isomerase is an enzymatic protein which is involved in the promotion of correct refolding of proteins through the establishment of correct disulfide bond formation. Protein disulfide isomerase was initially identified based upon its ability to catalyze the renaturation of reduced denatured RNAse (Goldberger et al., *J. Biol. Chem.* 239:1406-1410 (1964) and Epstein et al., *Cold Spring Harbor Symp. Quant. Biol.* 28:439-449 (1963)). Protein disulfide isomerase has been shown to be a resident enzyme of the endoplasmic reticulum which is retained in the endoplasmic reticulum via a -KDEL or -HDEL amino acid sequence at its C-terminus.

Given the importance of disulfide bond-forming enzymes and their potential uses in a number of different applications, for example in increasing the yield of correct refolding of recombinantly produced proteins, efforts are currently being undertaken by both industry and academia to identify new, native proteins having homology to protein disulfide isomerase. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel protein disulfide isomerase homologs. We herein describe a novel polypeptide having homology to protein disulfide isomerase, designated herein as PRO268.

47. PRO330

Prolyl 4-hydroxylase is an enzyme which functions to post-translationally hydroxylate proline residues at the Y position of the amino acid sequence Gly-X-Y, which is a repeating three amino acid sequence found in both collagen and procollagen. Hydroxylation of proline residues at the Y position of the Gly-X-Y amino acid triplet to form 4-hydroxyproline residues at those positions is required before newly synthesized collagen polypeptide chains may fold into their proper three-dimensional triple-helical conformation. If hydroxylation does not occur, synthesized collagen polypeptides remain non-helical, are poorly secreted by cells and cannot assemble into stable functional collagen fibrils. Vuorio et al., *Proc. Natl. Acad. Sci. USA* 89:7467-7470 (1992). Prolyl 4-hydroxylase is comprised of at least two different polypeptide subunits, alpha and beta.

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)]. Based upon these efforts, Applicants have herein identified and describe a novel polypeptide having homology to the alpha subunit of prolyl 4-hydroxylase, designated herein as PRO330.

48. **PRO339 and PRO310**

Fringe is a protein which specifically blocks serrate-mediated activation of notch in the dorsal compartment of the Drosophila wing imaginal disc. Fleming, et al., Development, 124(15):2973-81 (1997). Therefore, fringe is of interest for both its role in development as well as its ability to regulate serrate, particularly serrate's signaling abilities. Also of interest are novel polypeptides which may have a role in development and/or the regulation of serrate-like molecules. Of particular interest are novel polypeptides having homology to fringe as identified and described herein, designated herein as PRO339 and PRO310 polypeptides.

49. **PRO244**

Lectins are a class of proteins comprising a region that binds carbohydrates specifically and non-covalently. Numerous lectins have been identified in higher animals, both membrane-bound and soluble, and have been implicated in a variety of cell-recognition phenomena and tumor metastasis.

Most lectins can be classified as either C-type (calcium-dependent) or S-type (thiol-dependent).

Lectins are thought to play a role in regulating cellular events that are initiated at the level of the plasma membrane. For example, plasma membrane associated molecules are involved in the activation of various subsets of lymphoid cells, e.g. T-lymphocytes, and it is known that cell surface molecules are responsible for activation of these cells and consequently their response during an immune reaction.

A particular group of cell adhesion molecules, selectins, belong in the superfamily of C-type lectins. This group includes L-selectin (peripheral lymph node homing receptor (pnHR), LEC-CAM-1, LAM-1, $gp90^{MEL}$, $gp100^{MEL}$, $gp110^{MEL}$, MEL-14 antigen, Leu-8 antigen, TQ-1 antigen, DREG antigen), E-selectin (LEC-CAM-2, LECAM-2, ELAM-1), and P-selectin (LEC-CAM-3, LECAM-3, GMP-140, PADGEM). The structure of selectins consists of a C-type lectin (carbohydrate binding) domain, an epidermal growth factor-like (EGF-like) motif, and variable numbers of complement regulatory (CR) motifs. Selectins are associated with leukocyte adhesion, e.g. the attachment of neutrophils to venular endothelial cells adjacent to inflammation (E-selectin), or with the trafficking of lymphocytes from blood to secondary lymphoid organs, e.g. lymph nodes and Peyer's patches (L-selectin).

Another exemplary lectin is the cell-associated macrophage antigen, Mac-2 that is believed to be involved in cell adhesion and immune responses. Macrophages also express a lectin that recognizes Tn Ag, a human carcinoma-associated epitope.

Another C-type lectin is CD95 (Fas antigen/APO-1) that is an important mediator of immunologically relevant regulated or programmed cell death (apoptosis). "Apoptosis" is a non-necrotic cell death that takes place in metazoan animal cells following activation of an intrinsic cell suicide program. The cloning of Fas antigen is described in PCT publication WO 91/10448, and European patent application EP510691. The mature Fas molecule consists of 319 amino acids of which 157 are extracellular, 17 constitute the transmembrane domain, and 145 are intracellular. Increased levels of Fas expression at T cell surface have been associated with tumor cells and HIV-infected cells. Ligation of CD95 triggers apoptosis in the presence of interleukin-1 (IL-2).

C-type lectins also include receptors for oxidized low-density lipoprotein (LDL). This suggests a possible role in the pathogenesis of atherosclerosis.

We herein describe the identification and characterization of novel polypeptides having homology to

C-type lectins, designated herein as PRO244 polypeptides.

SUMMARY OF THE INVENTION

1. PRO211 and PRO217

Applicants have identified cDNA clones that encode novel polypeptides having homology to EGF, designated in the present application as "PRO211" and "PRO217" polypeptides.

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO211 or PRO217 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding EGF-like homologue PRO211 and PRO217 polypeptides of Fig. 2 (SEQ ID NO:2) and/or 4 (SEQ ID NO:4) indicated in Fig. 1 (SEQ ID NO1) and/or Fig. 3 (SEQ ID NO:3), respectively, or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO211 and PRO217 EGF-like homologue PRO211 and PRO217 polypeptides. In particular, the invention provides isolated native sequence PRO211 and PRO217 EGF-like homologue polypeptides, which in one embodiment, includes an amino acid sequence comprising residues: 1 to 353 of Fig. 2 (SEQ ID NO:2) or (2) 1 to 379 of Fig. 4 (SEQ ID NO: 4).

2. PRO230

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO230".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO230 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO230 polypeptide having amino acid residues 1 through 467 of Figure 6 (SEQ ID NO:12), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO230 polypeptide. In particular, the invention provides isolated native sequence PRO230 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 through 467 of Figure 6 (SEQ ID NO:12).

In another embodiment, the invention provides an expressed sequence tag (EST) comprising the nucleotide sequence of SEQ ID NO:13 (Figure 7) which is herein designated as DNA20088.

3. PRO232

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO232".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO232 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO232 polypeptide having amino acid residues 1 to 114 of Figure 9 (SEQ ID NO:18), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO232 polypeptide. In particular, the invention provides isolated native sequence PRO232 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 114 of Figure 9 (SEQ ID NO:18).

4. **PRO187**

Applicants have identified a cDNA clone that encodes a novel polypeptide, designated in the present application as "PRO187".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO187 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO187 polypeptide of Figure 11 (SEQ ID NO:23), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides a nucleic acid comprising the coding sequence of Figure 10 (SEQ ID NO:22) or its complement. In another aspect, the invention provides a nucleic acid of the full length protein of clone DNA27864-1155, deposited with the ATCC under accession number ATCC 209375, alternatively the coding sequence of clone DNA27864-1155, deposited under accession number ATCC 209375.

In yet another embodiment, the invention provides isolated PRO187 polypeptide. In particular, the invention provides isolated native sequence PRO187 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 205 of Figure 11 (SEQ ID NO:23). Alternatively, the invention provides a polypeptide encoded by the nucleic acid deposited under accession number ATCC 209375.

5. **PRO265**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO265".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO265 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO265 polypeptide having amino acid residues 1 to 660 of Figure 13 (SEQ ID NO:28), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO265 polypeptide. In particular, the invention provides isolated native sequence PRO265 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 660 of Figure 13 (SEQ ID NO:28). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO265 polypeptide.

6. **PRO219**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO219".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO219 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO219 polypeptide having amino acid residues 1 to 915 of Figure 15 (SEQ ID NO:34), or is complementary to such

encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO219 polypeptide. In particular, the invention provides isolated native sequence PRO219 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 915 of Figure 15 (SEQ ID NO:34).

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7. PRO246

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO246".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO246 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO246 polypeptide having amino acid residues 1 to 390 of Figure 17 (SEQ ID NO:39), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO246 polypeptide. In particular, the invention provides isolated native sequence PRO246 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 390 of Figure 17 (SEQ ID NO:39). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO246 polypeptide.

8. PRO228

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to CD97, EMR1 and latrophilin, wherein the polypeptide is designated in the present application as "PRO228".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO228 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO228 polypeptide having amino acid residues 1 to 690 of Figure 19 (SEQ ID NO:49), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO228 polypeptide. In particular, the invention provides isolated native sequence PRO228 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 690 of Figure 19 (SEQ ID NO:49). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO228 polypeptide.

In another embodiment, the invention provides an expressed sequence tag (EST) comprising the nucleotide sequence of SEQ ID NO:50, designated herein as DNA21951.

9. PRO533

Applicants have identified a cDNA clone (DNA49435-1219) that encodes a novel polypeptide, designated in the present application as PRO533.

In one embodiment, the invention provides an isolated nucleic acid molecule having at least about 80% sequence identity to (a) a DNA molecule encoding a PRO533 polypeptide comprising the sequence of amino

acids 23 to 216 of Figure 22 (SEQ ID NO:59), or (b) the complement of the DNA molecule of (a). The sequence identity preferably is about 85%, more preferably about 90%, most preferably about 95%. In one aspect, the isolated nucleic acid has at least about 80%, preferably at least about 85%, more preferably at least about 90%, and most preferably at least about 95% sequence identity with a polypeptide having amino acid residues 23 to 216 of Figure 22 (SEQ ID NO:59). Preferably, the highest degree of sequence identity occurs within the secreted portion (amino acids 23 to 216 of Figure 22, SEQ ID NO:59). In a further embodiment, the isolated nucleic acid molecule comprises DNA encoding a PRO533 polypeptide having amino acid residues 1 to 216 of Figure 22 (SEQ ID NO:59), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides a nucleic acid of the full length protein of clone DNA49435-1219, deposited with the ATCC under accession number ATCC 209480.

In yet another embodiment, the invention provides isolated PRO533 polypeptide. In particular, the invention provides isolated native sequence PRO533 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 23 to 216 of Figure 22 (SEQ ID NO:59). Native PRO533 polypeptides with or without the native signal sequence (amino acids 1 to 22 in Figure 22 (SEQ ID NO:59)), and with or without the initiating methionine are specifically included. Alternatively, the invention provides a PRO533 polypeptide encoded by the nucleic acid deposited under accession number ATCC 209480.

10. PRO245

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO245".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO245 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO245 polypeptide having amino acid residues 1 to 312 of Fig. 24 (SEQ ID NO:64), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO245 polypeptide. In particular, the invention provides isolated native sequence PRO245 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 312 of Figure 24 (SEQ ID NO:64).

11. PRO220, PRO221 and PRO227

Applicants have identified cDNA clones that each encode novel polypeptides, all having leucine rich repeats. These polypeptides are designated in the present application as PRO220, PRO221 and PRO227.

In one embodiment, the invention provides isolated nucleic acid molecules comprising DNA respectively encoding PRO220, PRO221 and PRO227, respectively. In one aspect, provided herein is an isolated nucleic acid comprises DNA encoding the PRO220 polypeptide having amino acid residues 1 through 708 of Figure 26 (SEQ ID NO:69), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. Also provided herein is an isolated nucleic acid comprises DNA encoding the PRO221 polypeptide having amino acid residues 1 through 259 of

Figure 28 (SEQ ID NO:71), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. Moreover, also provided herein is an isolated nucleic acid comprises DNA encoding the PRO227 polypeptide having amino acid residues 1 through 620 of Figure 30 (SEQ ID NO:73), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO220, PRO221 and PRO227 polypeptides. In particular, provided herein is the isolated native sequence for the PRO220 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 708 of Figure 26 (SEQ ID NO:69). Additionally provided herein is the isolated native sequence for the PRO221 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 259 of Figure 28 (SEQ ID NO:71). Moreover, provided herein is the isolated native sequence for the PRO227 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 620 of Figure 30 (SEQ ID NO:73).

12. **PRO258**

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to CRTAM and poliovirus receptor precursors, wherein the polypeptide is designated in the present application as "PRO258".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO258 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO258 polypeptide having amino acid residues 1 to 398 of Figure 32 (SEQ ID NO:84), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO258 polypeptide. In particular, the invention provides isolated native sequence PRO258 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 398 of Figure 32 (SEQ ID NO:84). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO258 polypeptide.

13. **PRO266**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO266".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO266 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO266 polypeptide having amino acid residues 1 to 696 of Figure 34 (SEQ ID NO:91), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO266 polypeptide. In particular, the invention provides isolated native sequence PRO266 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 696 of Figure 34 (SEQ ID NO:91).

14. **PRO269**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as PRO269.

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO269 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO269 polypeptide having amino acid residues 1 to 490 of Fig. 36 (SEQ ID NO:96), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO269 polypeptide. In particular, the invention provides isolated native sequence PRO269 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 490 of Figure 36 (SEQ ID NO:96). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO269 polypeptide.

15. **PRO287**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO287".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO287 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO287 polypeptide having amino acid residues 1 to 415 of Fig. 38 (SEQ ID NO:104), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO287 polypeptide. In particular, the invention provides isolated native sequence PRO287 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 415 of Figure 38 (SEQ ID NO:104).

16. **PRO214**

Applicants have identified a cDNA clone that encodes a novel polypeptide, designated in the present application as "PRO214".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO214 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO214 polypeptide of Fig. 40 (SEQ ID NO:109), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides a nucleic acid comprising the coding sequence of Fig. 39 (SEQ ID NO:108) or its complement. In another aspect, the invention provides a nucleic acid of the full length protein of clone DNA32286-1191, deposited with ATCC under accession number ATCC 209385.

In yet another embodiment, the invention provides isolated PRO214 polypeptide. In particular, the invention provides isolated native sequence PRO214 polypeptide, which in one embodiment, includes an amino acid sequence comprising the residues of Figure 40 (SEQ ID NO:109). Alternatively, the invention provides a polypeptide encoded by the nucleic acid deposited under accession number ATCC 209385.

17. PRO317

Applicants have identified a cDNA clone that encodes a novel polypeptide, designated in the present application as "PRO317".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding PRO317 polypeptide. In one aspect, the isolated nucleic acid comprises DNA (SEQ ID NO:113) encoding PRO317 polypeptide having amino acid residues 1 to 366 of Fig. 42, or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO317 polypeptide. In particular, the invention provides isolated native-sequence PRO317 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 366 of Figure 42 (SEQ ID NO:114).

In yet another embodiment, the invention supplies a method of detecting the presence of PRO317 in a sample, the method comprising:

a) contacting a detectable anti-PRO317 antibody with a sample suspected of containing PRO317; and

b) detecting binding of the antibody to the sample; wherein the sample is selected from the group consisting of a body fluid, a tissue sample, a cell extract, and a cell culture medium.

In a still further embodiment a method is provided for determining the presence of PRO317 mRNA in a sample, the method comprising:

a) contacting a sample suspected of containing PRO317 mRNA with a detectable nucleic acid probe that hybridizes under moderate to stringent conditions to PRO317 mRNA; and

b) detecting hybridization of the probe to the sample.

Preferably, in this method the sample is a tissue sample and the detecting step is by *in situ* hybridization, or the sample is a cell extract and detection is by Northern analysis.

Further, the invention provides a method for treating a PRO317-associated disorder comprising administering to a mammal an effective amount of the PRO317 polypeptide or a composition thereof containing a carrier, or with an effective amount of a PRO317 agonist or PRO317 antagonist, such as an antibody which binds specifically to PRO317.

18. PRO301

Applicants have identified a cDNA clone (DNA40628-1216) that encodes a novel polypeptide, designated in the present application as "PRO301".

In one embodiment, the invention provides an isolated nucleic acid molecule having at least about 80% sequence identity to (a) a DNA molecule encoding a PRO301 polypeptide comprising the sequence of amino acids 28 to 258 of Fig. 44 (SEQ ID NO:119), or (b) the complement of the DNA molecule of (a). The sequence identity preferably is about 85%, more preferably about 90%, most preferably about 95%. In one aspect, the isolated nucleic acid has at least about 80%, preferably at least about 85%, more preferably at least about 90%, and most preferably at least about 95% sequence identity with a polypeptide having amino acid residues 28 to 258 of Fig. 44 (SEQ ID NO:119). Preferably, the highest degree of sequence identity occurs within the

extracellular domains (amino acids 28 to 258 of Fig. 44, SEQ ID NO:119). In a further embodiment, the isolated nucleic acid molecule comprises DNA encoding a PRO301 polypeptide having amino acid residues 28 to 299 of Fig. 44 (SEQ ID NO:119), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides a nucleic acid of the full length protein of clone DNA40628-1216, deposited with the ATCC under accession number ATCC 209432, alternatively the coding sequence of clone DNA40628-1216, deposited under accession number ATCC 209432.

In yet another embodiment, the invention provides isolated PRO301 polypeptide. In particular, the invention provides isolated native sequence PRO301 polypeptide, which in one embodiment, includes an amino acid sequence comprising the extracellular domain residues 28 to 258 of Figure 44 (SEQ ID NO:119). Native PRO301 polypeptides with or without the native signal sequence (amino acids 1 to 27 in Figure 44 (SEQ ID NO:119), and with or without the initiating methionine are specifically included. Additionally, the sequences of the invention may also comprise the transmembrane domain (residues 236 to about 258 in Figure 44; SEQ ID NO:119) and/or the intracellular domain (about residue 259 to 299 in Figure 44; SEQ ID NO:119). Alternatively, the invention provides a PRO301 polypeptide encoded by the nucleic acid deposited under accession number ATCC 209432.

19. PRO224

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO224".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO224 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO224 polypeptide having amino acid residues 1 to 282 of Figure 46 (SEQ ID NO:127), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO224 polypeptide. In particular, the invention provides isolated native sequence PRO224 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 282 of Figure 46 (SEQ ID NO:127).

20. PRO222

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO222".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO222 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO222 polypeptide having amino acid residues 1 to 490 of Fig. 48 (SEQ ID NO:132), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO222 polypeptide. In particular, the invention provides isolated native sequence PRO222 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 490 of Figure 48 (SEQ ID NO:132).

21. PRO234

Applicants have identified a cDNA clone that encodes a novel lectin polypeptide molecule, designated in the present application as "PRO234".

In one embodiment, the invention provides an isolated nucleic acid encoding a novel lectin comprising DNA encoding a PRO234 polypeptide. In one aspect, the isolated nucleic acid comprises the DNA encoding PRO234 polypeptides having amino acid residues 1 to 382 of Fig. 50 (SEQ ID NO:137), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides an isolated nucleic acid molecule comprising the nucleotide sequence of Fig. 49 (SEQ ID NO:136).

In another embodiment, the invention provides isolated novel PRO234 polypeptides. In particular, the invention provides isolated native sequence PRO234 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 382 of Figure 50 (SEQ ID NO:137).

In yet another embodiment, the invention provides oligonucleotide probes useful for isolating genomic and cDNA nucleotide sequences.

22. PRO231

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to a putative acid phosphatase, wherein the polypeptide is designated in the present application as "PRO231".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO231 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO231 polypeptide having amino acid residues 1 to 428 of Fig. 52 (SEQ ID NO:142), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO231 polypeptide. In particular, the invention provides isolated native sequence PRO231 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 428 of Figure 52 (SEQ ID NO:142).

23. PRO229

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to scavenger receptors wherein the polypeptide is designated in the present application as "PRO229".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO229 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO229 polypeptide having amino acid residues 1 to 347 of Figure 54 (SEQ ID NO:148), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO229 polypeptide. In particular, the invention provides isolated native sequence PRO229 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 347 of Figure 54 (SEQ ID NO:148).

24. PRO238

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to reductase, wherein the polypeptide is designated in the present application as "PRO238".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO238 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO238 polypeptide having amino acid residues 1 to 310 of Figure 56 (SEQ ID NO:153), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO238 polypeptide. In particular, the invention provides isolated native sequence PRO238 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 310 of Figure 56 (SEQ ID NO:153).

25. PRO233

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO233".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO233 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO233 polypeptide having amino acid residues 1 to 300 of Figure 58 (SEQ ID NO:159), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO233 polypeptide. In particular, the invention provides isolated native sequence PRO233 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 300 of Figure 58 (SEQ ID NO:159).

26. PRO223

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to serine carboxypeptidase polypeptides, wherein the polypeptide is designated in the present application as "PRO223".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO223 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO223 polypeptide having amino acid residues 1 to 476 of Figure 60 (SEQ ID NO:164), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO223 polypeptide. In particular, the

invention provides isolated native sequence PRO223 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 476 of Figure 60 (SEQ ID NO:164).

27. PRO235

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO235".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO235 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO235 polypeptide having amino acid residues 1 to 552 of Figure 62 (SEQ ID NO:170), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO235 polypeptide. In particular, the invention provides isolated native sequence PRO235 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 552 of Figure 62 (SEQ ID NO:170).

28. PRO236 and PRO262

Applicants have identified cDNA clones that encode novel polypeptides having homology to β-galactosidase, wherein those polypeptides are designated in the present application as "PRO236" and "PRO262".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO236 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO236 polypeptide having amino acid residues 1 to 636 of Figure 64 (SEQ ID NO:175), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO262 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO262 polypeptide having amino acid residues 1 to 654 of Figure 66 (SEQ ID NO:177), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO236 polypeptide. In particular, the invention provides isolated native sequence PRO236 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 636 of Figure 64 (SEQ ID NO:175).

In another embodiment, the invention provides isolated PRO262 polypeptide. In particular, the invention provides isolated native sequence PRO262 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 654 of Figure 66 (SEQ ID NO:177).

29. PRO239

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO239".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO239 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO239 polypeptide having amino acid residues 1 to 501 of Figure 68 (SEQ ID NO:185), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO239 polypeptide. In particular, the invention provides isolated native sequence PRO239 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 501 of Figure 68 (SEQ ID NO:185).

30. PRO257

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO257".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO257 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO257 polypeptide having amino acid residues 1 to 607 of Figure 70 (SEQ ID NO:190), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO257 polypeptide. In particular, the invention provides isolated native sequence PRO257 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 607 of Figure 70 (SEQ ID NO:190). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO257 polypeptide.

31. PRO260

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO260".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO260 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO260 polypeptide having amino acid residues 1 to 467 of Figure 72 (SEQ ID NO:195), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO260 polypeptide. In particular, the invention provides isolated native sequence PRO260 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 467 of Figure 72 (SEQ ID NO:195).

32. PRO263

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to CD44 antigen, wherein the polypeptide is designated in the present application as "PRO263".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO263 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO263

polypeptide having amino acid residues 1 to 322 of Figure 74 (SEQ ID NO:201), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO263 polypeptide. In particular, the invention provides isolated native sequence PRO263 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 322 of Figure 74 (SEQ ID NO:201). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO263 polypeptide.

33. PRO270

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO270".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO270 polypeptide. In one aspect, the isolated nucleic acid comprises DNA whivch includes the sequence encoding the PRO270 polypeptide having amino acid residues 1 to 296 of Fig. 76 (SEQ ID NO:207), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO270 polypeptide. In particular, the invention provides isolated native sequence PRO270 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 296 of Figure 76 (SEQ ID NO:207).

34. PRO271

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to the proteoglycan link protein, wherein the polypeptide is designated in the present application as "PRO271".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO271 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO271 polypeptide having amino acid residues 1 to 360 of Figure 78 (SEQ ID NO:213), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO271 polypeptide. In particular, the invention provides isolated native sequence PRO271 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 360 of Figure 78 (SEQ ID NO:213).

35. PRO272

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO272".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO272 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO272 polypeptide having amino acid residues 1 to 328 of Figure 80 (SEQ ID NO:221), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under

high stringency conditions.

In another embodiment, the invention provides isolated PRO272 polypeptide. In particular, the invention provides isolated native sequence PRO272 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 328 of Figure 80 (SEQ ID NO:211).

36. PRO294

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO294".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO294 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO294 polypeptide having amino acid residues 1 to 550 of Figure 82 (SEQ ID NO:227), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO294 polypeptide. In particular, the invention provides isolated native sequence PRO294 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 550 of Figure 82 (SEQ ID NO:227).

37. PRO295

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO295".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO295 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO295 polypeptide having amino acid residues 1 to 350 of Figure 84 (SEQ ID NO:236), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO295 polypeptide. In particular, the invention provides isolated native sequence PRO295 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 350 of Figure 84 (SEQ ID NO:236).

38. PRO293

Applicants have identified a cDNA clone that encodes a novel human neuronal leucine rich repeat polypeptide, wherein the polypeptide is designated in the present application as "PRO293".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO293 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO293 polypeptide having amino acid residues 1 to 713 of Figure 86 (SEQ ID NO:245), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO293 polypeptide. In particular, the

invention provides isolated native sequence PRO293 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 713 of Figure 86 (SEQ ID NO:245). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO293 polypeptide.

39. PRO247

Applicants have identified a cDNA clone that encodes a novel polypeptide having leucine rich repeats wherein the polypeptide is designated in the present application as "PRO247".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO247 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO247 polypeptide having amino acid residues 1 to 546 of Figure 88 (SEQ ID NO:250), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO247 polypeptide. In particular, the invention provides isolated native sequence PRO247 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 546 of Figure 88 (SEQ ID NO:250). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO247 polypeptide.

40. PRO302, PRO303, PRO304, PRO307 and PRO343

Applicants have identified cDNA clones that encode novel polypeptides having homology to various proteases, wherein those polypeptide are designated in the present application as "PRO302", "PRO303", "PRO304", "PRO307" and "PRO343" polypeptides.

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO302 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO302 polypeptide having amino acid residues 1 to 452 of Figure 90 (SEQ ID NO:255), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO303 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO303 polypeptide having amino acid residues 1 to 314 of Figure 92 (SEQ ID NO:257), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In yet another embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO304 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO304 polypeptide having amino acid residues 1 to 556 of Figure 94 (SEQ ID NO:259), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO307 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO307 polypeptide having amino acid residues 1 to 383 of Figure 96 (SEQ ID NO:261), or is complementary to such

encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO343 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO343 polypeptide having amino acid residues 1 to 317 of Figure 98 (SEQ ID NO:263), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO302 polypeptide. In particular, the invention provides isolated native sequence PRO302 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 452 of Figure 90 (SEQ ID NO:255).

In another embodiment, the invention provides isolated PRO303 polypeptide. In particular, the invention provides isolated native sequence PRO303 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 314 of Figure 92 (SEQ ID NO:257).

In another embodiment, the invention provides isolated PRO304 polypeptide. In particular, the invention provides isolated native sequence PRO304 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 556 of Figure 94 (SEQ ID NO:259).

In another embodiment, the invention provides isolated PRO307 polypeptide. In particular, the invention provides isolated native sequence PRO307 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 383 of Figure 96 (SEQ ID NO:261).

In another embodiment, the invention provides isolated PRO343 polypeptide. In particular, the invention provides isolated native sequence PRO343 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 317 of Figure 98 (SEQ ID NO:263).

41. PRO328

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO328".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO328 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO328 polypeptide having amino acid residues 1 to 463 of Figure 100 (SEQ ID NO:285), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO328 polypeptide. In particular, the invention provides isolated native sequence PRO328 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 463 of Figure 100 (SEQ ID NO:285). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO306 polypeptide.

42. PRO335, PRO331 and PRO326

Applicants have identified three cDNA clones that respectively encode three novel polypeptides, each having leucine rich repeats and homology to LIG-1 and ALS. These polypeptides are designated in the present application as PRO335, PRO331 and PRO326, respectively.

In one embodiment, the invention provides three isolated nucleic acid molecules comprising DNA respectively encoding PRO335, PRO331 and PRO326, respectively. In one aspect, herein is provided an isolated nucleic acid comprising DNA encoding the PRO335 polypeptide having amino acid residues 1 through 1059 of Figure 102 (SEQ ID NO:290), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. Also provided herein is an isolated nucleic acid comprises DNA encoding the PRO331 polypeptide having amino acid residues 1 through 640 of Figure 104 (SEQ ID NO:292), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. Additionally provided herein is an isolated nucleic acid comprises DNA encoding the PRO326 polypeptide having amino acid residues 1 through 1119 of Figure 106 (SEQ ID NO:294), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO335, PRO331 and PRO326 polypeptides or extracellular domains thereof. In particular, the invention provides isolated native sequence for the PRO335 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 through 1059 of Figure 102 (SEQ ID NO:290). Also provided herein is the isolated native sequence for the PRO331 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 through 640 of Figure 104 (SEQ ID NO:292). Also provided herein is the isolated native sequence for the PRO326 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 through 1119 of Figure 106 (SEQ ID NO:294).

43. PRO332

Applicants have identified a cDNA clone (DNA40982-1235) that encodes a novel polypeptide, designated in the present application as "PRO332."

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA having at least about 80% sequence identity to (a) a DNA molecule encoding a PRO358 polypeptide comprising the sequence of amino acids 49 to 642 of Fig. 108 (SEQ ID NO:310), or (b) the complement of the DNA molecule of (a). The sequence identity preferably is about 85%, more preferably about 90%, most preferably about 95%. In one aspect, the isolated nucleic acid has at least about 80%, preferably at least about 85%, more preferably at least about 90%, and most preferably at least about 95% sequence identity with a polypeptide having amino acid residues 1 to 642 of Fig. 108 (SEQ ID NO:310). Preferably, the highest degree of sequence identity occurs within the leucine-rich repeat domains (amino acids 116 to 624 of Fig. 108, SEQ ID NO:310). In a further embodiment, the isolated nucleic acid molecule comprises DNA encoding a PRO332 polypeptide having amino acid residues 49 to 642 of Fig. 108 (SEQ ID NO:310), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO332 polypeptides. In particular, the invention provides isolated native sequence PRO332 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 49 to 624 of Figure 108 (SEQ ID NO:310). Native PRO332 polypeptides with or without the native signal sequence (amino acids 1 to 48 in Figure 108, SEQ ID NO:310), and with or without the initiating methionine are specifically included.

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44. PRO334

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to fibulin and fibrillin, wherein the polypeptide is designated in the present application as "PRO334".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO334 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO334 polypeptide having amino acid residues 1 to 509 of Figure 110 (SEQ ID NO:315), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO334 polypeptide. In particular, the invention provides isolated native sequence PRO334 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 509 of Figure 110 (SEQ ID NO:315).

45. PRO346

Applicants have identified a cDNA clone (DNA44167-1243) that encodes a novel polypeptide, designated in the present application as "PRO346."

In one embodiment, the invention provides an isolated nucleic acid molecule having at least about 80% sequence identity to (a) a DNA molecule encoding a PRO346 polypeptide comprising the sequence of amino acids 19 to 339 of Fig. 112 (SEQ ID NO: 320), or (b) the complement of the DNA molecule of (a). The sequence identity preferably is about 85%, more preferably about 90%, most preferably about 95%. In one aspect, the isolated nucleic acid has at least about 80%, preferably at least about 85%, more preferably at least about 90%, and most preferably at least about 95% sequence identity with a polypeptide having amino acid residues 19 to 339 of Fig. 112 (SEQ ID NO:320). Preferably, the highest degree of sequence identity occurs within the extracellular domains (amino acids 19 to 339 of Fig. 112, SEQ ID NO:320). In alternative embodiments, the polypeptide by which the homology is measured comprises the residues 1-339, 19-360 or 19-450 of Fig. 112, SEQ ID NO:320). In a further embodiment, the isolated nucleic acid molecule comprises DNA encoding a PRO346 polypeptide having amino acid residues 19 to 339 of Fig. 112 (SEQ ID NO:320), alternatively residues 1-339, 19-360 or 19-450 of Fig. 112 (SEQ ID NO:320) or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides a nucleic acid of the full length protein of clone DNA44167-1243, deposited with the ATCC under accession number ATCC 209434, alternatively the coding sequence of clone DNA44167-1243, deposited under accession number ATCC 209434.

In yet another embodiment, the invention provides isolated PRO346 polypeptide. In particular, the invention provides isolated native sequence PRO346 polypeptide, which in one embodiment, includes an amino

acid sequence comprising residues 19 to 339 of Figure 112 (SEQ ID NO:320). Native PRO346 polypeptides with or without the native signal sequence (residues 1 to 18 in Figure 112 (SEQ ID NO:320), with or without the initiating methionine, with or without the transmembrane domain (residues 340 to 360) and with or without the intracellular domain (residues 361 to 450) are specifically included. Alternatively, the invention provides a PRO346 polypeptide encoded by the nucleic acid deposited under accession number ATCC 209434.

46. PRO268

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to protein disulfide isomerase, wherein the polypeptide is designated in the present application as "PRO268".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO268 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO268 polypeptide having amino acid residues 1 to 280 of Figure 114 (SEQ ID NO:325), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO268 polypeptide. In particular, the invention provides isolated native sequence PRO268 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 280 of Figure 114 (SEQ ID NO:325). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO268 polypeptide.

47. PRO330

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to the alpha subunit of prolyl 4-hydroxylase, wherein the polypeptide is designated in the present application as "PRO330".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO330 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO330 polypeptide having amino acid residues 1 to 533 of Figure 116 (SEQ ID NO:332), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO330 polypeptide. In particular, the invention provides isolated native sequence PRO330 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 533 of Figure 116 (SEQ ID NO:332).

48. PRO339 and PRO310

Applicants have identified two cDNA clones wherein each clone encodes a novel polypeptide having homology to fringe, wherein the polypeptides are designated in the present application as "PRO339" and "PRO310".

In one embodiment, the invention provides isolated nucleic acid molecules comprising DNA encoding a PRO339 and/or a PRO310 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO339 polypeptide having amino acid residues 1 to 772 of Figure 118 (SEQ ID NO:339), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally,

under high stringency conditions. In another aspect, the isolated nucleic acid comprises DNA encoding the PRO310 polypeptide having amino acid residues 1 to 318 of Figure 120 (SEQ ID NO:341), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO339 as well as isolated PRO310 polypeptides. In particular, the invention provides isolated native sequence PRO339 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 772 of Figure 118 (SEQ ID NO:339). The invention further provides isolated native sequence PRO310 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 318 of Figure 120 (SEQ ID NO:341).

49. PRO244

Applicants have identified a cDNA clone that encodes a novel polypeptide, designated in the present application as "PRO244".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding PRO244 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding PRO244 polypeptide having amino acid residues 1 to 219 of Fig. 122 (SEQ ID NO:377), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO244 polypeptide. In particular, the invention provides isolated native sequence PRO244 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 219 of Figure 122 (SEQ ID NO:377).

50. Additional Embodiments

In other embodiments of the present invention, the invention provides vectors comprising DNA encoding any of the herein described polypeptides. Host cell comprising any such vector are also provided. By way of example, the host cells may be CHO cells, *E. coli*, or yeast. A process for producing any of the herein described polypeptides is further provided and comprises culturing host cells under conditions suitable for expression of the desired polypeptide and recovering the desired polypeptide from the cell culture.

In other embodiments, the invention provides chimeric molecules comprising any of the herein described polypeptides fused to a heterologous polypeptide or amino acid sequence. Example of such chimeric molecules comprise any of the herein described polypeptides fused to an epitope tag sequence or a Fc region of an immunoglobulin.

In another embodiment, the invention provides an antibody which specifically binds to any of the above or below described polypeptides. Optionally, the antibody is a monoclonal antibody, humanized antibody, antibody fragment or single-chain antibody.

In yet other embodiments, the invention provides oligonucleotide probes useful for isolating genomic and cDNA nucleotide sequences, wherein those probes may be derived from any of the above or below described nucleotide sequences.

In other embodiments, the invention provides an isolated nucleic acid molecule comprising a nucleotide

sequence that encodes a PRO polypeptide.

In one aspect, the isolated nucleic acid molecule comprises a nucleotide sequence having at least about 80% sequence identity, preferably at least about 81% sequence identity, more preferably at least about 82% sequence identity, yet more preferably at least about 83% sequence identity, yet more preferably at least about 84% sequence identity, yet more preferably at least about 85% sequence identity, yet more preferably at least about 86% sequence identity, yet more preferably at least about 87% sequence identity, yet more preferably at least about 88% sequence identity, yet more preferably at least about 89% sequence identity, yet more preferably at least about 90% sequence identity, yet more preferably at least about 91% sequence identity, yet more preferably at least about 92% sequence identity, yet more preferably at least about 93% sequence identity, yet more preferably at least about 94% sequence identity, yet more preferably at least about 95% sequence identity, yet more preferably at least about 96% sequence identity, yet more preferably at least about 97% sequence identity, yet more preferably at least about 98% sequence identity and yet more preferably at least about 99% sequence identity to (a) a DNA molecule encoding a PRO polypeptide having a full-length amino acid sequence as disclosed herein, an amino acid sequence lacking the signal peptide as disclosed herein or an extracellular domain of a transmembrane protein, with or without the signal peptide, as disclosed herein, or (b) the complement of the DNA molecule of (a).

In other aspects, the isolated nucleic acid molecule comprises a nucleotide sequence having at least about 80% sequence identity, preferably at least about 81% sequence identity, more preferably at least about 82% sequence identity, yet more preferably at least about 83% sequence identity, yet more preferably at least about 84% sequence identity, yet more preferably at least about 85% sequence identity, yet more preferably at least about 86% sequence identity, yet more preferably at least about 87% sequence identity, yet more preferably at least about 88% sequence identity, yet more preferably at least about 89% sequence identity, yet more preferably at least about 90% sequence identity, yet more preferably at least about 91% sequence identity, yet more preferably at least about 92% sequence identity, yet more preferably at least about 93% sequence identity, yet more preferably at least about 94% sequence identity, yet more preferably at least about 95% sequence identity, yet more preferably at least about 96% sequence identity, yet more preferably at least about 97% sequence identity, yet more preferably at least about 98% sequence identity and yet more preferably at least about 99% sequence identity to (a) a DNA molecule comprising the coding sequence of a full-length PRO polypeptide cDNA as disclosed herein, the coding sequence of a PRO polypeptide lacking the signal peptide as disclosed herein or the coding sequence of an extracellular domain of a transmembrane PRO polypeptide, with or without the signal peptide, as disclosed herein, or (b) the complement of the DNA molecule of (a).

In a further aspect, the invention concerns an isolated nucleic acid molecule comprising a nucleotide sequence having at least about 80% sequence identity, preferably at least about 81% sequence identity, more preferably at least about 82% sequence identity, yet more preferably at least about 83% sequence identity, yet more preferably at least about 84% sequence identity, yet more preferably at least about 85% sequence identity, yet more preferably at least about 86% sequence identity, yet more preferably at least about 87% sequence identity, yet more preferably at least about 88% sequence identity, yet more preferably at least about 89% sequence identity, yet more preferably at least about 90% sequence identity, yet more preferably at least about 91% sequence identity, yet more preferably at least about 92% sequence identity, yet more preferably at least

about 93% sequence identity, yet more preferably at least about 94% sequence identity, yet more preferably at least about 95% sequence identity, yet more preferably at least about 96% sequence identity, yet more preferably at least about 97% sequence identity, yet more preferably at least about 98% sequence identity and yet more preferably at least about 99% sequence identity to (a) a DNA molecule that encodes the same mature polypeptide encoded by any of the human protein cDNAs deposited with the ATCC as disclosed herein, or (b) the complement of the DNA molecule of (a).

Another aspect the invention provides an isolated nucleic acid molecule comprising a nucleotide sequence encoding a PRO polypeptide which is either transmembrane domain-deleted or transmembrane domain-inactivated, or is complementary to such encoding nucleotide sequence, wherein the transmembrane domain(s) of such polypeptide are disclosed herein. Therefore, soluble extracellular domains of the herein described PRO polypeptides are contemplated.

Another embodiment is directed to fragments of a PRO polypeptide coding sequence, or the complement thereof, that may find use as, for example, hybridization probes or for encoding fragments of a PRO polypeptide that may optionally encode a polypeptide comprising a binding site for an anti-PRO antibody. Such nucleic acid fragments are usually at least about 20 nucleotides in length, preferably at least about 30 nucleotides in length, more preferably at least about 40 nucleotides in length, yet more preferably at least about 50 nucleotides in length, yet more preferably at least about 60 nucleotides in length, yet more preferably at least about 70 nucleotides in length, yet more preferably at least about 80 nucleotides in length, yet more preferably at least about 90 nucleotides in length, yet more preferably at least about 100 nucleotides in length, yet more preferably at least about 110 nucleotides in length, yet more preferably at least about 120 nucleotides in length, yet more preferably at least about 130 nucleotides in length, yet more preferably at least about 140 nucleotides in length, yet more preferably at least about 150 nucleotides in length, yet more preferably at least about 160 nucleotides in length, yet more preferably at least about 170 nucleotides in length, yet more preferably at least about 180 nucleotides in length, yet more preferably at least about 190 nucleotides in length, yet more preferably at least about 200 nucleotides in length, yet more preferably at least about 250 nucleotides in length, yet more preferably at least about 300 nucleotides in length, yet more preferably at least about 350 nucleotides in length, yet more preferably at least about 400 nucleotides in length, yet more preferably at least about 450 nucleotides in length, yet more preferably at least about 500 nucleotides in length, yet more preferably at least about 600 nucleotides in length, yet more preferably at least about 700 nucleotides in length, yet more preferably at least about 800 nucleotides in length, yet more preferably at least about 900 nucleotides in length and yet more preferably at least about 1000 nucleotides in length, wherein in this context the term "about" means the referenced nucleotide sequence length plus or minus 10% of that referenced length. It is noted that novel fragments of a PRO polypeptide-encoding nucleotide sequence may be determined in a routine manner by aligning the PRO polypeptide-encoding nucleotide sequence with other known nucleotide sequences using any of a number of well known sequence alignment programs and determining which PRO polypeptide-encoding nucleotide sequence fragment(s) are novel. All of such PRO polypeptide-encoding nucleotide sequences are contemplated herein. Also contemplated are the PRO polypeptide fragments encoded by these nucleotide molecule fragments, preferably those PRO polypeptide fragments that comprise a binding site for an anti-PRO antibody.

In another embodiment, the invention provides isolated PRO polypeptide encoded by any of the isolated

nucleic acid sequences hereinabove identified.

In a certain aspect, the invention concerns an isolated PRO polypeptide, comprising an amino acid sequence having at least about 80% sequence identity, preferably at least about 81% sequence identity, more preferably at least about 82% sequence identity, yet more preferably at least about 83% sequence identity, yet more preferably at least about 84% sequence identity, yet more preferably at least about 85% sequence identity, yet more preferably at least about 86% sequence identity, yet more preferably at least about 87% sequence identity, yet more preferably at least about 88% sequence identity, yet more preferably at least about 89% sequence identity, yet more preferably at least about 90% sequence identity, yet more preferably at least about 91% sequence identity, yet more preferably at least about 92% sequence identity, yet more preferably at least about 93% sequence identity, yet more preferably at least about 94% sequence identity, yet more preferably at least about 95% sequence identity, yet more preferably at least about 96% sequence identity, yet more preferably at least about 97% sequence identity, yet more preferably at least about 98% sequence identity and yet more preferably at least about 99% sequence identity to a PRO polypeptide having a full-length amino acid sequence as disclosed herein, an amino acid sequence lacking the signal peptide as disclosed herein or an extracellular domain of a transmembrane protein, with or without the signal peptide, as disclosed herein.

In a further aspect, the invention concerns an isolated PRO polypeptide comprising an amino acid sequence having at least about 80% sequence identity, preferably at least about 81% sequence identity, more preferably at least about 82% sequence identity, yet more preferably at least about 83% sequence identity, yet more preferably at least about 84% sequence identity, yet more preferably at least about 85% sequence identity, yet more preferably at least about 86% sequence identity, yet more preferably at least about 87% sequence identity, yet more preferably at least about 88% sequence identity, yet more preferably at least about 89% sequence identity, yet more preferably at least about 90% sequence identity, yet more preferably at least about 91% sequence identity, yet more preferably at least about 92% sequence identity, yet more preferably at least about 93% sequence identity, yet more preferably at least about 94% sequence identity, yet more preferably at least about 95% sequence identity, yet more preferably at least about 96% sequence identity, yet more preferably at least about 97% sequence identity, yet more preferably at least about 98% sequence identity and yet more preferably at least about 99% sequence identity to an amino acid sequence encoded by any of the human protein cDNAs deposited with the ATCC as disclosed herein.

In a further aspect, the invention concerns an isolated PRO polypeptide comprising an amino acid sequence scoring at least about 80% positives, preferably at least about 81% positives, more preferably at least about 82% positives, yet more preferably at least about 83% positives, yet more preferably at least about 84% positives, yet more preferably at least about 85% positives, yet more preferably at least about 86% positives, yet more preferably at least about 87% positives, yet more preferably at least about 88% positives, yet more preferably at least about 89% positives, yet more preferably at least about 90% positives, yet more preferably at least about 91% positives, yet more preferably at least about 92% positives, yet more preferably at least about 93% positives, yet more preferably at least about 94% positives, yet more preferably at least about 95% positives, yet more preferably at least about 96% positives, yet more preferably at least about 97% positives, yet more preferably at least about 98% positives and yet more preferably at least about 99% positives when compared with the amino acid sequence of a PRO polypeptide having a full-length amino acid sequence as

disclosed herein, an amino acid sequence lacking the signal peptide as disclosed herein or an extracellular domain of a transmembrane protein, with or without the signal peptide, as disclosed herein.

In a specific aspect, the invention provides an isolated PRO polypeptide without the N-terminal signal sequence and/or the initiating methionine and is encoded by a nucleotide sequence that encodes such an amino acid sequence as hereinbefore described. Processes for producing the same are also herein described, wherein those processes comprise culturing a host cell comprising a vector which comprises the appropriate encoding nucleic acid molecule under conditions suitable for expression of the PRO polypeptide and recovering the PRO polypeptide from the cell culture.

Another aspect the invention provides an isolated PRO polypeptide which is either transmembrane domain-deleted or transmembrane domain-inactivated. Processes for producing the same are also herein described, wherein those processes comprise culturing a host cell comprising a vector which comprises the appropriate encoding nucleic acid molecule under conditions suitable for expression of the PRO polypeptide and recovering the PRO polypeptide from the cell culture.

In yet another embodiment, the invention concerns agonists and antagonists of a native PRO polypeptide as defined herein. In a particular embodiment, the agonist or antagonist is an anti-PRO antibody or a small molecule.

In a further embodiment, the invention concerns a method of identifying agonists or antagonists to a PRO polypeptide which comprise contacting the PRO polypeptide with a candidate molecule and monitoring a biological activity mediated by said PRO polypeptide. Preferably, the PRO polypeptide is a native PRO polypeptide.

In a still further embodiment, the invention concerns a composition of matter comprising a PRO polypeptide, or an agonist or antagonist of a PRO polypeptide as herein described, or an anti-PRO antibody, in combination with a carrier. Optionally, the carrier is a pharmaceutically acceptable carrier.

Another embodiment of the present invention is directed to the use of a PRO polypeptide, or an agonist or antagonist thereof as hereinbefore described, or an anti-PRO antibody, for the preparation of a medicament useful in the treatment of a condition which is responsive to the PRO polypeptide, an agonist or antagonist thereof or an anti-PRO antibody.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows a nucleotide sequence (SEQ ID NO:1) of a native sequence PRO211 cDNA, wherein SEQ ID NO:1 is a clone designated herein as "DNA32292-1131".

Figure 2 shows the amino acid sequence (SEQ ID NO:2) derived from the coding sequence of SEQ ID NO:1 shown in Figure 1.

Figure 3 shows a nucleotide sequence (SEQ ID NO:3) of a native sequence PRO217 cDNA, wherein SEQ ID NO:3 is a clone designated herein as "DNA33094-1131".

Figure 4 shows the amino acid sequence (SEQ ID NO:4) derived from the coding sequence of SEQ ID NO:3 shown in Figure 3.

Figure 5 shows a nucleotide sequence (SEQ ID NO:11) of a native sequence PRO230 cDNA, wherein SEQ ID NO:11 is a clone designated herein as "DNA33223-1136".

Figure 6 shows the amino acid sequence (SEQ ID NO:12) derived from the coding sequence of SEQ ID NO:11 shown in Figure 5.

Figure 7 shows a nucleotide sequence designated herein as DNA20088 (SEQ ID NO:13).

Figure 8 shows a nucleotide sequence (SEQ ID NO:17) of a native sequence PRO232 cDNA, wherein SEQ ID NO:17 is a clone designated herein as "DNA34435-1140".

Figure 9 shows the amino acid sequence (SEQ ID NO:18) derived from the coding sequence of SEQ ID NO:17 shown in Figure 8.

Figure 10 shows a nucleotide sequence (SEQ ID NO:22) of a native sequence PRO187 cDNA, wherein SEQ ID NO:22 is a clone designated herein as "DNA27864-1155".

Figure 11 shows the amino acid sequence (SEQ ID NO:23) derived from the coding sequence of SEQ ID NO:22 shown in Figure 10.

Figure 12 shows a nucleotide sequence (SEQ ID NO:27) of a native sequence PRO265 cDNA, wherein SEQ ID NO:27 is a clone designated herein as "DNA36350-1158".

Figure 13 shows the amino acid sequence (SEQ ID NO:28) derived from the coding sequence of SEQ ID NO:27 shown in Figure 12.

Figure 14 shows a nucleotide sequence (SEQ ID NO:33) of a native sequence PRO219 cDNA, wherein SEQ ID NO:33 is a clone designated herein as "DNA32290-1164".

Figure 15 shows the amino acid sequence (SEQ ID NO:34) derived from the coding sequence of SEQ ID NO:33 shown in Figure 14.

Figure 16 shows a nucleotide sequence (SEQ ID NO:38) of a native sequence PRO246 cDNA, wherein SEQ ID NO:38 is a clone designated herein as "DNA35639-1172".

Figure 17 shows the amino acid sequence (SEQ ID NO:39) derived from the coding sequence of SEQ ID NO:38 shown in Figure 16.

Figure 18 shows a nucleotide sequence (SEQ ID NO:48) of a native sequence PRO228 cDNA, wherein SEQ ID NO:48 is a clone designated herein as "DNA33092-1202".

Figure 19 shows the amino acid sequence (SEQ ID NO:49) derived from the coding sequence of SEQ ID NO:48 shown in Figure 18.

Figure 20 shows a nucleotide sequence designated herein as DNA21951 (SEQ ID NO:50).

Figure 21 shows a nucleotide sequence (SEQ ID NO:58) of a native sequence PRO533 cDNA, wherein SEQ ID NO:58 is a clone designated herein as "DNA49435-1219".

Figure 22 shows the amino acid sequence (SEQ ID NO:59) derived from the coding sequence of SEQ ID NO:58 shown in Figure 21.

Figure 23 shows a nucleotide sequence (SEQ ID NO:63) of a native sequence PRO245 cDNA, wherein SEQ ID NO:63 is a clone designated herein as "DNA35638-1141".

Figure 24 shows the amino acid sequence (SEQ ID NO:64) derived from the coding sequence of SEQ ID NO:63 shown in Figure 23.

Figure 25 shows a nucleotide sequence (SEQ ID NO:68) of a native sequence PRO220 cDNA, wherein SEQ ID NO:68 is a clone designated herein as "DNA32298-1132".

Figure 26 shows the amino acid sequence (SEQ ID NO:69) derived from the coding sequence of SEQ

ID NO:68 shown in Figure 25.

Figure 27 shows a nucleotide sequence (SEQ ID NO:70) of a native sequence PRO221 cDNA, wherein SEQ ID NO:70 is a clone designated herein as "DNA33089-1132".

Figure 28 shows the amino acid sequence (SEQ ID NO:71) derived from the coding sequence of SEQ ID NO:70 shown in Figure 27.

Figure 29 shows a nucleotide sequence (SEQ ID NO:72) of a native sequence PRO227 cDNA, wherein SEQ ID NO:72 is a clone designated herein as "DNA33786-1132".

Figure 30 shows the amino acid sequence (SEQ ID NO:73) derived from the coding sequence of SEQ ID NO:72 shown in Figure 29.

Figure 31 shows a nucleotide sequence (SEQ ID NO:83) of a native sequence PRO258 cDNA, wherein SEQ ID NO:83 is a clone designated herein as "DNA35918-1174".

Figure 32 shows the amino acid sequence (SEQ ID NO:84) derived from the coding sequence of SEQ ID NO:83 shown in Figure 31.

Figure 33 shows a nucleotide sequence (SEQ ID NO:90) of a native sequence PRO266 cDNA, wherein SEQ ID NO:90 is a clone designated herein as "DNA37150-1178".

Figure 34 shows the amino acid sequence (SEQ ID NO:91) derived from the coding sequence of SEQ ID NO:90 shown in Figure 33.

Figure 35 shows a nucleotide sequence (SEQ ID NO:95) of a native sequence PRO269 cDNA, wherein SEQ ID NO:95 is a clone designated herein as "DNA38260-1180".

Figure 36 shows the amino acid sequence (SEQ ID NO:96) derived from the coding sequence of SEQ ID NO:95 shown in Figure 35.

Figure 37 shows a nucleotide sequence (SEQ ID NO:103) of a native sequence PRO287 cDNA, wherein SEQ ID NO:103 is a clone designated herein as "DNA39969-1185".

Figure 38 shows the amino acid sequence (SEQ ID NO:104) derived from the coding sequence of SEQ ID NO:103 shown in Figure 37.

Figure 39 shows a nucleotide sequence (SEQ ID NO:108) of a native sequence PRO214 cDNA, wherein SEQ ID NO:108 is a clone designated herein as "DNA32286-1191".

Figure 40 shows the amino acid sequence (SEQ ID NO:109) derived from the coding sequence of SEQ ID NO:108 shown in Figure 39.

Figure 41 shows a nucleotide sequence (SEQ ID NO:113) of a native sequence PRO317 cDNA, wherein SEQ ID NO:113 is a clone designated herein as "DNA33461-1199".

Figure 42 shows the amino acid sequence (SEQ ID NO:114) derived from the coding sequence of SEQ ID NO:113 shown in Figure 41.

Figure 43 shows a nucleotide sequence (SEQ ID NO:118) of a native sequence PRO301 cDNA, wherein SEQ ID NO:118 is a clone designated herein as "DNA40628-1216".

Figure 44 shows the amino acid sequence (SEQ ID NO:119) derived from the coding sequence of SEQ ID NO:118 shown in Figure 43.

Figure 45 shows a nucleotide sequence (SEQ ID NO:126) of a native sequence PRO224 cDNA, wherein

SEQ ID NO:126 is a clone designated herein as "DNA33221-1133".

Figure 46 shows the amino acid sequence (SEQ ID NO:127) derived from the coding sequence of SEQ ID NO:126 shown in Figure 45.

Figure 47 shows a nucleotide sequence (SEQ ID NO:131) of a native sequence PRO222 cDNA, wherein SEQ ID NO:131 is a clone designated herein as "DNA33107-1135".

Figure 48 shows the amino acid sequence (SEQ ID NO:132) derived from the coding sequence of SEQ ID NO:131 shown in Figure 47.

Figure 49 shows a nucleotide sequence (SEQ ID NO:136) of a native sequence PRO234 cDNA, wherein SEQ ID NO:136 is a clone designated herein as "DNA35557-1137".

Figure 50 shows the amino acid sequence (SEQ ID NO:137) derived from the coding sequence of SEQ ID NO:136 shown in Figure 49.

Figure 51 shows a nucleotide sequence (SEQ ID NO:141) of a native sequence PRO231 cDNA, wherein SEQ ID NO:141 is a clone designated herein as "DNA34434-1139".

Figure 52 shows the amino acid sequence (SEQ ID NO:142) derived from the coding sequence of SEQ ID NO:141 shown in Figure 51.

Figure 53 shows a nucleotide sequence (SEQ ID NO:147) of a native sequence PRO229 cDNA, wherein SEQ ID NO:147 is a clone designated herein as "DNA33100-1159".

Figure 54 shows the amino acid sequence (SEQ ID NO:148) derived from the coding sequence of SEQ ID NO:147 shown in Figure 53.

Figure 55 shows a nucleotide sequence (SEQ ID NO:152) of a native sequence PRO238 cDNA, wherein SEQ ID NO:152 is a clone designated herein as "DNA35600-1162".

Figure 56 shows the amino acid sequence (SEQ ID NO:153) derived from the coding sequence of SEQ ID NO:152 shown in Figure 55.

Figure 57 shows a nucleotide sequence (SEQ ID NO:158) of a native sequence PRO233 cDNA, wherein SEQ ID NO:158 is a clone designated herein as "DNA34436-1238".

Figure 58 shows the amino acid sequence (SEQ ID NO:159) derived from the coding sequence of SEQ ID NO:158 shown in Figure 57.

Figure 59 shows a nucleotide sequence (SEQ ID NO:163) of a native sequence PRO223 cDNA, wherein SEQ ID NO:163 is a clone designated herein as "DNA33206-1165".

Figure 60 shows the amino acid sequence (SEQ ID NO:164) derived from the coding sequence of SEQ ID NO:163 shown in Figure 59.

Figure 61 shows a nucleotide sequence (SEQ ID NO:169) of a native sequence PRO235 cDNA, wherein SEQ ID NO:169 is a clone designated herein as "DNA35558-1167".

Figure 62 shows the amino acid sequence (SEQ ID NO:170) derived from the coding sequence of SEQ ID NO:169 shown in Figure 61.

Figure 63 shows a nucleotide sequence (SEQ ID NO:174) of a native sequence PRO236 cDNA, wherein SEQ ID NO:174 is a clone designated herein as "DNA35599-1168".

Figure 64 shows the amino acid sequence (SEQ ID NO:175) derived from the coding sequence of SEQ ID NO:174 shown in Figure 63.

Figure 65 shows a nucleotide sequence (SEQ ID NO:176) of a native sequence PRO262 cDNA, wherein SEQ ID NO:176 is a clone designated herein as "DNA36992-1168".

Figure 66 shows the amino acid sequence (SEQ ID NO:177) derived from the coding sequence of SEQ ID NO:176 shown in Figure 65.

Figure 67 shows a nucleotide sequence (SEQ ID NO:184) of a native sequence PRO239 cDNA, wherein SEQ ID NO:184 is a clone designated herein as "DNA34407-1169".

Figure 68 shows the amino acid sequence (SEQ ID NO:185) derived from the coding sequence of SEQ ID NO:184 shown in Figure 67.

Figure 69 shows a nucleotide sequence (SEQ ID NO:189) of a native sequence PRO257 cDNA, wherein SEQ ID NO:189 is a clone designated herein as "DNA35841-1173".

Figure 70 shows the amino acid sequence (SEQ ID NO:190) derived from the coding sequence of SEQ ID NO:189 shown in Figure 69.

Figure 71 shows a nucleotide sequence (SEQ ID NO:194) of a native sequence PRO260 cDNA, wherein SEQ ID NO:194 is a clone designated herein as "DNA33470-1175".

Figure 72 shows the amino acid sequence (SEQ ID NO:195) derived from the coding sequence of SEQ ID NO:194 shown in Figure 71.

Figure 73 shows a nucleotide sequence (SEQ ID NO:200) of a native sequence PRO263 cDNA, wherein SEQ ID NO:200 is a clone designated herein as "DNA34431-1177".

Figure 74 shows the amino acid sequence (SEQ ID NO:201) derived from the coding sequence of SEQ ID NO:200 shown in Figure 73.

Figure 75 shows a nucleotide sequence (SEQ ID NO:206) of a native sequence PRO270 cDNA, wherein SEQ ID NO:206 is a clone designated herein as "DNA39510-1181".

Figure 76 shows the amino acid sequence (SEQ ID NO:207) derived from the coding sequence of SEQ ID NO:206 shown in Figure 75.

Figure 77 shows a nucleotide sequence (SEQ ID NO:212) of a native sequence PRO271 cDNA, wherein SEQ ID NO:212 is a clone designated herein as "DNA39423-1182".

Figure 78 shows the amino acid sequence (SEQ ID NO:213) derived from the coding sequence of SEQ ID NO:212 shown in Figure 77.

Figure 79 shows a nucleotide sequence (SEQ ID NO:220) of a native sequence PRO272 cDNA, wherein SEQ ID NO:220 is a clone designated herein as "DNA40620-1183".

Figure 80 shows the amino acid sequence (SEQ ID NO:221) derived from the coding sequence of SEQ ID NO:220 shown in Figure 79.

Figure 81 shows a nucleotide sequence (SEQ ID NO:226) of a native sequence PRO294 cDNA, wherein SEQ ID NO:226 is a clone designated herein as "DNA40604-1187".

Figure 82 shows the amino acid sequence (SEQ ID NO:227) derived from the coding sequence of SEQ ID NO:226 shown in Figure 81.

Figure 83 shows a nucleotide sequence (SEQ ID NO:235) of a native sequence PRO295 cDNA, wherein SEQ ID NO:235 is a clone designated herein as "DNA38268-1188".

Figure 84 shows the amino acid sequence (SEQ ID NO:236) derived from the coding sequence of SEQ ID NO:235 shown in Figure 83.

Figure 85 shows a nucleotide sequence (SEQ ID NO:244) of a native sequence PRO293 cDNA, wherein SEQ ID NO:244 is a clone designated herein as "DNA37151-1193".

Figure 86 shows the amino acid sequence (SEQ ID NO:245) derived from the coding sequence of SEQ ID NO:244 shown in Figure 85.

Figure 87 shows a nucleotide sequence (SEQ ID NO:249) of a native sequence PRO247 cDNA, wherein SEQ ID NO:249 is a clone designated herein as "DNA35673-1201".

Figure 88 shows the amino acid sequence (SEQ ID NO:250) derived from the coding sequence of SEQ ID NO:249 shown in Figure 87.

Figure 89 shows a nucleotide sequence (SEQ ID NO:254) of a native sequence PRO302 cDNA, wherein SEQ ID NO:254 is a clone designated herein as "DNA40370-1217".

Figure 90 shows the amino acid sequence (SEQ ID NO:255) derived from the coding sequence of SEQ ID NO:254 shown in Figure 89.

Figure 91 shows a nucleotide sequence (SEQ ID NO:256) of a native sequence PRO303 cDNA, wherein SEQ ID NO:256 is a clone designated herein as "DNA42551-1217".

Figure 92 shows the amino acid sequence (SEQ ID NO:257) derived from the coding sequence of SEQ ID NO:256 shown in Figure 91.

Figure 93 shows a nucleotide sequence (SEQ ID NO:258) of a native sequence PRO304 cDNA, wherein SEQ ID NO:258 is a clone designated herein as "DNA39520-1217".

Figure 94 shows the amino acid sequence (SEQ ID NO:259) derived from the coding sequence of SEQ ID NO:258 shown in Figure 93.

Figure 95 shows a nucleotide sequence (SEQ ID NO:260) of a native sequence PRO307 cDNA, wherein SEQ ID NO:260 is a clone designated herein as "DNA41225-1217".

Figure 96 shows the amino acid sequence (SEQ ID NO:261) derived from the coding sequence of SEQ ID NO:260 shown in Figure 95.

Figure 97 shows a nucleotide sequence (SEQ ID NO:262) of a native sequence PRO343 cDNA, wherein SEQ ID NO:262 is a clone designated herein as "DNA43318-1217".

Figure 98 shows the amino acid sequence (SEQ ID NO:263) derived from the coding sequence of SEQ ID NO:262 shown in Figure 97.

Figure 99 shows a nucleotide sequence (SEQ ID NO:284) of a native sequence PRO328 cDNA, wherein SEQ ID NO:284 is a clone designated herein as "DNA40587-1231".

Figure 100 shows the amino acid sequence (SEQ ID NO:285) derived from the coding sequence of SEQ ID NO:284 shown in Figure 99.

Figure 101 shows a nucleotide sequence (SEQ ID NO:289) of a native sequence PRO335 cDNA, wherein SEQ ID NO:289 is a clone designated herein as "DNA41388-1234".

Figure 102 shows the amino acid sequence (SEQ ID NO:290) derived from the coding sequence of SEQ ID NO:289 shown in Figure 101.

Figure 103 shows a nucleotide sequence (SEQ ID NO:291) of a native sequence PRO331 cDNA,

wherein SEQ ID NO:291 is a clone designated herein as "DNA40981-1234".

Figure 104 shows the amino acid sequence (SEQ ID NO:292) derived from the coding sequence of SEQ ID NO:291 shown in Figure 103.

Figure 105 shows a nucleotide sequence (SEQ ID NO:293) of a native sequence PRO326 cDNA, wherein SEQ ID NO:293 is a clone designated herein as "DNA37140-1234".

Figure 106 shows the amino acid sequence (SEQ ID NO:294) derived from the coding sequence of SEQ ID NO:293 shown in Figure 105.

Figure 107 shows a nucleotide sequence (SEQ ID NO:309) of a native sequence PRO332 cDNA, wherein SEQ ID NO:309 is a clone designated herein as "DNA40982-1235".

Figure 108 shows the amino acid sequence (SEQ ID NO:310) derived from the coding sequence of SEQ ID NO:309 shown in Figure 107.

Figure 109 shows a nucleotide sequence (SEQ ID NO:314) of a native sequence PRO334 cDNA, wherein SEQ ID NO:314 is a clone designated herein as "DNA41379-1236".

Figure 110 shows the amino acid sequence (SEQ ID NO:315) derived from the coding sequence of SEQ ID NO:314 shown in Figure 109.

Figure 111 shows a nucleotide sequence (SEQ ID NO:319) of a native sequence PRO346 cDNA, wherein SEQ ID NO:319 is a clone designated herein as "DNA44167-1243".

Figure 112 shows the amino acid sequence (SEQ ID NO:320) derived from the coding sequence of SEQ ID NO:319 shown in Figure 111.

Figure 113 shows a nucleotide sequence (SEQ ID NO:324) of a native sequence PRO268 cDNA, wherein SEQ ID NO:324 is a clone designated herein as "DNA39427-1179".

Figure 114 shows the amino acid sequence (SEQ ID NO:325) derived from the coding sequence of SEQ ID NO:324 shown in Figure 113.

Figure 115 shows a nucleotide sequence (SEQ ID NO:331) of a native sequence PRO330 cDNA, wherein SEQ ID NO:331 is a clone designated herein as "DNA40603-1232".

Figure 116 shows the amino acid sequence (SEQ ID NO:332) derived from the coding sequence of SEQ ID NO:331 shown in Figure 115.

Figure 117 shows a nucleotide sequence (SEQ ID NO:338) of a native sequence PRO339 cDNA, wherein SEQ ID NO:338 is a clone designated herein as "DNA43466-1225".

Figure 118 shows the amino acid sequence (SEQ ID NO:339) derived from the coding sequence of SEQ ID NO:338 shown in Figure 117.

Figure 119 shows a nucleotide sequence (SEQ ID NO:340) of a native sequence PRO310 cDNA, wherein SEQ ID NO:340 is a clone designated herein as "DNA43046-1225".

Figure 120 shows the amino acid sequence (SEQ ID NO:341) derived from the coding sequence of SEQ ID NO:340 shown in Figure 119.

Figure 121 shows a nucleotide sequence (SEQ ID NO:376) of a native sequence PRO244 cDNA, wherein SEQ ID NO:376 is a clone designated herein as "DNA35668-1171".

Figure 122 shows the amino acid sequence (SEQ ID NO:377) derived from the coding sequence of SEQ ID NO:376 shown in Figure 121.

Figure 123 shows a nucleotide sequence (SEQ ID NO:422) of a native sequence PRO1868 cDNA, wherein SEQ ID NO:422 is a clone designated herein as "DNA77624-2515".

Figure 124 shows the amino acid sequence (SEQ ID NO:423) derived from the coding sequence of SEQ ID NO:422 shown in Figure 123.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

I. Definitions

The terms "PRO polypeptide" and "PRO" as used herein and when immediately followed by a numerical designation refer to various polypeptides, wherein the complete designation (i.e., PRO/number) refers to specific polypeptide sequences as described herein. The terms "PRO/number polypeptide" and "PRO/number" wherein the term "number" is provided as an actual numerical designation as used herein encompass native sequence polypeptides and polypeptide variants (which are further defined herein). The PRO polypeptides described herein may be isolated from a variety of sources, such as from human tissue types or from another source, or prepared by recombinant or synthetic methods.

A "native sequence PRO polypeptide" comprises a polypeptide having the same amino acid sequence as the corresponding PRO polypeptide derived from nature. Such native sequence PRO polypeptides can be isolated from nature or can be produced by recombinant or synthetic means. The term "native sequence PRO polypeptide" specifically encompasses naturally-occurring truncated or secreted forms of the specific PRO polypeptide (*e.g.*, an extracellular domain sequence), naturally-occurring variant forms (*e.g.*, alternatively spliced forms) and naturally-occurring allelic variants of the polypeptide. In various embodiments of the invention, the native sequence PRO polypeptides disclosed herein are mature or full-length native sequence polypeptides comprising the full-length amino acids sequences shown in the accompanying figures. Start and stop codons are shown in bold font and underlined in the figures. However, while the PRO polypeptide disclosed in the accompanying figures are shown to begin with methionine residues designated herein as amino acid position 1 in the figures, it is conceivable and possible that other methionine residues located either upstream or downstream from the amino acid position 1 in the figures may be employed as the starting amino acid residue for the PRO polypeptides.

The PRO polypeptide "extracellular domain" or "ECD" refers to a form of the PRO polypeptide which is essentially free of the transmembrane and cytoplasmic domains. Ordinarily, a PRO polypeptide ECD will have less than 1% of such transmembrane and/or cytoplasmic domains and preferably, will have less than 0.5% of such domains. It will be understood that any transmembrane domains identified for the PRO polypeptides of the present invention are identified pursuant to criteria routinely employed in the art for identifying that type of hydrophobic domain. The exact boundaries of a transmembrane domain may vary but most likely by no more than about 5 amino acids at either end of the domain as initially identified herein. Optionally, therefore, an extracellular domain of a PRO polypeptide may contain from about 5 or fewer amino acids on either side of the transmembrane domain/extracellular domain boundary as identified in the Examples or specification and such polypeptides, with or without the associated signal peptide, and nucleic acid encoding them, are contemplated by the present invention.

The approximate location of the "signal peptides" of the various PRO polypeptides disclosed herein are shown in the present specification and/or the accompanying figures. It is noted, however, that the C-terminal boundary of a signal peptide may vary, but most likely by no more than about 5 amino acids on either side of the signal peptide C-terminal boundary as initially identified herein, wherein the C-terminal boundary of the signal peptide may be identified pursuant to criteria routinely employed in the art for identifying that type of amino acid sequence element (e.g., Nielsen et al., Prot. Eng. 10:1-6 (1997) and von Heinje et al., Nucl. Acids.

Res. 14:4683-4690 (1986)). Moreover, it is also recognized that, in some cases, cleavage of a signal sequence from a secreted polypeptide is not entirely uniform, resulting in more than one secreted species. These mature polypeptides, where the signal peptide is cleaved within no more than about 5 amino acids on either side of the C-terminal boundary of the signal peptide as identified herein, and the polynucleotides encoding them, are contemplated by the present invention.

"PRO polypeptide variant" means an active PRO polypeptide as defined above or below having at least about 80% amino acid sequence identity with a full-length native sequence PRO polypeptide sequence as disclosed herein, a PRO polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a PRO polypeptide, with or without the signal peptide, as disclosed herein or any other fragment of a full-length PRO polypeptide sequence as disclosed herein. Such PRO polypeptide variants include, for instance, PRO polypeptides wherein one or more amino acid residues are added, or deleted, at the N- or C-terminus of the full-length native amino acid sequence. Ordinarily, a PRO polypeptide variant will have at least about 80% amino acid sequence identity, preferably at least about 81% amino acid sequence identity, more preferably at least about 82% amino acid sequence identity, more preferably at least about 83% amino acid sequence identity, more preferably at least about 84% amino acid sequence identity, more preferably at least about 85% amino acid sequence identity, more preferably at least about 86% amino acid sequence identity, more preferably at least about 87% amino acid sequence identity, more preferably at least about 88% amino acid sequence identity, more preferably at least about 89% amino acid sequence identity, more preferably at least about 90% amino acid sequence identity, more preferably at least about 91% amino acid sequence identity, more preferably at least about 92% amino acid sequence identity, more preferably at least about 93% amino acid sequence identity, more preferably at least about 94% amino acid sequence identity, more preferably at least about 95% amino acid sequence identity, more preferably at least about 96% amino acid sequence identity, more preferably at least about 97% amino acid sequence identity, more preferably at least about 98% amino acid sequence identity and most preferably at least about 99% amino acid sequence identity with a full-length native sequence PRO polypeptide sequence as disclosed herein, a PRO polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a PRO polypeptide, with or without the signal peptide, as disclosed herein or any other specifically defined fragment of a full-length PRO polypeptide sequence as disclosed herein. Ordinarily, PRO variant polypeptides are at least about 10 amino acids in length, often at least about 20 amino acids in length, more often at least about 30 amino acids in length, more often at least about 40 amino acids in length, more often at least about 50 amino acids in length, more often at least about 60 amino acids in length, more often at least about 70 amino acids in length, more often at least about 80 amino acids in length, more often at least about 90 amino acids in length, more often at least about 100 amino acids in length, more often at least about 150 amino acids in length, more often at least about 200 amino acids in length, more often at least about 300 amino acids in length, or more.

"Percent (%) amino acid sequence identity" with respect to the PRO polypeptide sequences identified herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the specific PRO polypeptide sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid

sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared. For purposes herein, however, % amino acid sequence identity values are generated using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in Table 1 below. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc. and the source code shown in Table 1 below has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available through Genentech, Inc., South San Francisco, California or may be compiled from the source code provided in Table 1 below. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison parameters are set by the ALIGN-2 program and do not vary.

In situations where ALIGN-2 is employed for amino acid sequence comparisons, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) is calculated as follows:

100 times the fraction X/Y

where X is the number of amino acid residues scored as identical matches by the sequence alignment program ALIGN-2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A. As examples of % amino acid sequence identity calculations using this method, Tables 2 and 3 demonstrate how to calculate the % amino acid sequence identity of the amino acid sequence designated "Comparison Protein" to the amino acid sequence designated "PRO", wherein "PRO" represents the amino acid sequence of a hypothetical PRO polypeptide of interest, "Comparison Protein" represents the amino acid sequence of a polypeptide against which the "PRO" polypeptide of interest is being compared, and "X, "Y" and "Z" each represent different hypothetical amino acid residues.

Unless specifically stated otherwise, all % amino acid sequence identity values used herein are obtained as described in the immediately preceding paragraph using the ALIGN-2 computer program. However, % amino acid sequence identity values may also be obtained as described below by using the WU-BLAST-2 computer program (Altschul et al., Methods in Enzymology 266:460-480 (1996)). Most of the WU-BLAST-2 search parameters are set to the default values. Those not set to default values, i.e., the adjustable parameters, are set with the following values: overlap span = 1, overlap fraction = 0.125, word threshold (T) = 11, and scoring matrix = BLOSUM62. When WU-BLAST-2 is employed, a % amino acid sequence identity value is determined by dividing (a) the number of matching identical amino acid residues between the amino acid sequence of the PRO polypeptide of interest having a sequence derived from the native PRO polypeptide and

the comparison amino acid sequence of interest (i.e., the sequence against which the PRO polypeptide of interest is being compared which may be a PRO variant polypeptide) as determined by WU-BLAST-2 by (b) the total number of amino acid residues of the PRO polypeptide of interest. For example, in the statement "a polypeptide comprising an the amino acid sequence A which has or having at least 80% amino acid sequence identity to the amino acid sequence B", the amino acid sequence A is the comparison amino acid sequence of interest and the amino acid sequence B is the amino acid sequence of the PRO polypeptide of interest.

Percent amino acid sequence identity may also be determined using the sequence comparison program NCBI-BLAST2 (Altschul et al., Nucleic Acids Res. 25:3389-3402 (1997)). The NCBI-BLAST2 sequence comparison program may be downloaded from http://www.ncbi.nlm.nih.gov. NCBI-BLAST2 uses several search parameters, wherein all of those search parameters are set to default values including, for example, unmask = yes, strand = all, expected occurrences = 10, minimum low complexity length = 15/5, multi-pass e-value = 0.01, constant for multi-pass = 25, dropoff for final gapped alignment = 25 and scoring matrix = BLOSUM62.

In situations where NCBI-BLAST2 is employed for amino acid sequence comparisons, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) is calculated as follows:

100 times the fraction X/Y

where X is the number of amino acid residues scored as identical matches by the sequence alignment program NCBI-BLAST2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A.

"PRO variant polynucleotide" or "PRO variant nucleic acid sequence" means a nucleic acid molecule which encodes an active PRO polypeptide as defined below and which has at least about 80% nucleic acid sequence identity with a nucleotide acid sequence encoding a full-length native sequence PRO polypeptide sequence as disclosed herein, a full-length native sequence PRO polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a PRO polypeptide, with or without the signal peptide, as disclosed herein or any other fragment of a full-length PRO polypeptide sequence as disclosed herein. Ordinarily, a PRO variant polynucleotide will have at least about 80% nucleic acid sequence identity, more preferably at least about 81% nucleic acid sequence identity, more preferably at least about 82% nucleic acid sequence identity, more preferably at least about 83% nucleic acid sequence identity, more preferably at least about 84% nucleic acid sequence identity, more preferably at least about 85% nucleic acid sequence identity, more preferably at least about 86% nucleic acid sequence identity, more preferably at least about 87% nucleic acid sequence identity, more preferably at least about 88% nucleic acid sequence identity, more preferably at least about 89% nucleic acid sequence identity, more preferably at least about 90% nucleic acid sequence identity, more preferably at least about 91% nucleic acid sequence identity, more preferably at least about 92%

nucleic acid sequence identity, more preferably at least about 93% nucleic acid sequence identity, more preferably at least about 94% nucleic acid sequence identity, more preferably at least about 95% nucleic acid sequence identity, more preferably at least about 96% nucleic acid sequence identity, more preferably at least about 97% nucleic acid sequence identity, more preferably at least about 98% nucleic acid sequence identity and yet more preferably at least about 99% nucleic acid sequence identity with a nucleic acid sequence encoding a full-length native sequence PRO polypeptide sequence as disclosed herein, a full-length native sequence PRO polypeptide sequence lacking the signal peptide as disclosed herein, an extracellular domain of a PRO polypeptide, with or without the signal sequence, as disclosed herein or any other fragment of a full-length PRO polypeptide sequence as disclosed herein. Variants do not encompass the native nucleotide sequence.

Ordinarily, PRO variant polynucleotides are at least about 30 nucleotides in length, often at least about 60 nucleotides in length, more often at least about 90 nucleotides in length, more often at least about 120 nucleotides in length, more often at least about 150 nucleotides in length, more often at least about 180 nucleotides in length, more often at least about 210 nucleotides in length, more often at least about 240 nucleotides in length, more often at least about 270 nucleotides in length, more often at least about 300 nucleotides in length, more often at least about 450 nucleotides in length, more often at least about 600 nucleotides in length, more often at least about 900 nucleotides in length, or more.

"Percent (%) nucleic acid sequence identity" with respect to PRO-encoding nucleic acid sequences identified herein is defined as the percentage of nucleotides in a candidate sequence that are identical with the nucleotides in the PRO nucleic acid sequence of interest, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining percent nucleic acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN or Megalign (DNASTAR) software. For purposes herein, however, % nucleic acid sequence identity values are generated using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in Table 1 below. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc. and the source code shown in Table 1 below has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available through Genentech, Inc., South San Francisco, California or may be compiled from the source code provided in Table 1 below. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison parameters are set by the ALIGN-2 program and do not vary.

In situations where ALIGN-2 is employed for nucleic acid sequence comparisons, the % nucleic acid sequence identity of a given nucleic acid sequence C to, with, or against a given nucleic acid sequence D (which can alternatively be phrased as a given nucleic acid sequence C that has or comprises a certain % nucleic acid sequence identity to, with, or against a given nucleic acid sequence D) is calculated as follows:

100 times the fraction W/Z

where W is the number of nucleotides scored as identical matches by the sequence alignment program ALIGN-2

in that program's alignment of C and D, and where Z is the total number of nucleotides in D. It will be appreciated that where the length of nucleic acid sequence C is not equal to the length of nucleic acid sequence D, the % nucleic acid sequence identity of C to D will not equal the % nucleic acid sequence identity of D to C. As examples of % nucleic acid sequence identity calculations, Tables 4 and 5, demonstrate how to calculate the % nucleic acid sequence identity of the nucleic acid sequence designated "Comparison DNA" to the nucleic acid sequence designated "PRO-DNA", wherein "PRO-DNA" represents a hypothetical PRO-encoding nucleic acid sequence of interest, "Comparison DNA" represents the nucleotide sequence of a nucleic acid molecule against which the "PRO-DNA" nucleic acid molecule of interest is being compared, and "N", "L" and "V" each represent different hypothetical nucleotides.

Unless specifically stated otherwise, all % nucleic acid sequence identity values used herein are obtained as described in the immediately preceding paragraph using the ALIGN-2 computer program. However, % nucleic acid sequence identity values may also be obtained as described below by using the WU-BLAST-2 computer program (Altschul et al., Methods in Enzymology 266:460-480 (1996)). Most of the WU-BLAST-2 search parameters are set to the default values. Those not set to default values, i.e., the adjustable parameters, are set with the following values: overlap span = 1, overlap fraction = 0.125, word threshold (T) = 11, and scoring matrix = BLOSUM62. When WU-BLAST-2 is employed, a % nucleic acid sequence identity value is determined by dividing (a) the number of matching identical nucleotides between the nucleic acid sequence of the PRO polypeptide-encoding nucleic acid molecule of interest having a sequence derived from the native sequence PRO polypeptide-encoding nucleic acid and the comparison nucleic acid molecule of interest (i.e., the sequence against which the PRO polypeptide-encoding nucleic acid molecule of interest is being compared which may be a variant PRO polynucleotide) as determined by WU-BLAST-2 by (b) the total number of nucleotides of the PRO polypeptide-encoding nucleic acid molecule of interest. For example, in the statement "an isolated nucleic acid molecule comprising a nucleic acid sequence A which has or having at least 80% nucleic acid sequence identity to the nucleic acid sequence B", the nucleic acid sequence A is the comparison nucleic acid molecule of interest and the nucleic acid sequence B is the nucleic acid sequence of the PRO polypeptide-encoding nucleic acid molecule of interest.

Percent nucleic acid sequence identity may also be determined using the sequence comparison program NCBI-BLAST2 (Altschul et al., Nucleic Acids Res. 25:3389-3402 (1997)). The NCBI-BLAST2 sequence comparison program may be downloaded from http://www.ncbi.nlm.nih.gov. NCBI-BLAST2 uses several search parameters, wherein all of those search parameters are set to default values including, for example, unmask = yes, strand = all, expected occurrences = 10, minimum low complexity length = 15/5, multi-pass e-value = 0.01, constant for multi-pass = 25, dropoff for final gapped alignment = 25 and scoring matrix = BLOSUM62.

In situations where NCBI-BLAST2 is employed for sequence comparisons, the % nucleic acid sequence identity of a given nucleic acid sequence C to, with, or against a given nucleic acid sequence D (which can alternatively be phrased as a given nucleic acid sequence C that has or comprises a certain % nucleic acid sequence identity to, with, or against a given nucleic acid sequence D) is calculated as follows:

100 times the fraction W/Z

where W is the number of nucleotides scored as identical matches by the sequence alignment program NCBI-BLAST2 in that program's alignment of C and D, and where Z is the total number of nucleotides in D. It will be appreciated that where the length of nucleic acid sequence C is not equal to the length of nucleic acid sequence D, the % nucleic acid sequence identity of C to D will not equal the % nucleic acid sequence identity of D to C.

In other embodiments, PRO variant polynucleotides are nucleic acid molecules that encode an active PRO polypeptide and which are capable of hybridizing, preferably under stringent hybridization and wash conditions, to nucleotide sequences encoding a full-length PRO polypeptide as disclosed herein. PRO variant polypeptides may be those that are encoded by a PRO variant polynucleotide.

The term "positives", in the context of sequence comparison performed as described above, includes residues in the sequences compared that are not identical but have similar properties (e.g. as a result of conservative substitutions, see Table 6 below). For purposes herein, the % value of positives is determined by dividing (a) the number of amino acid residues scoring a positive value between the PRO polypeptide amino acid sequence of interest having a sequence derived from the native PRO polypeptide sequence and the comparison amino acid sequence of interest (i.e., the amino acid sequence against which the PRO polypeptide sequence is being compared) as determined in the BLOSUM62 matrix of WU-BLAST-2 by (b) the total number of amino acid residues of the PRO polypeptide of interest.

Unless specifically stated otherwise, the % value of positives is calculated as described in the immediately preceding paragraph. However, in the context of the amino acid sequence identity comparisons performed as described for ALIGN-2 and NCBI-BLAST-2 above, includes amino acid residues in the sequences compared that are not only identical, but also those that have similar properties. Amino acid residues that score a positive value to an amino acid residue of interest are those that are either identical to the amino acid residue of interest or are a preferred substitution (as defined in Table 6 below) of the amino acid residue of interest.

For amino acid sequence comparisons using ALIGN-2 or NCBI-BLAST2, the % value of positives of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % positives to, with, or against a given amino acid sequence B) is calculated as follows:

100 times the fraction X/Y

where X is the number of amino acid residues scoring a positive value as defined above by the sequence alignment program ALIGN-2 or NCBI-BLAST2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % positives of A to B will not equal the % positives of B to A.

"Isolated," when used to describe the various polypeptides disclosed herein, means polypeptide that has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous

solutes. In preferred embodiments, the polypeptide will be purified (1) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (2) to homogeneity by SDS-PAGE under non-reducing or reducing conditions using Coomassie blue or, preferably, silver stain. Isolated polypeptide includes polypeptide *in situ* within recombinant cells, since at least one component of the PRO polypeptide natural environment will not be present. Ordinarily, however, isolated polypeptide will be prepared by at least one purification step.

An "isolated" PRO polypeptide-encoding nucleic acid or other polypeptide-encoding nucleic acid is a nucleic acid molecule that is identified and separated from at least one contaminant nucleic acid molecule with which it is ordinarily associated in the natural source of the polypeptide-encoding nucleic acid. An isolated polypeptide-encoding nucleic acid molecule is other than in the form or setting in which it is found in nature. Isolated polypeptide-encoding nucleic acid molecules therefore are distinguished from the specific polypeptide-encoding nucleic acid molecule as it exists in natural cells. However, an isolated polypeptide-encoding nucleic acid molecule includes polypeptide-encoding nucleic acid molecules contained in cells that ordinarily express the polypeptide where, for example, the nucleic acid molecule is in a chromosomal location different from that of natural cells.

The term "control sequences" refers to DNA sequences necessary for the expression of an operably linked coding sequence in a particular host organism. The control sequences that are suitable for prokaryotes, for example, include a promoter, optionally an operator sequence, and a ribosome binding site. Eukaryotic cells are known to utilize promoters, polyadenylation signals, and enhancers.

Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

The term "antibody" is used in the broadest sense and specifically covers, for example, single anti-PRO monoclonal antibodies (including agonist, antagonist, and neutralizing antibodies), anti-PRO antibody compositions with polyepitopic specificity, single chain anti-PRO antibodies, and fragments of anti-PRO antibodies (see below). The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally-occurring mutations that may be present in minor amounts.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends on the ability of denatured DNA to reanneal when complementary strands are present in an environment below their melting temperature. The higher the degree

of desired homology between the probe and hybridizable sequence, the higher the relative temperature which can be used. As a result, it follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel et al., Current Protocols in Molecular Biology, Wiley Interscience Publishers, (1995).

"Stringent conditions" or "high stringency conditions", as defined herein, may be identified by those that: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42°C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 μg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) and 50% formamide at 55°C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C.

"Moderately stringent conditions" may be identified as described by Sambrook et al., Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 1989, and include the use of washing solution and hybridization conditions (e.g., temperature, ionic strength and %SDS) less stringent that those described above. An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising: 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/ml denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc. as necessary to accommodate factors such as probe length and the like.

The term "epitope tagged" when used herein refers to a chimeric polypeptide comprising a PRO polypeptide fused to a "tag polypeptide". The tag polypeptide has enough residues to provide an epitope against which an antibody can be made, yet is short enough such that it does not interfere with activity of the polypeptide to which it is fused. The tag polypeptide preferably also is fairly unique so that the antibody does not substantially cross-react with other epitopes. Suitable tag polypeptides generally have at least six amino acid residues and usually between about 8 and 50 amino acid residues (preferably, between about 10 and 20 amino acid residues).

As used herein, the term "immunoadhesin" designates antibody-like molecules which combine the binding specificity of a heterologous protein (an "adhesin") with the effector functions of immunoglobulin constant domains. Structurally, the immunoadhesins comprise a fusion of an amino acid sequence with the desired binding specificity which is other than the antigen recognition and binding site of an antibody (i.e., is "heterologous"), and an immunoglobulin constant domain sequence. The adhesin part of an immunoadhesin molecule typically is a contiguous amino acid sequence comprising at least the binding site of a receptor or a ligand. The immunoglobulin constant domain sequence in the immunoadhesin may be obtained from any immunoglobulin, such as IgG-1, IgG-2, IgG-3, or IgG-4 subtypes, IgA (including IgA-1 and IgA-2), IgE, IgD or IgM.

"Active" or "activity" for the purposes herein refers to form(s) of a PRO polypeptide which retain a biological and/or an immunological activity of native or naturally-occurring PRO, wherein "biological" activity refers to a biological function (either inhibitory or stimulatory) caused by a native or naturally-occurring PRO other than the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring PRO and an "immunological" activity refers to the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring PRO.

The term "antagonist" is used in the broadest sense, and includes any molecule that partially or fully blocks, inhibits, or neutralizes a biological activity of a native PRO polypeptide disclosed herein. In a similar manner, the term "agonist" is used in the broadest sense and includes any molecule that mimics a biological activity of a native PRO polypeptide disclosed herein. Suitable agonist or antagonist molecules specifically include agonist or antagonist antibodies or antibody fragments, fragments or amino acid sequence variants of native PRO polypeptides, peptides, antisense oligonucleotides, small organic molecules, etc. Methods for identifying agonists or antagonists of a PRO polypeptide may comprise contacting a PRO polypeptide with a candidate agonist or antagonist molecule and measuring a detectable change in one or more biological activities normally associated with the PRO polypeptide.

"Treatment" refers to both therapeutic treatment and prophylactic or preventative measures, wherein the object is to prevent or slow down (lessen) the targeted pathologic condition or disorder. Those in need of treatment include those already with the disorder as well as those prone to have the disorder or those in whom the disorder is to be prevented.

"Chronic" administration refers to administration of the agent(s) in a continuous mode as opposed to an acute mode, so as to maintain the initial therapeutic effect (activity) for an extended period of time. "Intermittent" administration is treatment that is not consecutively done without interruption, but rather is cyclic in nature.

"Mammal" for purposes of treatment refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, cats, cattle, horses, sheep, pigs, goats, rabbits, etc. Preferably, the mammal is human.

Administration "in combination with" one or more further therapeutic agents includes simultaneous (concurrent) and consecutive administration in any order.

"Carriers" as used herein include pharmaceutically acceptable carriers, excipients, or stabilizers which are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the physiologically acceptable carrier is an aqueous pH buffered solution. Examples of physiologically acceptable carriers include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptide; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN™, polyethylene glycol (PEG), and PLURONICS™.

"Antibody fragments" comprise a portion of an intact antibody, preferably the antigen binding or

variable region of the intact antibody. Examples of antibody fragments include Fab, Fab', $F(ab')_2$, and Fv fragments; diabodies; linear antibodies (Zapata et al., Protein Eng. 8(10): 1057-1062 [1995]); single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site, and a residual "Fc" fragment, a designation reflecting the ability to crystallize readily. Pepsin treatment yields an $F(ab')_2$ fragment that has two antigen-combining sites and is still capable of cross-linking antigen.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and -binding site. This region consists of a dimer of one heavy- and one light-chain variable domain in tight, non-covalent association. It is in this configuration that the three CDRs of each variable domain interact to define an antigen-binding site on the surface of the V_H-V_L dimer. Collectively, the six CDRs confer antigen-binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab fragments differ from Fab' fragments by the addition of a few residues at the carboxy terminus of the heavy chain CH1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. $F(ab')_2$ antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

The "light chains" of antibodies (immunoglobulins) from any vertebrate species can be assigned to one of two clearly distinct types, called kappa and lambda, based on the amino acid sequences of their constant domains.

Depending on the amino acid sequence of the constant domain of their heavy chains, immunoglobulins can be assigned to different classes. There are five major classes of immunoglobulins: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), e.g., IgG1, IgG2, IgG3, IgG4, IgA, and IgA2.

"Single-chain Fv" or "sFv" antibody fragments comprise the V_H and V_L domains of antibody, wherein these domains are present in a single polypeptide chain. Preferably, the Fv polypeptide further comprises a polypeptide linker between the V_H and V_L domains which enables the sFv to form the desired structure for antigen binding. For a review of sFv, see Pluckthun in The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenburg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994).

The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy-chain variable domain (V_H) connected to a light-chain variable domain (V_L) in the same polypeptide chain (V_H-V_L). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger et al., Proc. Natl. Acad. Sci. USA, 90:6444-6448 (1993).

An "isolated" antibody is one which has been identified and separated and/or recovered from a

component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the antibody will be purified (1) to greater than 95% by weight of antibody as determined by the Lowry method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue or, preferably, silver stain. Isolated antibody includes the antibody in situ within recombinant cells since at least one component of the antibody's natural environment will not be present. Ordinarily, however, isolated antibody will be prepared by at least one purification step.

The word "label" when used herein refers to a detectable compound or composition which is conjugated directly or indirectly to the antibody so as to generate a "labeled" antibody. The label may be detectable by itself (e.g. radioisotope labels or fluorescent labels) or, in the case of an enzymatic label, may catalyze chemical alteration of a substrate compound or composition which is detectable.

By "solid phase" is meant a non-aqueous matrix to which the antibody of the present invention can adhere. Examples of solid phases encompassed herein include those formed partially or entirely of glass (e.g., controlled pore glass), polysaccharides (e.g., agarose), polyacrylamides, polystyrene, polyvinyl alcohol and silicones. In certain embodiments, depending on the context, the solid phase can comprise the well of an assay plate; in others it is a purification column (e.g., an affinity chromatography column). This term also includes a discontinuous solid phase of discrete particles, such as those described in U.S. Patent No. 4,275,149.

A "liposome" is a small vesicle composed of various types of lipids, phospholipids and/or surfactant which is useful for delivery of a drug (such as a PRO polypeptide or antibody thereto) to a mammal. The components of the liposome are commonly arranged in a bilayer formation, similar to the lipid arrangement of biological membranes.

A "small molecule" is defined herein to have a molecular weight below about 500 Daltons.

"PRO317-associated disorder" refers to a pathological condition or disease wherein PRO317 is over- or underexpressed. Such disorders include diseases of the female genital tract or of the endometrium of a mammal, including hyperplasia, endometritis, endometriosis, wherein the patient is at risk for infertility due to endometrial factor, endometrioma, and endometrial cancer, especially those diseases involving abnormal bleeding such as a gynecological disease. They also include diseases involving angiogenesis, wherein the angiogenesis results in a pathological condition, such as cancer involving solid tumors (the therapy for the disorder would result in decreased vascularization and a decline in growth and metastasis of a variety of tumors). Alternatively, the angiogenesis may be beneficial, such as for ischemia, especially coronary ischemia. Hence, these disorders include those found in patients whose hearts are functioning but who have a blocked blood supply due to atherosclerotic coronary artery disease, and those with a functioning but underperfused heart, including patients with coronary arterial disease who are not optimal candidates for angioplasty and coronary artery by-pass surgery. The disorders also include diseases involving the kidney or originating from the kidney tissue, such as polycystic kidney disease and chronic and acute renal failure.

Table 1

```
/*
 *
 * C-C increased from 12 to 15
 * Z is average of EQ
 * B is average of ND
 * match with stop is _M; stop-stop = 0; J (joker) match = 0
 */
#define  _M      -8      /* value of a match with a stop */

int      _day[26][26] = {
/*      A  B  C  D  E  F  G  H  I  J  K  L  M  N  O  P  Q  R  S  T  U  V  W  X  Y  Z */
/* A */    { 2, 0,-2, 0, 0,-4, 1,-1,-1, 0,-1,-2,-1, 0,_M, 1, 0,-2, 1, 1, 0, 0,-6, 0,-3, 0},
/* B */    { 0, 3,-4, 3, 2,-5, 0, 1,-2, 0, 0,-3,-2, 2,_M,-1, 1, 0, 0, 0, 0,-2,-5, 0,-3, 1},
/* C */    {-2,-4,15,-5,-5,-4,-3,-3,-2, 0,-5,-6,-5,-4,_M,-3,-5,-4, 0,-2, 0,-2,-8, 0, 0,-5},
/* D */    { 0, 3,-5, 4, 3,-6, 1, 1,-2, 0, 0,-4,-3, 2,_M,-1, 2,-1, 0, 0, 0,-2,-7, 0,-4, 2},
/* E */    { 0, 2,-5, 3, 4,-5, 0, 1,-2, 0, 0,-3,-2, 1,_M,-1, 2,-1, 0, 0, 0,-2,-7, 0,-4, 3},
/* F */    {-4,-5,-4,-6,-5, 9,-5,-2, 1, 0,-5, 2, 0,-4,_M,-5,-5,-4,-3,-3, 0,-1, 0, 0, 7,-5},
/* G */    { 1, 0,-3, 1, 0,-5, 5,-2,-3, 0,-2,-4,-3, 0,_M,-1,-1,-3, 1, 0, 0,-1,-7, 0,-5, 0},
/* H */    {-1, 1,-3, 1, 1,-2,-2, 6,-2, 0, 0,-2,-2, 2,_M, 0, 3, 2,-1,-1, 0,-2,-3, 0, 0, 2},
/* I */    {-1,-2,-2,-2,-2, 1,-3,-2, 5, 0,-2, 2, 2,-2,_M,-2,-2,-2,-1, 0, 0, 4,-5, 0,-1,-2},
/* J */    { 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,_M, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0},
/* K */    {-1, 0,-5, 0, 0,-5,-2, 0,-2, 0, 5,-3, 0, 1,_M,-1, 1, 3, 0, 0, 0,-2,-3, 0,-4, 0},
/* L */    {-2,-3,-6,-4,-3, 2,-4,-2, 2, 0,-3, 6, 4,-3,_M,-3,-2,-3,-3,-1, 0, 2,-2, 0,-1,-2},
/* M */    {-1,-2,-5,-3,-2, 0,-3,-2, 2, 0, 0, 4, 6,-2,_M,-2,-1, 0,-2,-1, 0, 2,-4, 0,-2,-1},
/* N */    { 0, 2,-4, 2, 1,-4, 0, 2,-2, 0, 1,-3,-2, 2,_M,-1, 1, 0, 1, 0, 0,-2,-4, 0,-2, 1},
/* O */    {_M,_M,_M,_M,_M,_M,_M,_M,_M,_M,_M,_M,_M,_M, 0,_M,_M,_M,_M,_M,_M,_M,_M,_M,_M,_M},
/* P */    { 1,-1,-3,-1,-1,-5,-1, 0,-2, 0,-1,-3,-2,-1,_M, 6, 0, 0, 1, 0, 0,-1,-6, 0,-5, 0},
/* Q */    { 0, 1,-5, 2, 2,-5,-1, 3,-2, 0, 1,-2,-1, 1,_M, 0, 4, 1,-1,-1, 0,-2,-5, 0,-4, 3},
/* R */    {-2, 0,-4,-1,-1,-4,-3, 2,-2, 0, 3,-3, 0, 0,_M, 0, 1, 6, 0,-1, 0,-2, 2, 0,-4, 0},
/* S */    { 1, 0, 0, 0, 0,-3, 1,-1,-1, 0, 0,-3,-2, 1,_M, 1,-1, 0, 2, 1, 0,-1,-2, 0,-3, 0},
/* T */    { 1, 0,-2, 0, 0,-3, 0,-1, 0, 0, 0,-1,-1, 0,_M, 0,-1,-1, 1, 3, 0, 0,-5, 0,-3, 0},
/* U */    { 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,_M, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0},
/* V */    { 0,-2,-2,-2,-2,-1,-1,-2, 4, 0,-2, 2, 2,-2,_M,-1,-2,-2,-1, 0, 0, 4,-6, 0,-2,-2},
/* W */    {-6,-5,-8,-7,-7, 0,-7,-3,-5, 0,-3,-2,-4,-4,_M,-6,-5, 2,-2,-5, 0,-6,17, 0, 0,-6},
/* X */    { 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,_M, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0},
/* Y */    {-3,-3, 0,-4,-4, 7,-5, 0,-1, 0,-4,-1,-2,-2,_M,-5,-4,-4,-3,-3, 0,-2, 0, 0,10,-4},
/* Z */    { 0, 1,-5, 2, 3,-5, 0, 2,-2, 0, 0,-2,-1, 1,_M, 0, 3, 0, 0, 0, 0,-2,-6, 0,-4, 4}
};
```

Table 1 (cont')

```
/*
 */
#include <stdio.h>
#include <ctype.h>

#define  MAXJMP      16      /* max jumps in a diag */
#define  MAXGAP      24      /* don't continue to penalize gaps larger than this */
#define  JMPS        1024    /* max jmps in an path */
#define  MX          4       /* save if there's at least MX-1 bases since last jmp */

#define  DMAT        3       /* value of matching bases */
#define  DMIS        0       /* penalty for mismatched bases */
#define  DINS0       8       /* penalty for a gap */
#define  DINS1       1       /* penalty per base */
#define  PINS0       8       /* penalty for a gap */
#define  PINS1       4       /* penalty per residue */

struct jmp {
        short           n[MAXJMP];      /* size of jmp (neg for dely) */
        unsigned short  x[MAXJMP];      /* base no. of jmp in seq x */
};                                      /* limits seq to 2^16 -1 */

struct diag {
        int             score;          /* score at last jmp */
        long            offset;         /* offset of prev block */
        short           ijmp;           /* current jmp index */
        struct jmp      jp;             /* list of jmps */
};

struct path {
        int     spc;            /* number of leading spaces */
        short   n[JMPS];        /* size of jmp (gap) */
        int     x[JMPS];        /* loc of jmp (last elem before gap) */
};

char            *ofile;                 /* output file name */
char            *namex[2];              /* seq names: getseqs() */
char            *prog;                  /* prog name for err msgs */
char            *seqx[2];               /* seqs: getseqs() */
int             dmax;                   /* best diag: nw() */
int             dmax0;                  /* final diag */
int             dna;                    /* set if dna: main() */
int             endgaps;                /* set if penalizing end gaps */
int             gapx, gapy;             /* total gaps in seqs */
int             len0, len1;             /* seq lens */
int             ngapx, ngapy;           /* total size of gaps */
int             smax;                   /* max score: nw() */
int             *xbm;                   /* bitmap for matching */
long            offset;                 /* current offset in jmp file */
struct  diag    *dx;                    /* holds diagonals */
struct  path    pp[2];                  /* holds path for seqs */

char            *calloc(), *malloc(), *index(), *strcpy();
char            *getseq(), *g_calloc();
```

Table 1 (cont')

```
/* Needleman-Wunsch alignment program
 *
 * usage: progs file1 file2
 *   where file1 and file2 are two dna or two protein sequences.
 *   The sequences can be in upper- or lower-case an may contain ambiguity
 *   Any lines beginning with ';', '>' or '<' are ignored
 *   Max file length is 65535 (limited by unsigned short x in the jmp struct)
 *   A sequence with 1/3 or more of its elements ACGTU is assumed to be DNA
 *   Output is in the file "align.out"
 *
 * The program may create a tmp file in /tmp to hold info about traceback.
 * Original version developed under BSD 4.3 on a vax 8650
 */
#include "nw.h"
#include "day.h"

static    _dbval[26] = {
        1,14,2,13,0,0,4,11,0,0,12,0,3,15,0,0,0,5,6,8,8,7,9,0,10,0
};

static    _pbval[26] = {
        1, 2|(1<<('D'-'A'))|(1<<('N'-'A')), 4, 8, 16, 32, 64,
        128, 256, 0xFFFFFFF, 1<<10, 1<<11, 1<<12, 1<<13, 1<<14,
        1<<15, 1<<16, 1<<17, 1<<18, 1<<19, 1<<20, 1<<21, 1<<22,
        1<<23, 1<<24, 1<<25|(1<<('E'-'A'))|(1<<('Q'-'A'))
};

main(ac, av)                                                    main
        int     ac;
        char    *av[];
{
        prog = av[0];
        if (ac != 3) {
                fprintf(stderr,"usage: %s file1 file2\n", prog);
                fprintf(stderr,"where file1 and file2 are two dna or two protein sequences.\n");
                fprintf(stderr,"The sequences can be in upper- or lower-case\n");
                fprintf(stderr,"Any lines beginning with ';' or '<' are ignored\n");
                fprintf(stderr,"Output is in the file \"align.out\"\n");
                exit(1);
        }
        namex[0] = av[1];
        namex[1] = av[2];
        seqx[0] = getseq(namex[0], &len0);
        seqx[1] = getseq(namex[1], &len1);
        xbm = (dna)? _dbval : _pbval;

        endgaps = 0;                    /* 1 to penalize endgaps */
        ofile = "align.out";            /* output file */

        nw();                   /* fill in the matrix, get the possible jmps */
        readjmps();             /* get the actual jmps */
        print();                /* print stats, alignment */

        cleanup(0);             /* unlink any tmp files */
}
```

Table 1 (cont')

```
/* do the alignment, return best score: main()
 * dna: values in Fitch and Smith, PNAS, 80, 1382-1386, 1983
 * pro: PAM 250 values
 * When scores are equal, we prefer mismatches to any gap, prefer
 * a new gap to extending an ongoing gap, and prefer a gap in seqx
 * to a gap in seq y.
 */
nw()                                                                    nw
{
        char            *px, *py;               /* seqs and ptrs */
        int             *ndely, *dely;  /* keep track of dely */
        int             ndelx, delx;    /* keep track of delx */
        int             *tmp;           /* for swapping row0, row1 */
        int             mis;            /* score for each type */
        int             ins0, ins1;     /* insertion penalties */
        register        id;             /* diagonal index */
        register        ij;             /* jmp index */
        register        *col0, *col1;   /* score for curr, last row */
        register        xx, yy;         /* index into seqs */

        dx = (struct diag *)g_calloc("to get diags", len0+len1+1, sizeof(struct diag));

        ndely = (int *)g_calloc("to get ndely", len1+1, sizeof(int));
        dely = (int *)g_calloc("to get dely", len1+1, sizeof(int));
        col0 = (int *)g_calloc("to get col0", len1+1, sizeof(int));
        col1 = (int *)g_calloc("to get col1", len1+1, sizeof(int));
        ins0 = (dna)? DINS0 : PINS0;
        ins1 = (dna)? DINS1 : PINS1;

        smax = -10000;
        if (endgaps) {
                for (col0[0] = dely[0] = -ins0, yy = 1; yy <= len1; yy++) {
                        col0[yy] = dely[yy] = col0[yy-1] - ins1;
                        ndely[yy] = yy;
                }
                col0[0] = 0;            /* Waterman Bull Math Biol 84 */
        }
        else
                for (yy = 1; yy <= len1; yy++)
                        dely[yy] = -ins0;

        /* fill in match matrix
         */
        for (px = seqx[0], xx = 1; xx <= len0; px++, xx++) {
                /* initialize first entry in col
                 */
                if (endgaps) {
                        if (xx == 1)
                                col1[0] = delx = -(ins0+ins1);
                        else
                                col1[0] = delx = col0[0] - ins1;
                        ndelx = xx;
                }
                else {
                        col1[0] = 0;
                        delx = -ins0;
                        ndelx = 0;
                }
```

Table 1 (cont')


```
for (py = seqx[1], yy = 1; yy <= len1; py++, yy++) {
        mis = col0[yy-1];
        if (dna)
                mis += (xbm[*px-'A']&xbm[*py-'A'])? DMAT : DMIS;
        else
                mis += _day[*px-'A'][*py-'A'];

        /* update penalty for del in x seq;
         * favor new del over ongong del
         * ignore MAXGAP if weighting endgaps
         */
        if (endgaps || ndely[yy] < MAXGAP) {
                if (col0[yy] - ins0 >= dely[yy]) {
                        dely[yy] = col0[yy] - (ins0+ins1);
                        ndely[yy] = 1;
                } else {
                        dely[yy] -= ins1;
                        ndely[yy]++;
                }
        } else {
                if (col0[yy] - (ins0+ins1) >= dely[yy]) {
                        dely[yy] = col0[yy] - (ins0+ins1);
                        ndely[yy] = 1;
                } else
                        ndely[yy]++;
        }

        /* update penalty for del in y seq;
         * favor new del over ongong del
         */
        if (endgaps || ndelx < MAXGAP) {
                if (col1[yy-1] - ins0 >= delx) {
                        delx = col1[yy-1] - (ins0+ins1);
                        ndelx = 1;
                } else {
                        delx -= ins1;
                        ndelx++;
                }
        } else {
                if (col1[yy-1] - (ins0+ins1) >= delx) {
                        delx = col1[yy-1] - (ins0+ins1);
                        ndelx = 1;
                } else
                        ndelx++;
        }

        /* pick the maximum score; we're favoring
         * mis over any del and delx over dely
         */
```

Table 1 (cont')


```
                id = xx - yy + len1 - 1;
                if (mis >= delx && mis >= dely[yy])
                        col1[yy] = mis;
                else if (delx >= dely[yy]) {
                        col1[yy] = delx;
                        ij = dx[id].ijmp;
                        if (dx[id].jp.n[0] && (!dna  || (ndelx >= MAXJMP
                        && xx > dx[id].jp.x[ij]+MX) || mis > dx[id].score+DINS0)) {
                                dx[id].ijmp++;
                                if (++ij >= MAXJMP) {
                                        writejmps(id);
                                        ij = dx[id].ijmp = 0;
                                        dx[id].offset = offset;
                                        offset += sizeof(struct jmp) + sizeof(offset);
                                }
                        }
                        dx[id].jp.n[ij] = ndelx;
                        dx[id].jp.x[ij] = xx;
                        dx[id].score = delx;
                }
                else {
                        col1[yy] = dely[yy];
                        ij = dx[id].ijmp;
        if (dx[id].jp.n[0] && (!dna || (ndely[yy] >= MAXJMP
                        && xx > dx[id].jp.x[ij]+MX) || mis > dx[id].score+DINS0)) {
                                dx[id].ijmp++;
                                if (++ij >= MAXJMP) {
                                        writejmps(id);
                                        ij = dx[id].ijmp = 0;
                                        dx[id].offset = offset;
                                        offset += sizeof(struct jmp) + sizeof(offset);
                                }
                        }
                        dx[id].jp.n[ij] = -ndely[yy];
                        dx[id].jp.x[ij] = xx;
                        dx[id].score = dely[yy];
                }
                if (xx == len0 && yy < len1) {
                        /* last col
                         */
                        if (endgaps)
                                col1[yy] -= ins0+ins1*(len1-yy);
                        if (col1[yy] > smax) {
                                smax = col1[yy];
                                dmax = id;
                        }
                }
        }
        if (endgaps && xx < len0)
                col1[yy-1] -= ins0+ins1*(len0-xx);
        if (col1[yy-1] > smax) {
                smax = col1[yy-1];
                dmax = id;
        }
        tmp = col0; col0 = col1; col1 = tmp;
}
(void) free((char *)ndely);
(void) free((char *)dely);
(void) free((char *)col0);
(void) free((char *)col1);
}
```

Table 1 (cont')

```
/*
 *
 * print() -- only routine visible outside this module
 *
 * static:
 * getmat() -- trace back best path, count matches: print()
 * pr_align() -- print alignment of described in array p[]: print()
 * dumpblock() -- dump a block of lines with numbers, stars: pr_align()
 * nums() -- put out a number line: dumpblock()
 * putline() -- put out a line (name, [num], seq, [num]): dumpblock()
 * stars() - -put a line of stars: dumpblock()
 * stripname() -- strip any path and prefix from a seqname
 */

#include "nw.h"

#define SPC     3
#define P_LINE  256     /* maximum output line */
#define P_SPC   3       /* space between name or num and seq */

extern  _day[26][26];
int     olen;           /* set output line length */
FILE    *fx;            /* output file */

print()
{
        int     lx, ly, firstgap, lastgap;      /* overlap */

        if ((fx = fopen(ofile, "w")) == 0) {
                fprintf(stderr,"%s: can't write %s\n", prog, ofile);
                cleanup(1);
        }
        fprintf(fx, "< first sequence: %s (length = %d)\n", namex[0], len0);
        fprintf(fx, "< second sequence: %s (length = %d)\n", namex[1], len1);
        olen = 60;
        lx = len0;
        ly = len1;
        firstgap = lastgap = 0;
        if (dmax < len1 - 1) {          /* leading gap in x */
                pp[0].spc = firstgap = len1 - dmax - 1;
                ly -= pp[0].spc;
        }
        else if (dmax > len1 - 1) {     /* leading gap in y */
                pp[1].spc = firstgap = dmax - (len1 - 1);
                lx -= pp[1].spc;
        }
        if (dmax0 < len0 - 1) {         /* trailing gap in x */
                lastgap = len0 - dmax0 -1;
                lx -= lastgap;
        }
        else if (dmax0 > len0 - 1) {    /* trailing gap in y */
                lastgap = dmax0 - (len0 - 1);
                ly -= lastgap;
        }
        getmat(lx, ly, firstgap, lastgap);
        pr_align();
}
```

print

Table 1 (cont')

```
/*
 * trace back the best path, count matches
 */
static
getmat(lx, ly, firstgap, lastgap)                                    getmat
        int     lx, ly;                 /* "core" (minus endgaps) */
        int     firstgap, lastgap;      /* leading trailing overlap */
{
        int             nm, i0, i1, siz0, siz1;
        char            outx[32];
        double          pct;
        register        n0, n1;
        register char   *p0, *p1;

        /* get total matches, score
         */
        i0 = i1 = siz0 = siz1 = 0;
        p0 = seqx[0] + pp[1].spc;
        p1 = seqx[1] + pp[0].spc;
        n0 = pp[1].spc + 1;
        n1 = pp[0].spc + 1;

        nm = 0;
        while ( *p0 && *p1 ) {
                if (siz0) {
                        p1++;
                        n1++;
                        siz0--;
                }
                else if (siz1) {
                        p0++;
                        n0++;
                        siz1--;
                }
                else {
                        if (xbm[*p0-'A']&xbm[*p1-'A'])
                                nm++;
                        if (n0++ == pp[0].x[i0])
                                siz0 = pp[0].n[i0++];
                        if (n1++ == pp[1].x[i1])
                                siz1 = pp[1].n[i1++];
                        p0++;
                        p1++;
                }
        }

        /* pct homology:
         * if penalizing endgaps, base is the shorter seq
         * else, knock off overhangs and take shorter core
         */
        if (endgaps)
                lx = (len0 < len1)? len0 : len1;
        else
                lx = (lx < ly)? lx : ly;
        pct = 100.*(double)nm/(double)lx;
        fprintf(fx, "\n");
        fprintf(fx, "< %d match%s in an overlap of %d: %.2f percent similarity\n",
                nm, (nm == 1)? "" : "es", lx, pct);
```

Table 1 (cont')

...getmat

```
        fprintf(fx, "<gaps in first sequence: %d", gapx);
        if (gapx) {
                (void) sprintf(outx, " (%d %s%s)",
                        ngapx, (dna)? "base":"residue", (ngapx == 1)? "":"s");
                fprintf(fx,"%s", outx);

        fprintf(fx, ", gaps in second sequence: %d", gapy);
        if (gapy) {
                (void) sprintf(outx, " (%d %s%s)",
                        ngapy, (dna)? "base":"residue", (ngapy == 1)? "":"s");
                fprintf(fx,"%s", outx);
        }
        if (dna)
                fprintf(fx,
                "\n<score: %d (match = %d, mismatch = %d, gap penalty = %d + %d per base)\n",
                smax, DMAT, DMIS, DINS0, DINS1);
        else
                fprintf(fx,
                "\n<score: %d (Dayhoff PAM 250 matrix, gap penalty = %d + %d per residue)\n",
                smax, PINS0, PINS1);
        if (endgaps)
                fprintf(fx,
                "<endgaps penalized. left endgap: %d %s%s, right endgap: %d %s%s\n",
                firstgap, (dna)? "base" : "residue", (firstgap == 1)? "" : "s",
                lastgap, (dna)? "base" : "residue", (lastgap == 1)? "" : "s");
        else
                fprintf(fx, "<endgaps not penalized\n");
}

static          nm;             /* matches in core -- for checking */
static          lmax;           /* lengths of stripped file names */
static          ij[2];          /* jmp index for a path */
static          nc[2];          /* number at start of current line */
static          ni[2];          /* current elem number -- for gapping */
static          siz[2];
static char     *ps[2];         /* ptr to current element */
static char     *po[2];         /* ptr to next output char slot */
static char     out[2][P_LINE]; /* output line */
static char     star[P_LINE];   /* set by stars() */

/*
 * print alignment of described in struct path pp[]
 */
static
```

pr_align

```
pr_align()
{
        int             nn;     /* char count */
        int             more;
        register        i;

        for (i = 0, lmax = 0; i < 2; i++) {
                nn = stripname(namex[i]);
                if (nn > lmax)
                        lmax = nn;

                nc[i] = 1;
                ni[i] = 1;
                siz[i] = ij[i] = 0;
                ps[i] = seqx[i];
                po[i] = out[i];
        }
```

Table 1 (cont')

...pr_align

```
        for (nn = nm = 0, more = 1; more; ) {
                for (i = more = 0; i < 2; i++) {
                        /*
                         * do we have more of this sequence?
                         */
                        if (!*ps[i])
                                continue;

                        more++;

                        if (pp[i].spc) {        /* leading space */
                                *po[i]++ = ' ';
                                pp[i].spc--;
                        }
                        else if (siz[i]) {      /* in a gap */
                                *po[i]++ = '-';
                                siz[i]--;
                        }
                        else {                  /* we're putting a seq element
                                                 */
                                *po[i] = *ps[i];
                                if (islower(*ps[i]))
                                        *ps[i] = toupper(*ps[i]);
                                po[i]++;
                                ps[i]++;

                                /*
                                 * are we at next gap for this seq?
                                 */
                                if (ni[i] == pp[i].x[ij[i]]) {
                                        /*
                                         * we need to merge all gaps
                                         * at this location
                                         */
                                        siz[i] = pp[i].n[ij[i]++];
                                        while (ni[i] == pp[i].x[ij[i]])
                                                siz[i] += pp[i].n[ij[i]++];
                                }
                                ni[i]++;
                        }
                }
                if (++nn == olen || !more && nn) {
                        dumpblock();
                        for (i = 0; i < 2; i++)
                                po[i] = out[i];
                        nn = 0;
                }
        }
}

/*
 * dump a block of lines, including numbers, stars: pr_align()
 */
static
dumpblock()                                                    dumpblock
{
        register  i;

        for (i = 0; i < 2; i++)
                *po[i]-- = '\0';
```

Table 1 (cont')


```
        (void) putc('\n', fx);
        for (i = 0; i < 2; i++) {
                if (*out[i] && (*out[i] != ' ' || *(po[i]) != ' ')) {
                        if (i == 0)
                                nums(i);
                        if (i == 0 && *out[1])
                                stars();
                        putline(i);
                        if (i == 0 && *out[1])
                                fprintf(fx, star);
                        if (i == 1)
                                nums(i);
                }
        }
}

/*
 * put out a number line: dumpblock()
 */
static
nums(ix)                                                        nums
        int     ix;     /* index in out[] holding seq line */
{
        char            nline[P_LINE];
        register        i, j;
        register char   *pn, *px, *py;

        for (pn = nline, i = 0; i < lmax+P_SPC; i++, pn++)
                *pn = ' ';
        for (i = nc[ix], py = out[ix]; *py; py++, pn++) {
                if (*py == ' ' || *py == '-')
                        *pn = ' ';
                else {
                        if (i%10 == 0 || (i == 1 && nc[ix] != 1)) {
                                j = (i < 0)? -i : i;
                                for (px = pn; j; j /= 10, px--)
                                        *px = j%10 + '0';
                                if (i < 0)
                                        *px = '-';
                        }
                        else
                                *pn = ' ';
                        i++;
                }
        }
        *pn = '\0';
        nc[ix] = i;
        for (pn = nline; *pn; pn++)
                (void) putc(*pn, fx);
        (void) putc('\n', fx);
}

/*
 * put out a line (name, [num], seq, [num]): dumpblock()
 */
static
putline(ix)                                                     putline
        int     ix;
{
```

Table 1 (cont')

...putline

```
        int             i;
        register char   *px;

        for (px = namex[ix], i = 0; *px && *px != ':'; px++, i++)
                (void) putc(*px, fx);
        for (; i < lmax+P_SPC; i++)
                (void) putc(' ', fx);

        /* these count from 1:
         * ni[] is current element (from 1)
         * nc[] is number at start of current line
         */
        for (px = out[ix]; *px; px++)
                (void) putc(*px&0x7F, fx);
        (void) putc('\n', fx);
}

/*
 * put a line of stars (seqs always in out[0], out[1]): dumpblock()
 */
static
stars()                                                          stars
{
        int             i;
        register char   *p0, *p1, cx, *px;

        if (!*out[0] || (*out[0] == ' ' && *(po[0]) == ' ') ||
            !*out[1] || (*out[1] == ' ' && *(po[1]) == ' '))
                return;
        px = star;
        for (i = lmax+P_SPC; i; i--)
                *px++ = ' ';

        for (p0 = out[0], p1 = out[1]; *p0 && *p1; p0++, p1++) {
                if (isalpha(*p0) && isalpha(*p1)) {

                        if (xbm[*p0-'A']&xbm[*p1-'A']) {
                                cx = '*';
                                nm++;
                        }
                        else if (!dna && _day[*p0-'A'][*p1-'A'] > 0)
                                cx = '.';
                        else
                                cx = ' ';
                }
                else
                        cx = ' ';
                *px++ = cx;
        }
        *px++ = '\n';
        *px = '\0';
}
```

Table 1 (cont')

```
/*
 * strip path or prefix from pn, return len: pr_align()
 */
static
stripname(pn)
        char    *pn;    /* file name (may be path) */
{
        register char   *px, *py;

        py = 0;
        for (px = pn; *px; px++)
                if (*px == '/')
                        py = px + 1;
        if (py)
                (void) strcpy(pn, py);
        return(strlen(pn));

}
```

stripname

Table 1 (cont')

```
/*
 * cleanup() -- cleanup any tmp file
 * getseq() -- read in seq, set dna, len, maxlen
 * g_calloc() -- calloc() with error checkin
 * readjmps() -- get the good jmps, from tmp file if necessary
 * writejmps() -- write a filled array of jmps to a tmp file: nw()
 */
#include "nw.h"
#include <sys/file.h>

char    *jname = "/tmp/homgXXXXXX";             /* tmp file for jmps */
FILE    *fj;

int     cleanup();                              /* cleanup tmp file */
long    lseek();

/*
 * remove any tmp file if we blow
 */
cleanup(i)                                                          cleanup
        int     i;
{
        if (fj)
                (void) unlink(jname);
        exit(i);
}

/*
 * read, return ptr to seq, set dna, len, maxlen
 * skip lines starting with ';', '<', or '>'
 * seq in upper or lower case
 */
char    *
getseq(file, len)                                                   getseq
        char    *file;  /* file name */
        int     *len;   /* seq len */
{
        char            line[1024], *pseq;
        register char   *px, *py;
        int             natgc, tlen;
        FILE            *fp;

        if ((fp = fopen(file,"r")) == 0) {
                fprintf(stderr,"%s: can't read %s\n", prog, file);
                exit(1);
        }
        tlen = natgc = 0;
        while (fgets(line, 1024, fp)) {
                if (*line == ';' || *line == '<' || *line == '>')
                        continue;
                for (px = line; *px != '\n'; px++)
                        if (isupper(*px) || islower(*px))
                                tlen++;
        }
        if ((pseq = malloc((unsigned)(tlen+6))) == 0) {
                fprintf(stderr,"%s: malloc() failed to get %d bytes for %s\n", prog, tlen+6, file);
                exit(1);
        }
        pseq[0] = pseq[1] = pseq[2] = pseq[3] = '\0';
```

Table 1 (cont')

...getseq

```
        py = pseq + 4;
        *len = tlen;
        rewind(fp);

        while (fgets(line, 1024, fp)) {
                if (*line == ';' || *line == '<' || *line == '>')
                        continue;
                for (px = line; *px != '\n'; px++) {
                        if (isupper(*px))
                                *py++ = *px;
                        else if (islower(*px))
                                *py++ = toupper(*px);
                        if (index("ATGCU",*(py-1)))
                                natgc++;
                }
        }
        *py++ = '\0';
        *py = '\0';
        (void) fclose(fp);
        dna = natgc > (tlen/3);
        return(pseq+4);
}

char    *
g_calloc(msg, nx, sz)                                                   g_calloc
        char    *msg;           /* program, calling routine */
        int     nx, sz;         /* number and size of elements */
{
        char            *px, *calloc();

        if ((px = calloc((unsigned)nx, (unsigned)sz)) == 0) {
                if (*msg) {
                        fprintf(stderr, "%s: g_calloc() failed %s (n=%d, sz=%d)\n", prog, msg, nx, sz);
                        exit(1);
                }
        }
        return(px);
}

/*
 * get final jmps from dx[] or tmp file, set pp[], reset dmax: main()
 */
readjmps()                                                              readjmps
{
        int             fd = -1;
        int             siz, i0, i1;
        register        i, j, xx;

        if (fj) {
                (void) fclose(fj);
                if ((fd = open(jname, O_RDONLY, 0)) < 0) {
                        fprintf(stderr, "%s: can't open() %s\n", prog, jname);
                        cleanup(1);
                }
        }
        for (i = i0 = i1 = 0, dmax0 = dmax, xx = len0; ; i++) {
                while (1) {
                        for (j = dx[dmax].ijmp; j >= 0 && dx[dmax].jp.x[j] >= xx; j--)
                                ;
```

Table 1 (cont')


```
                if (j < 0 && dx[dmax].offset && fj) {
                        (void) lseek(fd, dx[dmax].offset, 0);
                        (void) read(fd, (char *)&dx[dmax].jp, sizeof(struct jmp));
                        (void) read(fd, (char *)&dx[dmax].offset, sizeof(dx[dmax].offset));
                        dx[dmax].ijmp = MAXJMP-1;
                }
                else
                        break;
        }
        if (i >= JMPS) {
                fprintf(stderr, "%s: too many gaps in alignment\n", prog);
                cleanup(1);
        }
        if (j >= 0) {
                siz = dx[dmax].jp.n[j];
                xx = dx[dmax].jp.x[j];
                dmax += siz;
                if (siz < 0) {                  /* gap in second seq */
                        pp[1].n[i1] = -siz;
                        xx += siz;
                        /* id = xx - yy + len1 - 1
                         */
                        pp[1].x[i1] = xx - dmax + len1 - 1;
                        gapy++;
                        ngapy -= siz;
/* ignore MAXGAP when doing endgaps */
                        siz = (-siz < MAXGAP || endgaps)? -siz : MAXGAP;
                        i1++;
                }
                else if (siz > 0) {  /* gap in first seq */
                        pp[0].n[i0] = siz;
                        pp[0].x[i0] = xx;
                        gapx++;
                        ngapx += siz;
/* ignore MAXGAP when doing endgaps */
                        siz = (siz < MAXGAP || endgaps)? siz : MAXGAP;
                        i0++;
                }
        }
        else
                break;
}

/* reverse the order of jmps
 */
for (j = 0, i0--; j < i0; j++, i0--) {
        i = pp[0].n[j]; pp[0].n[j] = pp[0].n[i0]; pp[0].n[i0] = i;
        i = pp[0].x[j]; pp[0].x[j] = pp[0].x[i0]; pp[0].x[i0] = i;
}
for (j = 0, i1--; j < i1; j++, i1--) {
        i = pp[1].n[j]; pp[1].n[j] = pp[1].n[i1]; pp[1].n[i1] = i;
        i = pp[1].x[j]; pp[1].x[j] = pp[1].x[i1]; pp[1].x[i1] = i;
}
if (fd >= 0)
        (void) close(fd);
if (fj) {
        (void) unlink(jname);
        fj = 0;
        offset = 0;
}
}
```

Table 1 (cont')

```
/*
 * write a filled jmp struct offset of the prev one (if any): nw()
 */
writejmps(ix)                                                    writejmps
        int     ix;
{
        char    *mktemp();

        if (!fj) {
                if (mktemp(jname) < 0) {
                        fprintf(stderr, "%s: can't mktemp() %s\n", prog, jname);
                        cleanup(1);
                }
                if ((fj = fopen(jname, "w")) == 0) {
                        fprintf(stderr, "%s: can't write %s\n", prog, jname);
                        exit(1);
                }
        }
        (void) fwrite((char *)&dx[ix].jp, sizeof(struct jmp), 1, fj);
        (void) fwrite((char *)&dx[ix].offset, sizeof(dx[ix].offset), 1, fj);
}
```

Table 2

PRO	XXXXXXXXXXXXXXX	(Length = 15 amino acids)
Comparison Protein	XXXXXYYYYYYY	(Length = 12 amino acids)

% amino acid sequence identity =

(the number of identically matching amino acid residues between the two polypeptide sequences as determined by ALIGN-2) divided by (the total number of amino acid residues of the PRO polypeptide) =

5 divided by 15 = 33.3%

Table 3

PRO	XXXXXXXXXX	(Length = 10 amino acids)
Comparison Protein	XXXXXYYYYYYZZYZ	(Length = 15 amino acids)

% amino acid sequence identity =

(the number of identically matching amino acid residues between the two polypeptide sequences as determined by ALIGN-2) divided by (the total number of amino acid residues of the PRO polypeptide) =

5 divided by 10 = 50%

Table 4

PRO-DNA	NNNNNNNNNNNNNN	(Length = 14 nucleotides)
Comparison DNA	NNNNNNLLLLLLLLL	(Length = 16 nucleotides)

% nucleic acid sequence identity =

(the number of identically matching nucleotides between the two nucleic acid sequences as determined by ALIGN-2) divided by (the total number of nucleotides of the PRO-DNA nucleic acid sequence) =

6 divided by 14 = 42.9%

Table 5

PRO-DNA	NNNNNNNNNNN	(Length = 12 nucleotides)
Comparison DNA	NNNNLLLVV	(Length = 9 nucleotides)

% nucleic acid sequence identity =

(the number of identically matching nucleotides between the two nucleic acid sequences as determined by ALIGN-2) divided by (the total number of nucleotides of the PRO-DNA nucleic acid sequence) =

4 divided by 12 = 33.3%

II. Compositions and Methods of the Invention

A. Full-Length PRO Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO polypeptides. In particular, cDNAs encoding various PRO polypeptides have been identified and isolated, as disclosed in further detail in the Examples below. It is noted that proteins produced in separate expression rounds may be given different PRO numbers but the UNQ number is unique for any given DNA and the encoded protein, and will not be changed. However, for sake of simplicity, in the present specification the protein encoded by the full length native nucleic acid molecules disclosed herein as well as all further native homologues and variants included in the foregoing definition of PRO, will be referred to as "PRO/number", regardless of their origin or mode of preparation.

As disclosed in the Examples below, various cDNA clones have been deposited with the ATCC. The actual nucleotide sequences of those clones can readily be determined by the skilled artisan by sequencing of the deposited clone using routine methods in the art. The predicted amino acid sequence can be determined from the nucleotide sequence using routine skill. For the PRO polypeptides and encoding nucleic acids described herein, Applicants have identified what is believed to be the reading frame best identifiable with the sequence information available at the time.

1. Full-length PRO211 and PRO217 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO211 and PRO217. In particular, Applicants have identified and isolated cDNA encoding PRO211 and PRO217 polypeptides, as disclosed in further detail in the Examples below. Using BLAST (FastA format) sequence alignment computer programs, Applicants found that cDNA sequences encoding full-length native sequence PRO211 and PRO217 have homologies to known proteins having EGF-like domains. Specifically, the cDNA sequence DNA32292-1131 (Figure 1, SEQ ID NO:1) has certain identify and a Blast score of 209 with PAC6_RAT and certain identify and a Blast score of 206 with Fibulin-1, isoform c precursor. The cDNA sequence DNA33094-1131 (Figure 3, SEQ ID NO:3) has 36% identity and a Blast score of 336 with eastern newt tenascin, and 37% identity and a Blast score of 331 with human tenascin-X precursor. Accordingly, it is presently believed that PRO211 and PRO217 polypeptides disclosed in the present application are newly identified members of the EGF-like family and possesses properties typical of the EGF-like protein family.

2. Full-length PRO230 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO230. In particular, Applicants have identified and isolated cDNA encoding a PRO230 polypeptide, as disclosed in further detail in the Examples below. Using known programs such as BLAST and FastA sequence alignment computer programs, Applicants found that a cDNA sequence encoding full-length native sequence PRO230 has 48% amino acid identity with the rabbit tubulointerstitial nephritis antigen precursor. Accordingly, it is presently believed that PRO230 polypeptide disclosed in the present application is a newly identified member of the tubulointerstitial nephritis antigen family and possesses

the ability to be recognized by human autoantibodies in certain forms of tubulointerstitial nephritis.

3. Full-length PRO232 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO232. In particular, Applicants have identified and isolated cDNA encoding a PRO232 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a portion of the full-length native sequence PRO232 (shown in Figure 9 and SEQ ID NO:18) has 35% sequence identity with a stem cell surface antigen from Gallus gallus. Accordingly, it is presently believed that the PRO232 polypeptide disclosed in the present application may be a newly identified stem cell antigen.

4. Full-length PRO187 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO187. In particular, Applicants have identified and isolated cDNA encoding a PRO187 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO187 (shown in Figure 15) has 74% amino acid sequence identity and BLAST score of 310 with various androgen-induced growth factors and FGF-8. Accordingly, it is presently believed that PRO187 polypeptide disclosed in the present application is a newly identified member of the FGF-8 protein family and may possess identify activity or property typical of the FGF-8-like protein family.

5. Full-length PRO265 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO265. In particular, Applicants have identified and isolated cDNA encoding a PRO265 polypeptide, as disclosed in further detail in the Examples below. Using programs such as BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO265 polypeptide have significant homology with the fibromodulin protein and fibromodulin precursor protein. Applicants have also found that the DNA encoding the PRO265 polypeptide has significant homology with platelet glycoprotein V, a member of the leucine rich related protein family involved in skin and wound repair. Accordingly, it is presently believed that PRO265 polypeptide disclosed in the present application is a newly identified member of the leucine rich repeat family and possesses protein protein binding capabilities, as well as be involved in skin and wound repair as typical of this family.

6. Full-length PRO219 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO219. In particular, Applicants have identified and isolated cDNA encoding a PRO219 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO219 polypeptide have significant homology with the mouse and human matrilin-2 precursor polypeptides. Accordingly, it is presently

believed that PRO219 polypeptide disclosed in the present application is related to the matrilin-2 precursor polypeptide.

7. **Full-length PRO246 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO246. In particular, Applicants have identified and isolated cDNA encoding a PRO246 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a portion of the PRO246 polypeptide has significant homology with the human cell surface protein HCAR. Accordingly, it is presently believed that PRO246 polypeptide disclosed in the present application may be a newly identified membrane-bound virus receptor or tumor cell-specific antigen.

8. **Full-length PRO228 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO228. In particular, Applicants have identified and isolated cDNA encoding a PRO228 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO228 polypeptide have significant homology with the EMR1 protein. Applicants have also found that the DNA encoding the PRO228 polypeptide has significant homology with latrophilin, macrophage-restricted cell surface glycoprotein, B0457.1 and leucocyte antigen CD97 precursor. Accordingly, it is presently believed that PRO228 polypeptide disclosed in the present application is a newly identified member of the seven transmembrane superfamily and possesses characteristics and functional properties typical of this family. In particular, it is believed that PRO228 is a new member of the subgroup within this family to which CD97 and EMR1 belong.

9. **Full-length PRO533 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO533. In particular, Applicants have identified and isolated cDNA encoding a PRO533 polypeptide, as disclosed in further detail in the Examples below. Using BLAST-2 and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO533 (shown in Figure 22 and SEQ ID NO:59) has a Blast score of 509 and 53% amino acid sequence identity with fibroblast growth factor (FGF). Accordingly, it is presently believed that PRO533 disclosed in the present application is a newly identified member of the fibroblast growth factor family and may possess activity typical of such polypeptides.

10. **Full-length PRO245 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO245. In particular, Applicants have identified and isolated cDNA encoding a PRO245 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a portion of the amino acid sequence of the

PRO245 polypeptide has 60% amino acid identity with the human c-myb protein. Accordingly, it is presently believed that the PRO245 polypeptide disclosed in the present application may be a newly identified member of the transmembrane protein tyrosine kinase family.

11. Full-length PRO220, PRO221 and PRO227 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO220, PRO221 and PRO227. In particular, Applicants have identified and isolated cDNAs encoding a PRO220, PRO221 and PRO227 polypeptide, respectively, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, PRO220 has amino acid identity with the amino acid sequence of a leucine rich protein wherein the identity is 87%. PRO220 additionally has amino acid identity with the neuronal leucine rich protein wherein the identity is 55%. The neuronal leucine rich protein is further described in Taguchi, *et al.*, Mol. Brain Res., 35:31-40 (1996).

PRO221 has amino acid identity with the SLIT protein precursor, wherein different portions of these two proteins have the respective percent identities of 39%, 38%, 34%, 31%, and 30%.

PRO227 has amino acid identity with the amino acid sequence of platelet glycoprotein V precursor. The same results were obtained for human glycoprotein V. Different portions of these two proteins show the following percent identities of 30%, 28%, 28%, 31%, 35%, 39% and 27%.

Accordingly, it is presently believed that PRO220, PRO221 and PRO227 polypeptides disclosed in the present application are newly identified members of the leucine rich repeat protein superfamily and that each possesses protein-protein binding capabilities typical of the leucine rich repeat protein superfamily. It is also believed that they have capabilities similar to those of SLIT, the leucine rich repeat protein and human glycoprotein V.

12. Full-length PRO258 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO258. In particular, Applicants have identified and isolated cDNA encoding a PRO258 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO258 polypeptide have significant homology with the CRTAM and poliovirus receptors. Accordingly, it is presently believed that PRO258 polypeptide disclosed in the present application is a newly identified member of the Ig superfamily and possesses virus receptor capabilities or regulates immune function as typical of this family.

13. Full-length PRO266 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO266. In particular, Applicants have identified and isolated cDNA encoding a PRO266 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO266 polypeptide have significant homology with the SLIT protein from Drosophilia. Accordingly, it is presently believed that PRO266 polypeptide disclosed in the present application is a newly identified member of the leucine rich repeat family

and possesses ligand-ligand binding activity and neuronal development typical of this family. SLIT has been shown to be useful in the study and treatment of Alzheimer's disease, *supra*, and thus, PRO266 may have involvement in the study and cure of this disease.

14. Full-length PRO269 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO269. In particular, Applicants have identified and isolated cDNA encoding a PRO269 polypeptide, as disclosed in further detail in the Examples below. Using BLAST, FastA and sequence alignment computer programs, Applicants found that the amino acid sequence encoded by nucleotides 314 to 1783 of the full-length native sequence PRO269 (shown in Figure 35 and SEQ ID NO:95) has significant homology to human urinary thrombomodulin and various thrombomodulin analogues respectively, to which it was aligned. Accordingly, it is presently believed that PRO269 polypeptide disclosed in the present application is a newly identified member of the thrombomodulin family.

15. Full-length PRO287 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO287. In particular, Applicants have identified and isolated cDNA encoding a PRO287 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO287 polypeptide have significant homology with the type 1 procollagen C-proteinase enhancer protein precursor and type 1 procollagen C-proteinase enhancer protein. Accordingly, it is presently believed that PRO287 polypeptide disclosed in the present application is a newly identified member of the C-proteinase enhancer protein family.

16. Full-length PRO214 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO214. In particular, Applicants have identified and isolated cDNA encoding a PRO214 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO214 polypeptide (shown in Figure 40 and SEQ ID NO:109) has 49% amino acid sequence identity with HT protein, a known member of the EGF-family. The comparison resulted in a BLAST score of 920, with 150 matching nucleotides. Accordingly, it is presently believed that the PRO214 polypeptide disclosed in the present application is a newly identified member of the family comprising EGF domains and may possess activities or properties typical of the EGF-domain containing family.

17. Full-length PRO317 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO317. In particular, cDNA encoding a PRO317 polypeptide has been identified and isolated, as disclosed in further detail in the Examples below. Using BLAST™ and FastA™ sequence alignment computer programs, it was found that a full-length native-sequence PRO317 (shown in

Figure 42 and SEQ ID NO:114) has 92% amino acid sequence identity with EBAF-1. Further, it is closely aligned with many other members of the TGF- superfamily.

Accordingly, it is presently believed that PRO317 disclosed in the present application is a newly identified member of the TGF- superfamily and may possess properties that are therapeutically useful in conditions of uterine bleeding, etc. Hence, PRO317 may be useful in diagnosing or treating abnormal bleeding involved in gynecological diseases, for example, to avoid or lessen the need for a hysterectomy. PRO317 may also be useful as an agent that affects angiogenesis in general, so PRO317 may be useful in anti-tumor indications, or conversely, in treating coronary ischemic conditions.

Library sources reveal that ESTs used to obtain the consensus DNA for generating PRO317 primers and probes were found in normal tissues (uterus, prostate, colon, and pancreas), in several tumors (colon, brain (twice), pancreas, and mullerian cell), and in a heart with ischemia. PRO317 has shown up in several tissues as well, but it does look to have a greater concentration in uterus. Hence, PRO317 may have a broader use by the body than EBAF-1. It is contemplated that, at least for some indications, PRO317 may have opposite effects from EBAF-1.

18. Full-length PRO301 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO301. In particular, Applicants have identified and isolated cDNA encoding a PRO301 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO301 (shown in Figure 44 and SEQ ID NO:119) has a Blast score of 246 corresponding to 30% amino acid sequence identity with human A33 antigen precursor. Accordingly, it is presently believed that PRO301 disclosed in the present application is a newly identified member of the A33 antigen protein family and may be expressed in human neoplastic diseases such as colorectal cancer.

19. Full-length PRO224 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO224. In particular, Applicants have identified and isolated cDNA encoding a PRO224 polypeptide, as disclosed in further detail in the Examples below. Using known programs such as BLAST and FastA sequence alignment computer programs, Applicants found that full-length native PRO224 (Figure 46, SEQ ID NO:127) has amino acid identity with apolipoprotein E receptor 2906 from homo sapiens. The alignments of different portions of these two polypeptides show amino acid identities of 37%, 36%, 30%, 44%, 44% and 28% respectively. Full-length native PRO224 (Figure 46, SEQ ID NO:127) also has amino acid identity with very low-density lipoprotein receptor precursor from gall. The alignments of different portions of these two polypeptides show amino acid identities of 38%, 37%, 42%, 33%, and 37% respectively. Additionally, full-length native PRO224 (Figure 46, SEQ ID NO:127) has amino acid identity with the chicken oocyte receptor P95 from Gallus gallus. The alignments of different portions of these two polypeptides show amino acid identities of 38%, 37%, 42%, 33%, and 37% respectively. Moreover, full-length native PRO224 (Figure 46, SEQ ID NO:127) has amino acid identity with very low density lipoprotein receptor

short form precursor from humans. The alignments of different portions of these two polypeptides show amino acid identities of 32%, 38%, 34%, 45%, and 31%, respectively. Accordingly, it is presently believed that PRO224 polypeptide disclosed in the present application is a newly identified member of the low density lipoprotein receptor family and possesses the structural characteristics required to have the functional ability to recognize and endocytose low density lipoproteins typical of the low density lipoprotein receptor family. (The alignments described above used the following scoring parameters: T=7, S+65, S2=36, Matrix: BLOSUM62.)

20. Full-length PRO222 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO222. In particular, Applicants have identified and isolated cDNA encoding a PRO222 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a sequence encoding full-length native sequence PRO222 (shown in Figure 48 and SEQ ID NO:132) has 25-26% amino acid identity with mouse complement factor h precursor, has 27-29% amino acid identity with complement receptor, has 25-47% amino acid identity with mouse complement C3b receptor type 2 long form precursor, has 40% amino acid identity with human hypothetical protein kiaa0247. Accordingly, it is presently believed that PRO222 polypeptide disclosed in the present application is a newly identified member of the complement receptor family and possesses activity typical of the complement receptor family.

21. Full-length PRO234 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO234. In particular, Applicants have identified and isolated cDNA encoding a PRO234 polypeptide, as disclosed in further detail in the Examples below. Using BLAST (FastA-format) sequence alignment computer programs, Applicants found that a cDNA sequence encoding full-length native sequence PRO234 has 31% identity and Blast score of 134 with E-selectin precursor. Accordingly, it is presently believed that the PRO234 polypeptides disclosed in the present application are newly identified members of the lectin/selectin family and possess activity typical of the lectin/selectin family.

22. Full-length PRO231 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO231. In particular, Applicants have identified and isolated cDNA encoding a PRO231 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that the full-length native sequence PRO231 polypeptide (shown in Figure 52 and SEQ ID NO:142) has 30 % and 31 % amino acid identity with human and rat prostatic acid phosphatase precursor proteins, respectively. Accordingly, it is presently believed that the PRO231 polypeptide disclosed in the present application may be a newly identified member of the acid phosphatase protein family.

23. Full-length PRO229 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO229. In particular, Applicants have identified and isolated cDNA encoding a PRO229 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO229 polypeptide have significant homology with antigen wc1.1, M130 antigen, T cell surface glycoprotein CD6 and CD6. It also is related to Sp-alpha. Accordingly, it is presently believed that PRO229 polypeptide disclosed in the present application is a newly identified member of the family containing scavenger receptor homology, a sequence motif found in a number of proteins involved in immune function and thus possesses immune function and /or segments which resist degradation, typical of this family.

24. **Full-length PRO238 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO238. In particular, Applicants have identified and isolated cDNA encoding a PRO238 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO238 polypeptide have significant homology with reductases, including oxidoreductase and fatty acyl-CoA reductase. Accordingly, it is presently believed that PRO238 polypeptide disclosed in the present application is a newly identified member of the reductase family and possesses reducing activity typical of the reductase family.

25. **Full-length PRO233 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO233. In particular, Applicants have identified and isolated cDNA encoding a PRO233 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO233 polypeptide have significant homology with the reductase protein. Applicants have also found that the DNA encoding the PRO233 polypeptide has significant homology with proteins from *Caenorhabditis elegans*. Accordingly, it is presently believed that PRO233 polypeptide disclosed in the present application is a newly identified member of the reductase family and possesses the ability to effect the redox state of the cell typical of the reductase family.

26. **Full-length PRO223 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO223. In particular, Applicants have identified and isolated cDNA encoding a PRO223 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that the PRO223 polypeptide has significant homology with various serine carboxypeptidase polypeptides. Accordingly, it is presently believed that PRO223 polypeptide disclosed in the present application is a newly identified serine carboxypeptidase.

27. **Full-length PRO235 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides

referred to in the present application as PRO235. In particular, Applicants have identified and isolated cDNA encoding a PRO235 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO235 polypeptide have significant homology with the various plexin proteins. Accordingly, it is presently believed that PRO235 polypeptide disclosed in the present application is a newly identified member of the plexin family and possesses cell adhesion properties typical of the plexin family.

28. Full-length PRO236 and PRO262 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO236 and PRO262. In particular, Applicants have identified and isolated cDNA encoding PRO236 and PRO262 polypeptides, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO236 and PRO262 polypeptides have significant homology with various β-galactosidase and β-galactosidase precursor polypeptides. Accordingly, it is presently believed that the PRO236 and PRO262 polypeptides disclosed in the present application are newly identified β-galactosidase homologs.

29. Full-length PRO239 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO239. In particular, Applicants have identified and isolated cDNA encoding a PRO239 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO239 polypeptide have significant homology with densin proteins. Accordingly, it is presently believed that PRO239 polypeptide disclosed in the present application is a newly identified member of the densin family and possesses cell adhesion and the ability to effect synaptic processes as is typical of the densin family.

30. Full-length PRO257 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO257. In particular, Applicants have identified and isolated cDNA encoding a PRO257 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO257 polypeptide have significant homology with the ebnerin precursor and ebnerin protein. Accordingly, it is presently believed that PRO257 polypeptide disclosed in the present application is a newly identified protein member which is related to the ebnerin protein.

31. Full-length PRO260 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO260. In particular, Applicants have identified and isolated cDNA encoding a PRO260 polypeptide, as disclosed in further detail in the Examples below. Using programs such as BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the

PRO260 polypeptide have significant homology with the alpha-l-fucosidase precursor. Accordingly, it is presently believed that PRO260 polypeptide disclosed in the present application is a newly identified member of the fucosidase family and possesses enzymatic activity related to fucose residues typical of the fucosidase family.

32. **Full-length PRO263 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO263. In particular, Applicants have identified and isolated cDNA encoding a PRO263 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO263 polypeptide have significant homology with the CD44 antigen and related proteins. Accordingly, it is presently believed that PRO263 polypeptide disclosed in the present application is a newly identified member of the CD44 antigen family and possesses at least one of the properties associated with these antigens, i.e., cancer and HIV marker, cell-cell or cell-matrix interactions, regulating cell traffic, lymph node homing, transmission of growth signals, and presentation of chemokines and growth facors to traveling cells.

33. **Full-length PRO270 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO270. In particular, Applicants have identified and isolated cDNA encoding a PRO270 polypeptide, as disclosed in further detail in the Examples below. Using BLAST, FastA and sequence alignment computer programs, Applicants found that various portions of the PRO270 polypeptide have significant homology with various thioredoxin proteins. Accordingly, it is presently believed that PRO270 polypeptide disclosed in the present application is a newly identified member of the thioredoxin family and possesses the ability to effect reduction-oxidation (redox) state typical of the thioredoxin family.

34. **Full-length PRO271 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO271. In particular, Applicants have identified and isolated cDNA encoding a PRO271 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that the PRO271 polypeptide has significant homology with various link proteins and precursors thereof. Accordingly, it is presently believed that PRO271 polypeptide disclosed in the present application is a newly identified link protein homolog.

35. **Full-length PRO272 Polypeptides**

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO272. In particular, Applicants have identified and isolated cDNA encoding a PRO272 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO272 polypeptide have significant homology with the human reticulocalbin protein and its precursors. Applicants have also found that

the DNA encoding the PRO272 polypeptide has significant homology with the mouse reticulocalbin precursor protein. Accordingly, it is presently believed that PRO272 polypeptide disclosed in the present application is a newly identified member of the reticulocalbin family and possesses the ability to bind calcium typical of the reticulocalbin family.

36. Full-length PRO294 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO294. In particular, Applicants have identified and isolated cDNA encoding a PRO294 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO294 polypeptide have significant homology with the various portions of a number of collagen proteins. Accordingly, it is presently believed that PRO294 polypeptide disclosed in the present application is a newly identified member of the collagen family.

37. Full-length PRO295 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO295. In particular, Applicants have identified and isolated cDNA encoding a PRO295 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO295 polypeptide have significant homology with integrin proteins. Accordingly, it is presently believed that PRO295 polypeptide disclosed in the present application is a newly identified member of the integrin family and possesses cell adhesion typical of the integrin family.

38. Full-length PRO293 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO293. In particular, Applicants have identified and isolated cDNA encoding a PRO293 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that portions of the PRO293 polypeptide have significant homology with the neuronal leucine rich repeat proteins 1 and 2, (NLRR-1 and NLRR-2), particularly NLRR-2. Accordingly, it is presently believed that PRO293 polypeptide disclosed in the present application is a newly identified member of the neuronal leucine rich repeat protein family and possesses ligand-ligand binding activity typical of the NRLL protein family.

39. Full-length PRO247 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO247. In particular, Applicants have identified and isolated cDNA encoding a PRO247 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO247 polypeptide have significant homology with densin. Applicants have also found that the DNA encoding the PRO247 polypeptide

has significant homology with a number of other proteins, including KIAA0231. Accordingly, it is presently believed that PRO247 polypeptide disclosed in the present application is a newly identified member of the leucine rich repeat family and possesses ligand binding abilities typical of this family.

40. Full-length PRO302, PRO303, PRO304, PRO307 and PRO343 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO302, PRO303, PRO304, PRO307 and PRO343. In particular, Applicants have identified and isolated cDNA encoding PRO302, PRO303, PRO304, PRO307 and PRO343 polypeptides, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO302, PRO303, PRO304, PRO307 and PRO343 polypeptides have significant homology with various protease proteins. Accordingly, it is presently believed that the PRO302, PRO303, PRO304, PRO307 and PRO343 polypeptides disclosed in the present application are newly identified protease proteins.

41. Full-length PRO328 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO328. In particular, Applicants have identified and isolated cDNA encoding a PRO328 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO328 polypeptide have significant homology with the human glioblastoma protein ("GLIP"). Further, Applicants found that various portions of the PRO328 polypeptide have significant homology with the cysteine rich secretory protein ("CRISP") as identified by BLAST homology [ECCRISP3_1, S68683, and CRS3_HUMAN]. Accordingly, it is presently believed that PRO328 polypeptide disclosed in the present application is a newly identified member of the GLIP or CRISP families and possesses transcriptional regulatory activity typical of the GLIP or CRISP families.

42. Full-length PRO335, PRO331 and PRO326 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO335, PRO331 or PRO326. In particular, Applicants have identified and isolated cDNA encoding a PRO335, PRO331 or PRO326 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO335, PRO331 or PRO326 polypeptide have significant homology with LIG-1, ALS and in the case of PRO331, additionally, decorin. Accordingly, it is presently believed that the PRO335, PRO331 and PRO326 polypeptides disclosed in the present application are newly identified members of the leucine rich repeat superfamily, and particularly, are related to LIG-1 and possess the biological functions of this family as discussed and referenced herein.

43. Full-length PRO332 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO332. In particular, Applicants have identified and isolated cDNA encoding PRO332 polypeptides, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO332 (shown in Figure 108 and SEQ ID NO:310) has about 30-40% amino acid sequence identity with a series of known proteoglycan sequences, including, for example, fibromodulin and fibromodulin precursor sequences of various species (FMOD_BOVIN, FMOD_CHICK, FMOD_RAT, FMOD_MOUSE, FMOD_HUMAN, P_R36773), osteomodulin sequences (AB000114_1, AB007848_1), decorin sequences (CFU83141_1, OCU03394_1, P R42266, P_R42267, P_R42260, P_R89439), keratan sulfate proteoglycans (BTU48360_1, AF022890_1), corneal proteoglycan (AF022256_1), and bone/cartilage proteoglycans and proteoglycane precursors (PGS1_BOVIN, PGS2_ MOUSE, PGS2_HUMAN). Accordingly, it is presently believed that PRO332 disclosed in the present application is a new proteoglycan-type molecule, and may play a role in regulating extracellular matrix, cartilage, and/or bone function.

44. Full-length PRO334 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO334. In particular, Applicants have identified and isolated cDNA encoding a PRO334 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO334 polypeptide have significant homology with fibulin and fibrillin. Accordingly, it is presently believed that PRO334 polypeptide disclosed in the present application is a newly identified member of the epidermal growth factor family and possesses properties and activities typical of this family.

45. Full-length PRO346 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO346. In particular, Applicants have identified and isolated cDNA encoding a PRO346 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO346 (shown in Figure 112 and SEQ ID NO:320) has 28 % amino acid sequence identity with carcinoembryonic antigen. Accordingly, it is presently believed that PRO346 disclosed in the present application is a newly identified member of the carcinoembryonic protein family and may be expressed in association with neoplastic tissue disorders.

46. Full-length PRO268 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO268. In particular, Applicants have identified and isolated cDNA encoding a PRO268 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that portions of the PRO268 polypeptide have significant homology with the various protein disulfide isomerase proteins. Accordingly, it is presently believed

that PRO268 polypeptide disclosed in the present application is a homolog of the protein disulfide isomerase p5 protein.

47. Full-length PRO330 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO330. In particular, Applicants have identified and isolated cDNA encoding a PRO330 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO330 polypeptide have significant homology with the murine prolyl 4-hydroxylase alpha-II subunit protein. Accordingly, it is presently believed that PRO330 polypeptide disclosed in the present application is a novel prolyl 4-hydroxylase subunit polypeptide.

48. Full-length PRO339 and PRO310 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO339 and PRO310. In particular, Applicants have identified and isolated cDNA encoding a PRO339 polypeptide, as disclosed in further detail in the Examples below. Applicants have also identified and isolated cDNA encoding a PRO310 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO339 and PRO310 polypeptides have significant homology with small secreted proteins from *C. elegans* and are distantly related to fringe. PRO339 also shows homology to collagen-like polymers. Sequences which were used to identify PRO310, designated herein as DNA40533 and DNA42267, also show homology to proteins from *C. elegans*. Accordingly, it is presently believed that the PRO339 and PRO310 polypeptides disclosed in the present application are newly identified member of the family of proteins involved in development, and which may have regulatory abilities similar to the capability of fringe to regulate serrate.

49. Full Length PRO244 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding C-type lectins referred to in the present application as PRO244. In particular, applicants have identified and isolated cDNA encoding PRO244 polypeptides, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that a full-length native sequence PRO244 (shown in Figure 122 and SEQ ID NO:377) has 43% amino acid sequence identity with the hepatic lectin gallus gallus (LECH-CHICK), and 42% amino acid sequence identity with an HIV gp120 binding C-type lectin (A46274). Accordingly, it is presently believed that PRO244 disclosed in the present application is a newly identified member of the C-lectin superfamily and may play a role in immune function, apoptosis, or in the pathogenesis of atherosclerosis. In addition, PRO244 may be useful in identifying tumor-associated epitopes.

B. PRO Polypeptide Variants

In addition to the full-length native sequence PRO polypeptides described herein, it is contemplated that

PRO variants can be prepared. PRO variants can be prepared by introducing appropriate nucleotide changes into the PRO DNA, and/or by synthesis of the desired PRO polypeptide. Those skilled in the art will appreciate that amino acid changes may alter post-translational processes of the PRO, such as changing the number or position of glycosylation sites or altering the membrane anchoring characteristics.

Variations in the native full-length sequence PRO or in various domains of the PRO described herein, can be made, for example, using any of the techniques and guidelines for conservative and non-conservative mutations set forth, for instance, in U.S. Patent No. 5,364,934. Variations may be a substitution, deletion or insertion of one or more codons encoding the PRO that results in a change in the amino acid sequence of the PRO as compared with the native sequence PRO. Optionally the variation is by substitution of at least one amino acid with any other amino acid in one or more of the domains of the PRO. Guidance in determining which amino acid residue may be inserted, substituted or deleted without adversely affecting the desired activity may be found by comparing the sequence of the PRO with that of homologous known protein molecules and minimizing the number of amino acid sequence changes made in regions of high homology. Amino acid substitutions can be the result of replacing one amino acid with another amino acid having similar structural and/or chemical properties, such as the replacement of a leucine with a serine, i.e., conservative amino acid replacements. Insertions or deletions may optionally be in the range of about 1 to 5 amino acids. The variation allowed may be determined by systematically making insertions, deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity exhibited by the full-length or mature native sequence.

PRO polypeptide fragments are provided herein. Such fragments may be truncated at the N-terminus or C-terminus, or may lack internal residues, for example, when compared with a full length native protein. Certain fragments lack amino acid residues that are not essential for a desired biological activity of the PRO polypeptide.

PRO fragments may be prepared by any of a number of conventional techniques. Desired peptide fragments may be chemically synthesized. An alternative approach involves generating PRO fragments by enzymatic digestion, e.g., by treating the protein with an enzyme known to cleave proteins at sites defined by particular amino acid residues, or by digesting the DNA with suitable restriction enzymes and isolating the desired fragment. Yet another suitable technique involves isolating and amplifying a DNA fragment encoding a desired polypeptide fragment, by polymerase chain reaction (PCR). Oligonucleotides that define the desired termini of the DNA fragment are employed at the 5' and 3' primers in the PCR. Preferably, PRO polypeptide fragments share at least one biological and/or immunological activity with the native PRO polypeptide disclosed herein.

In particular embodiments, conservative substitutions of interest are shown in Table 6 under the heading of preferred substitutions. If such substitutions result in a change in biological activity, then more substantial changes, denominated exemplary substitutions in Table 6, or as further described below in reference to amino acid classes, are introduced and the products screened.

Table 6

Original Residue	Exemplary Substitutions	Preferred Substitutions
Ala (A)	val; leu; ile	val
Arg (R)	lys; gln; asn	lys
Asn (N)	gln; his; lys; arg	gln
Asp (D)	glu	glu
Cys (C)	ser	ser
Gln (Q)	asn	asn
Glu (E)	asp	asp
Gly (G)	pro; ala	ala
His (H)	asn; gln; lys; arg	arg
Ile (I)	leu; val; met; ala; phe; norleucine	leu
Leu (L)	norleucine; ile; val; met; ala; phe	ile
Lys (K)	arg; gln; asn	arg
Met (M)	leu; phe; ile	leu
Phe (F)	leu; val; ile; ala; tyr	leu
Pro (P)	ala	ala
Ser (S)	thr	thr
Thr (T)	ser	ser
Trp (W)	tyr; phe	tyr
Tyr (Y)	trp; phe; thr; ser	phe
Val (V)	ile; leu; met; phe; ala; norleucine	leu

Substantial modifications in function or immunological identity of the PRO polypeptide are accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues are divided into groups based on common side-chain properties:

(1) hydrophobic: norleucine, met, ala, val, leu, ile;

(2) neutral hydrophilic: cys, ser, thr;

(3) acidic: asp, glu;

(4) basic: asn, gln, his, lys, arg;

(5) residues that influence chain orientation: gly, pro; and

(6) aromatic: trp, tyr, phe.

Non-conservative substitutions will entail exchanging a member of one of these classes for another class. Such substituted residues also may be introduced into the conservative substitution sites or, more preferably, into the remaining (non-conserved) sites.

The variations can be made using methods known in the art such as oligonucleotide-mediated (site-directed) mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis [Carter et al., Nucl. Acids Res., 13:4331 (1986); Zoller et al., Nucl. Acids Res., 10:6487 (1987)], cassette mutagenesis [Wells et al., Gene, 34:315 (1985)], restriction selection mutagenesis [Wells et al., Philos. Trans. R. Soc. London SerA, 317:415 (1986)] or other known techniques can be performed on the cloned DNA to produce the PRO variant

DNA.

Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous sequence. Among the preferred scanning amino acids are relatively small, neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant [Cunningham and Wells, Science, 244: 1081-1085 (1989)]. Alanine is also typically preferred because it is the most common amino acid. Further, it is frequently found in both buried and exposed positions [Creighton, The Proteins, (W.H. Freeman & Co., N.Y.); Chothia, J. Mol. Biol., 150:1 (1976)]. If alanine substitution does not yield adequate amounts of variant, an isoteric amino acid can be used.

C. Modifications of PRO

Covalent modifications of PRO are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of a PRO polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C- terminal residues of the PRO. Derivatization with bifunctional agents is useful, for instance, for crosslinking PRO to a water-insoluble support matrix or surface for use in the method for purifying anti-PRO antibodies, and vice-versa. Commonly used crosslinking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4-azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylpropionate), bifunctional maleimides such as bis-N-maleimido-1,8-octane and agents such as methyl-3-[(p-azidophenyl)dithio]propioimidate.

Other modifications include deamidation of glutaminyl and asparaginyl residues to the corresponding glutamyl and aspartyl residues, respectively, hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α-amino groups of lysine, arginine, and histidine side chains [T.E. Creighton, Proteins: Structure and Molecular Properties, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)], acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

Another type of covalent modification of the PRO polypeptide included within the scope of this invention comprises altering the native glycosylation pattern of the polypeptide. "Altering the native glycosylation pattern" is intended for purposes herein to mean deleting one or more carbohydrate moieties found in native sequence PRO (either by removing the underlying glycosylation site or by deleting the glycosylation by chemical and/or enzymatic means), and/or adding one or more glycosylation sites that are not present in the native sequence PRO. In addition, the phrase includes qualitative changes in the glycosylation of the native proteins, involving a change in the nature and proportions of the various carbohydrate moieties present.

Addition of glycosylation sites to the PRO polypeptide may be accomplished by altering the amino acid sequence. The alteration may be made, for example, by the addition of, or substitution by, one or more serine or threonine residues to the native sequence PRO (for O-linked glycosylation sites). The PRO amino acid sequence may optionally be altered through changes at the DNA level, particularly by mutating the DNA encoding the PRO polypeptide at preselected bases such that codons are generated that will translate into the desired amino acids.

Another means of increasing the number of carbohydrate moieties on the PRO polypeptide is by

chemical or enzymatic coupling of glycosides to the polypeptide. Such methods are described in the art, e.g., in WO 87/05330 published 11 September 1987, and in Aplin and Wriston, CRC Crit. Rev. Biochem., pp. 259-306 (1981).

Removal of carbohydrate moieties present on the PRO polypeptide may be accomplished chemically or enzymatically or by mutational substitution of codons encoding for amino acid residues that serve as targets for glycosylation. Chemical deglycosylation techniques are known in the art and described, for instance, by Hakimuddin, et al., Arch. Biochem. Biophys., 259:52 (1987) and by Edge et al., Anal. Biochem., 118:131 (1981). Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura et al., Meth. Enzymol., 138:350 (1987).

Another type of covalent modification of PRO comprises linking the PRO polypeptide to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol (PEG), polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

The PRO of the present invention may also be modified in a way to form a chimeric molecule comprising PRO fused to another, heterologous polypeptide or amino acid sequence.

In one embodiment, such a chimeric molecule comprises a fusion of the PRO with a tag polypeptide which provides an epitope to which an anti-tag antibody can selectively bind. The epitope tag is generally placed at the amino- or carboxyl- terminus of the PRO. The presence of such epitope-tagged forms of the PRO can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables the PRO to be readily purified by affinity purification using an anti-tag antibody or another type of affinity matrix that binds to the epitope tag. Various tag polypeptides and their respective antibodies are well known in the art. Examples include poly-histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags; the flu HA tag polypeptide and its antibody 12CA5 [Field et al., Mol. Cell. Biol., 8:2159-2165 (1988)]; the c-myc tag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibodies thereto [Evan et al., Molecular and Cellular Biology, 5:3610-3616 (1985)]; and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody [Paborsky et al., Protein Engineering, 3(6):547-553 (1990)]. Other tag polypeptides include the Flag-peptide [Hopp et al., BioTechnology, 6:1204-1210 (1988)]; the KT3 epitope peptide [Martin et al., Science, 255:192-194 (1992)]; an α-tubulin epitope peptide [Skinner et al., J. Biol. Chem., 266:15163-15166 (1991)]; and the T7 gene 10 protein peptide tag [Lutz-Freyermuth et al., Proc. Natl. Acad. Sci. USA, 87:6393-6397 (1990)].

In an alternative embodiment, the chimeric molecule may comprise a fusion of the PRO with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule (also referred to as an "immunoadhesin"), such a fusion could be to the Fc region of an IgG molecule. The Ig fusions preferably include the substitution of a soluble (transmembrane domain deleted or inactivated) form of a PRO polypeptide in place of at least one variable region within an Ig molecule. In a particularly preferred embodiment, the immunoglobulin fusion includes the hinge, CH2 and CH3, or the hinge, CH1, CH2 and CH3 regions of an IgG1 molecule. For the production of immunoglobulin fusions see also US Patent No. 5,428,130 issued June 27, 1995.

D. Preparation of PRO

The description below relates primarily to production of PRO by culturing cells transformed or

transfected with a vector containing PRO nucleic acid. It is, of course, contemplated that alternative methods, which are well known in the art, may be employed to prepare PRO. For instance, the PRO sequence, or portions thereof, may be produced by direct peptide synthesis using solid-phase techniques [see, e.g., Stewart et al., Solid-Phase Peptide Synthesis, W.H. Freeman Co., San Francisco, CA (1969); Merrifield, J. Am. Chem. Soc., 85:2149-2154 (1963)]. *In vitro* protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be accomplished, for instance, using an Applied Biosystems Peptide Synthesizer (Foster City, CA) using manufacturer's instructions. Various portions of the PRO may be chemically synthesized separately and combined using chemical or enzymatic methods to produce the full-length PRO.

1. Isolation of DNA Encoding PRO

DNA encoding PRO may be obtained from a cDNA library prepared from tissue believed to possess the PRO mRNA and to express it at a detectable level. Accordingly, human PRO DNA can be conveniently obtained from a cDNA library prepared from human tissue, such as described in the Examples. The PRO-encoding gene may also be obtained from a genomic library or by known synthetic procedures (e.g., automated nucleic acid synthesis).

Libraries can be screened with probes (such as antibodies to the PRO or oligonucleotides of at least about 20-80 bases) designed to identify the gene of interest or the protein encoded by it. Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the gene encoding PRO is to use PCR methodology [Sambrook et al., supra; Dieffenbach et al., PCR Primer: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1995)].

The Examples below describe techniques for screening a cDNA library. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized. The oligonucleotide is preferably labeled such that it can be detected upon hybridization to DNA in the library being screened. Methods of labeling are well known in the art, and include the use of radiolabels like ^{32}P-labeled ATP, biotinylation or enzyme labeling. Hybridization conditions, including moderate stringency and high stringency, are provided in Sambrook et al., supra.

Sequences identified in such library screening methods can be compared and aligned to other known sequences deposited and available in public databases such as GenBank or other private sequence databases. Sequence identity (at either the amino acid or nucleotide level) within defined regions of the molecule or across the full-length sequence can be determined using methods known in the art and as described herein.

Nucleic acid having protein coding sequence may be obtained by screening selected cDNA or genomic libraries using the deduced amino acid sequence disclosed herein for the first time, and, if necessary, using conventional primer extension procedures as described in Sambrook et al., supra, to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

2. Selection and Transformation of Host Cells

Host cells are transfected or transformed with expression or cloning vectors described herein for PRO

production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in Mammalian Cell Biotechnology: a Practical Approach, M. Butler, ed. (IRL Press, 1991) and Sambrook et al., supra.

Methods of eukaryotic cell transfection and prokaryotic cell transformation are known to the ordinarily skilled artisan, for example, $CaCl_2$, $CaPO_4$, liposome-mediated and electroporation. Depending on the host cell used, transformation is performed using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in Sambrook et al., supra, or electroporation is generally used for prokaryotes. Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) can be employed. General aspects of mammalian cell host system transfections have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, e.g., polybrene, polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown et al., Methods in Enzymology, 185:527-537 (1990) and Mansour et al., Nature, 336:348-352 (1988).

Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as *E. coli*. Various *E. coli* strains are publicly available, such as *E. coli* K12 strain MM294 (ATCC 31,446); *E. coli* X1776 (ATCC 31,537); *E. coli* strain W3110 (ATCC 27,325) and K5 772 (ATCC 53,635). Other suitable prokaryotic host cells include Enterobacteriaceae such as *Escherichia*, e.g., *E. coli*, *Enterobacter*, *Erwinia*, *Klebsiella*, *Proteus*, *Salmonella*, e.g., *Salmonella typhimurium*, *Serratia*, e.g., *Serratia marcescans*, and *Shigella*, as well as *Bacilli* such as *B. subtilis* and *B. licheniformis* (e.g., *B. licheniformis* 41P disclosed in DD 266,710 published 12 April 1989), *Pseudomonas* such as *P. aeruginosa*, and *Streptomyces*. These examples are illustrative rather than limiting. Strain W3110 is one particularly preferred host or parent host because it is a common host strain for recombinant DNA product fermentations. Preferably, the host cell secretes minimal amounts of proteolytic enzymes. For example, strain W3110 may be modified to effect a genetic mutation in the genes encoding proteins endogenous to the host, with examples of such hosts including *E. coli* W3110 strain 1A2, which has the complete genotype *tonA* ; *E. coli* W3110 strain 9E4, which has the complete genotype *tonA ptr3*; *E. coli* W3110 strain 27C7 (ATCC 55,244), which has the complete genotype *tonA ptr3 phoA E15 (argF-lac)169 degP ompT kan*^{r}; *E. coli* W3110 strain 37D6, which has the complete genotype *tonA ptr3 phoA E15 (argF-lac)169 degP ompT rbs7 ilvG kan*^{r}; *E. coli* W3110 strain 40B4, which is strain 37D6 with a non-kanamycin resistant *degP* deletion mutation; and an *E. coli* strain having mutant periplasmic protease disclosed in U.S. Patent No. 4,946,783 issued 7 August 1990. Alternatively, *in vitro* methods of cloning, e.g., PCR or other nucleic acid polymerase

reactions, are suitable.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for PRO-encoding vectors. *Saccharomyces cerevisiae* is a commonly used lower eukaryotic host microorganism. Others include *Schizosaccharomyces pombe* (Beach and Nurse, Nature, 290: 140 [1981]; EP 139,383 published 2 May 1985); *Kluyveromyces* hosts (U.S. Patent No. 4,943,529; Fleer et al., Bio/Technology, 9:968-975 (1991)) such as, e.g., *K. lactis* (MW98-8C, CBS683, CBS4574; Louvencourt et al., J. Bacteriol., 737 [1983]), *K. fragilis* (ATCC 12,424), *K. bulgaricus* (ATCC 16,045), *K. wickeramii* (ATCC 24,178), *K. waltii* (ATCC 56,500), *K. drosophilarum* (ATCC 36,906; Van den Berg et al., Bio/Technology, 8:135 (1990)), *K. thermotolerans*, and *K. marxianus; yarrowia* (EP 402,226); *Pichia pastoris* (EP 183,070; Sreekrishna et al., J. Basic Microbiol., 28:265-278 [1988]); *Candida; Trichoderma reesia* (EP 244,234); *Neurospora crassa* (Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 [1979]); *Schwanniomyces* such as *Schwanniomyces occidentalis* (EP 394,538 published 31 October 1990); and filamentous fungi such as, e.g., *Neurospora, Penicillium, Tolypocladium* (WO 91/00357 published 10 January 1991), and *Aspergillus* hosts such as *A. nidulans* (Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 [1983]; Tilburn et al., Gene, 26:205-221 [1983]; Yelton et al., Proc. Natl. Acad. Sci. USA, 81: 1470-1474 [1984]) and *A. niger* (Kelly and Hynes, EMBO J., 4:475-479 [1985]). Methylotropic yeasts are suitable herein and include, but are not limited to, yeast capable of growth on methanol selected from the genera consisting of *Hansenula, Candida, Kloeckera, Pichia, Saccharomyces, Torulopsis,* and *Rhodotorula*. A list of specific species that are exemplary of this class of yeasts may be found in C. Anthony, The Biochemistry of Methylotrophs, 269 (1982).

Suitable host cells for the expression of glycosylated PRO are derived from multicellular organisms. Examples of invertebrate cells include insect cells such as Drosophila S2 and Spodoptera Sf9, as well as plant cells. Examples of useful mammalian host cell lines include Chinese hamster ovary (CHO) and COS cells. More specific examples include monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 (1977)); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 (1980)); mouse sertoli cells (TM4, Mather, Biol. Reprod., 23:243-251 (1980)); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); and mouse mammary tumor (MMT 060562, ATCC CCL51). The selection of the appropriate host cell is deemed to be within the skill in the art.

3. Selection and Use of a Replicable Vector

The nucleic acid (e.g., cDNA or genomic DNA) encoding PRO may be inserted into a replicable vector for cloning (amplification of the DNA) or for expression. Various vectors are publicly available. The vector may, for example, be in the form of a plasmid, cosmid, viral particle, or phage. The appropriate nucleic acid sequence may be inserted into the vector by a variety of procedures. In general, DNA is inserted into an appropriate restriction endonuclease site(s) using techniques known in the art. Vector components generally include, but are not limited to, one or more of a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Construction of suitable vectors containing one or more of these components employs standard ligation techniques which are known to the skilled artisan.

The PRO may be produced recombinantly not only directly, but also as a fusion polypeptide with a heterologous polypeptide, which may be a signal sequence or other polypeptide having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the vector, or it may be a part of the PRO-encoding DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α-factor leaders, the latter described in U.S. Patent No. 5,010,182), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells.

Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for *Bacilli*.

An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the PRO-encoding nucleic acid, such as DHFR or thymidine kinase. An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described by Urlaub et al., Proc. Natl. Acad. Sci. USA, 77:4216 (1980). A suitable selection gene for use in yeast is the *trp*1 gene present in the yeast plasmid YRp7 [Stinchcomb et al., Nature, 282:39 (1979); Kingsman et al., Gene, 7:141 (1979); Tschemper et al., Gene, 10:157 (1980)]. The *trp*1 gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEP4-1 [Jones, Genetics, 85:12 (1977)].

Expression and cloning vectors usually contain a promoter operably linked to the PRO-encoding nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a variety of potential host cells are well known. Promoters suitable for use with prokaryotic hosts include the β-lactamase and lactose promoter systems [Chang et al., Nature, 275:615 (1978); Goeddel et al., Nature, 281:544 (1979)], alkaline phosphatase, a tryptophan (trp) promoter system [Goeddel, Nucleic Acids Res., 8:4057 (1980); EP 36,776], and hybrid promoters such as the tac promoter [deBoer et al., Proc. Natl. Acad. Sci. USA, 80:21-25 (1983)]. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding PRO.

Examples of suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase [Hitzeman et al., J. Biol. Chem., 255:2073 (1980)] or other glycolytic enzymes [Hess

et al., J. Adv. Enzyme Reg., 7:149 (1968); Holland, Biochemistry, 17:4900 (1978)], such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in EP 73,657.

PRO transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g., the actin promoter or an immunoglobulin promoter, and from heat-shock promoters, provided such promoters are compatible with the host cell systems.

Transcription of a DNA encoding the PRO by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α-fetoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. The enhancer may be spliced into the vector at a position 5' or 3' to the PRO coding sequence, but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3', untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding PRO.

Still other methods, vectors, and host cells suitable for adaptation to the synthesis of PRO in recombinant vertebrate cell culture are described in Gething et al., Nature, 293:620-625 (1981); Mantei et al., Nature, 281:40-46 (1979); EP 117,060; and EP 117,058.

4. Detecting Gene Amplification/Expression

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA [Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 (1980)], dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes

or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of cells or tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal, and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a native sequence PRO polypeptide or against a synthetic peptide based on the DNA sequences provided herein or against exogenous sequence fused to PRO DNA and encoding a specific antibody epitope.

5. Purification of Polypeptide

Forms of PRO may be recovered from culture medium or from host cell lysates. If membrane-bound, it can be released from the membrane using a suitable detergent solution (e.g. Triton-X 100) or by enzymatic cleavage. Cells employed in expression of PRO can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

It may be desired to purify PRO from recombinant cell proteins or polypeptides. The following procedures are exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex G-75; protein A Sepharose columns to remove contaminants such as IgG; and metal chelating columns to bind epitope-tagged forms of the PRO. Various methods of protein purification may be employed and such methods are known in the art and described for example in Deutscher, Methods in Enzymology, 182 (1990); Scopes, Protein Purification: Principles and Practice, Springer-Verlag, New York (1982). The purification step(s) selected will depend, for example, on the nature of the production process used and the particular PRO produced.

E. Uses for PRO

Nucleotide sequences (or their complement) encoding PRO have various applications in the art of molecular biology, including uses as hybridization probes, in chromosome and gene mapping and in the generation of anti-sense RNA and DNA. PRO nucleic acid will also be useful for the preparation of PRO polypeptides by the recombinant techniques described herein.

The full-length native sequence PRO gene, or portions thereof, may be used as hybridization probes for a cDNA library to isolate the full-length PRO cDNA or to isolate still other cDNAs (for instance, those encoding naturally-occurring variants of PRO or PRO from other species) which have a desired sequence identity to the native PRO sequence disclosed herein. Optionally, the length of the probes will be about 20 to about 50 bases. The hybridization probes may be derived from at least partially novel regions of the full length native nucleotide sequence wherein those regions may be determined without undue experimentation or from genomic sequences including promoters, enhancer elements and introns of native sequence PRO. By way of example,

a screening method will comprise isolating the coding region of the PRO gene using the known DNA sequence to synthesize a selected probe of about 40 bases. Hybridization probes may be labeled by a variety of labels, including radionucleotides such as ^{32}P or ^{35}S, or enzymatic labels such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems. Labeled probes having a sequence complementary to that of the PRO gene of the present invention can be used to screen libraries of human cDNA, genomic DNA or mRNA to determine which members of such libraries the probe hybridizes to. Hybridization techniques are described in further detail in the Examples below.

Any EST sequences disclosed in the present application may similarly be employed as probes, using the methods disclosed herein.

Other useful fragments of the PRO nucleic acids include antisense or sense oligonucleotides comprising a singe-stranded nucleic acid sequence (either RNA or DNA) capable of binding to target PRO mRNA (sense) or PRO DNA (antisense) sequences. Antisense or sense oligonucleotides, according to the present invention, comprise a fragment of the coding region of PRO DNA. Such a fragment generally comprises at least about 14 nucleotides, preferably from about 14 to 30 nucleotides. The ability to derive an antisense or a sense oligonucleotide, based upon a cDNA sequence encoding a given protein is described in, for example, Stein and Cohen (Cancer Res. 48:2659, 1988) and van der Krol et al. (BioTechniques 6:958, 1988).

Binding of antisense or sense oligonucleotides to target nucleic acid sequences results in the formation of duplexes that block transcription or translation of the target sequence by one of several means, including enhanced degradation of the duplexes, premature termination of transcription or translation, or by other means. The antisense oligonucleotides thus may be used to block expression of PRO proteins. Antisense or sense oligonucleotides further comprise oligonucleotides having modified sugar-phosphodiester backbones (or other sugar linkages, such as those described in WO 91/06629) and wherein such sugar linkages are resistant to endogenous nucleases. Such oligonucleotides with resistant sugar linkages are stable *in vivo* (i.e., capable of resisting enzymatic degradation) but retain sequence specificity to be able to bind to target nucleotide sequences.

Other examples of sense or antisense oligonucleotides include those oligonucleotides which are covalently linked to organic moieties, such as those described in WO 90/10048, and other moieties that increases affinity of the oligonucleotide for a target nucleic acid sequence, such as poly-(L-lysine). Further still, intercalating agents, such as ellipticine, and alkylating agents or metal complexes may be attached to sense or antisense oligonucleotides to modify binding specificities of the antisense or sense oligonucleotide for the target nucleotide sequence.

Antisense or sense oligonucleotides may be introduced into a cell containing the target nucleic acid sequence by any gene transfer method, including, for example, $CaPO_4$-mediated DNA transfection, electroporation, or by using gene transfer vectors such as Epstein-Barr virus. In a preferred procedure, an antisense or sense oligonucleotide is inserted into a suitable retroviral vector. A cell containing the target nucleic acid sequence is contacted with the recombinant retroviral vector, either *in vivo* or *ex vivo*. Suitable retroviral vectors include, but are not limited to, those derived from the murine retrovirus M-MuLV, N2 (a retrovirus derived from M-MuLV), or the double copy vectors designated DCT5A, DCT5B and DCT5C (see WO 90/13641).

Sense or antisense oligonucleotides also may be introduced into a cell containing the target nucleotide sequence by formation of a conjugate with a ligand binding molecule, as described in WO 91/04753. Suitable ligand binding molecules include, but are not limited to, cell surface receptors, growth factors, other cytokines, or other ligands that bind to cell surface receptors. Preferably, conjugation of the ligand binding molecule does not substantially interfere with the ability of the ligand binding molecule to bind to its corresponding molecule or receptor, or block entry of the sense or antisense oligonucleotide or its conjugated version into the cell.

Alternatively, a sense or an antisense oligonucleotide may be introduced into a cell containing the target nucleic acid sequence by formation of an oligonucleotide-lipid complex, as described in WO 90/10448. The sense or antisense oligonucleotide-lipid complex is preferably dissociated within the cell by an endogenous lipase.

Antisense RNA or DNA molecules are generally at least about 5 bases in length, about 10 bases in length, about 15 bases in length, about 20 bases in length, about 25 bases in length, about 30 bases in length, about 35 bases in length, about 40 bases in length, about 45 bases in length, about 50 bases in length, about 55 bases in length, about 60 bases in length, about 65 bases in length, about 70 bases in length, about 75 bases in length, about 80 bases in length, about 85 bases in length, about 90 bases in length, about 95 bases in length, about 100 bases in length, or more.

The probes may also be employed in PCR techniques to generate a pool of sequences for identification of closely related PRO coding sequences.

Nucleotide sequences encoding a PRO can also be used to construct hybridization probes for mapping the gene which encodes that PRO and for the genetic analysis of individuals with genetic disorders. The nucleotide sequences provided herein may be mapped to a chromosome and specific regions of a chromosome using known techniques, such as *in situ* hybridization, linkage analysis against known chromosomal markers, and hybridization screening with libraries.

When the coding sequences for PRO encode a protein which binds to another protein (example, where the PRO is a receptor), the PRO can be used in assays to identify the other proteins or molecules involved in the binding interaction. By such methods, inhibitors of the receptor/ligand binding interaction can be identified. Proteins involved in such binding interactions can also be used to screen for peptide or small molecule inhibitors or agonists of the binding interaction. Also, the receptor PRO can be used to isolate correlative ligand(s). Screening assays can be designed to find lead compounds that mimic the biological activity of a native PRO or a receptor for PRO. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates. Small molecules contemplated include synthetic organic or inorganic compounds. The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays and cell based assays, which are well characterized in the art.

Nucleic acids which encode PRO or its modified forms can also be used to generate either transgenic animals or "knock out" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. A transgenic animal (e.g., a mouse or rat) is an animal having cells that contain a transgene, which transgene was introduced into the animal or an ancestor of the animal at a prenatal, e.g., an embryonic stage. A transgene is a DNA which is integrated into the genome of a cell from which a transgenic animal develops. In one embodiment, cDNA encoding PRO can be used to clone genomic DNA encoding PRO in

accordance with established techniques and the genomic sequences used to generate transgenic animals that contain cells which express DNA encoding PRO. Methods for generating transgenic animals, particularly animals such as mice or rats, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009. Typically, particular cells would be targeted for PRO transgene incorporation with tissue-specific enhancers. Transgenic animals that include a copy of a transgene encoding PRO introduced into the germ line of the animal at an embryonic stage can be used to examine the effect of increased expression of DNA encoding PRO. Such animals can be used as tester animals for reagents thought to confer protection from, for example, pathological conditions associated with its overexpression. In accordance with this facet of the invention, an animal is treated with the reagent and a reduced incidence of the pathological condition, compared to untreated animals bearing the transgene, would indicate a potential therapeutic intervention for the pathological condition.

Alternatively, non-human homologues of PRO can be used to construct a PRO "knock out" animal which has a defective or altered gene encoding PRO as a result of homologous recombination between the endogenous gene encoding PRO and altered genomic DNA encoding PRO introduced into an embryonic stem cell of the animal. For example, cDNA encoding PRO can be used to clone genomic DNA encoding PRO in accordance with established techniques. A portion of the genomic DNA encoding PRO can be deleted or replaced with another gene, such as a gene encoding a selectable marker which can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector [see e.g., Thomas and Capecchi, Cell, 51:503 (1987) for a description of homologous recombination vectors]. The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected [see e.g., Li et al., Cell, 69:915 (1992)]. The selected cells are then injected into a blastocyst of an animal (e.g., a mouse or rat) to form aggregation chimeras [see e.g., Bradley, in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E. J. Robertson, ed. (IRL, Oxford, 1987), pp. 113-152]. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term to create a "knock out" animal. Progeny harboring the homologously recombined DNA in their germ cells can be identified by standard techniques and used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knockout animals can be characterized for instance, for their ability to defend against certain pathological conditions and for their development of pathological conditions due to absence of the PRO polypeptide.

Nucleic acid encoding the PRO polypeptides may also be used in gene therapy. In gene therapy applications, genes are introduced into cells in order to achieve *in vivo* synthesis of a therapeutically effective genetic product, for example for replacement of a defective gene. "Gene therapy" includes both conventional gene therapy where a lasting effect is achieved by a single treatment, and the administration of gene therapeutic agents, which involves the one time or repeated administration of a therapeutically effective DNA or mRNA. Antisense RNAs and DNAs can be used as therapeutic agents for blocking the expression of certain genes *in vivo*. It has already been shown that short antisense oligonucleotides can be imported into cells where they act as inhibitors, despite their low intracellular concentrations caused by their restricted uptake by the cell membrane. (Zamecnik *et al.*, Proc. Natl. Acad. Sci. USA 83:4143-4146 [1986]). The oligonucleotides can be

modified to enhance their uptake, e.g. by substituting their negatively charged phosphodiester groups by uncharged groups.

There are a variety of techniques available for introducing nucleic acids into viable cells. The techniques vary depending upon whether the nucleic acid is transferred into cultured cells *in vitro*, or *in vivo* in the cells of the intended host. Techniques suitable for the transfer of nucleic acid into mammalian cells *in vitro* include the use of liposomes, electroporation, microinjection, cell fusion, DEAE-dextran, the calcium phosphate precipitation method, etc. The currently preferred *in vivo* gene transfer techniques include transfection with viral (typically retroviral) vectors and viral coat protein-liposome mediated transfection (Dzau et al., Trends in Biotechnology 11, 205-210 [1993]). In some situations it is desirable to provide the nucleic acid source with an agent that targets the target cells, such as an antibody specific for a cell surface membrane protein or the target cell, a ligand for a receptor on the target cell, etc. Where liposomes are employed, proteins which bind to a cell surface membrane protein associated with endocytosis may be used for targeting and/or to facilitate uptake, e.g. capsid proteins or fragments thereof tropic for a particular cell type, antibodies for proteins which undergo internalization in cycling, proteins that target intracellular localization and enhance intracellular half-life. The technique of receptor-mediated endocytosis is described, for example, by Wu et al., J. Biol. Chem. 262, 4429-4432 (1987); and Wagner et al., Proc. Natl. Acad. Sci. USA 87, 3410-3414 (1990). For review of gene marking and gene therapy protocols see Anderson et al., Science 256, 808-813 (1992).

The PRO polypeptides described herein may also be employed as molecular weight markers for protein electrophoresis purposes and the isolated nucleic acid sequences may be used for recombinantly expressing those markers.

The nucleic acid molecules encoding the PRO polypeptides or fragments thereof described herein are useful for chromosome identification. In this regard, there exists an ongoing need to identify new chromosome markers, since relatively few chromosome marking reagents, based upon actual sequence data are presently available. Each PRO nucleic acid molecule of the present invention can be used as a chromosome marker.

The PRO polypeptides and nucleic acid molecules of the present invention may also be used for tissue typing, wherein the PRO polypeptides of the present invention may be differentially expressed in one tissue as compared to another. PRO nucleic acid molecules will find use for generating probes for PCR, Northern analysis, Southern analysis and Western analysis.

The PRO polypeptides described herein may also be employed as therapeutic agents. The PRO polypeptides of the present invention can be formulated according to known methods to prepare pharmaceutically useful compositions, whereby the PRO product hereof is combined in admixture with a pharmaceutically acceptable carrier vehicle. Therapeutic formulations are prepared for storage by mixing the active ingredient having the desired degree of purity with optional physiologically acceptable carriers, excipients or stabilizers (Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980)), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone, amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides and other

carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN™, PLURONICS™ or PEG.

The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes, prior to or following lyophilization and reconstitution.

Therapeutic compositions herein generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle.

The route of administration is in accord with known methods, e.g. injection or infusion by intravenous, intraperitoneal, intracerebral, intramuscular, intraocular, intraarterial or intralesional routes, topical administration, or by sustained release systems.

Dosages and desired drug concentrations of pharmaceutical compositions of the present invention may vary depending on the particular use envisioned. The determination of the appropriate dosage or route of administration is well within the skill of an ordinary physician. Animal experiments provide reliable guidance for the determination of effective doses for human therapy. Interspecies scaling of effective doses can be performed following the principles laid down by Mordenti, J. and Chappell, W. "The use of interspecies scaling in toxicokinetics" In Toxicokinetics and New Drug Development, Yacobi et al., Eds., Pergamon Press, New York 1989, pp. 42-96.

When *in vivo* administration of a PRO polypeptide or agonist or antagonist thereof is employed, normal dosage amounts may vary from about 10 ng/kg to up to 100 mg/kg of mammal body weight or more per day, preferably about 1 μg/kg/day to 10 mg/kg/day, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature; see, for example, U.S. Pat. Nos. 4,657,760; 5,206,344; or 5,225,212. It is anticipated that different formulations will be effective for different treatment compounds and different disorders, that administration targeting one organ or tissue, for example, may necessitate delivery in a manner different from that to another organ or tissue.

Where sustained-release administration of a PRO polypeptide is desired in a formulation with release characteristics suitable for the treatment of any disease or disorder requiring administration of the PRO polypeptide, microencapsulation of the PRO polypeptide is contemplated. Microencapsulation of recombinant proteins for sustained release has been successfully performed with human growth hormone (rhGH), interferon-(rhIFN-), interleukin-2, and MN rgp120. Johnson et al., Nat. Med., 2:795-799 (1996); Yasuda, Biomed. Ther., 27:1221-1223 (1993); Hora et al., Bio/Technology, 8:755-758 (1990); Cleland, "Design and Production of Single Immunization Vaccines Using Polylactide Polyglycolide Microsphere Systems," in Vaccine Design: The Subunit and Adjuvant Approach, Powell and Newman, eds, (Plenum Press: New York, 1995), pp. 439-462; WO 97/03692, WO 96/40072, WO 96/07399; and U.S. Pat. No. 5,654,010.

The sustained-release formulations of these proteins were developed using poly-lactic-coglycolic acid (PLGA) polymer due to its biocompatibility and wide range of biodegradable properties. The degradation products of PLGA, lactic and glycolic acids, can be cleared quickly within the human body. Moreover, the degradability of this polymer can be adjusted from months to years depending on its molecular weight and composition. Lewis, "Controlled release of bioactive agents from lactide/glycolide polymer," in: M. Chasin and R. Langer (Eds.), Biodegradable Polymers as Drug Delivery Systems (Marcel Dekker: New York, 1990),

pp. 1-41.

This invention encompasses methods of screening compounds to identify those that mimic the PRO polypeptide (agonists) or prevent the effect of the PRO polypeptide (antagonists). Screening assays for antagonist drug candidates are designed to identify compounds that bind or complex with the PRO polypeptides encoded by the genes identified herein, or otherwise interfere with the interaction of the encoded polypeptides with other cellular proteins. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates.

The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays, and cell-based assays, which are well characterized in the art.

All assays for antagonists are common in that they call for contacting the drug candidate with a PRO polypeptide encoded by a nucleic acid identified herein under conditions and for a time sufficient to allow these two components to interact.

In binding assays, the interaction is binding and the complex formed can be isolated or detected in the reaction mixture. In a particular embodiment, the PRO polypeptide encoded by the gene identified herein or the drug candidate is immobilized on a solid phase, e.g., on a microtiter plate, by covalent or non-covalent attachments. Non-covalent attachment generally is accomplished by coating the solid surface with a solution of the PRO polypeptide and drying. Alternatively, an immobilized antibody, e.g., a monoclonal antibody, specific for the PRO polypeptide to be immobilized can be used to anchor it to a solid surface. The assay is performed by adding the non-immobilized component, which may be labeled by a detectable label, to the immobilized component, e.g., the coated surface containing the anchored component. When the reaction is complete, the non-reacted components are removed, e.g., by washing, and complexes anchored on the solid surface are detected. When the originally non-immobilized component carries a detectable label, the detection of label immobilized on the surface indicates that complexing occurred. Where the originally non-immobilized component does not carry a label, complexing can be detected, for example, by using a labeled antibody specifically binding the immobilized complex.

If the candidate compound interacts with but does not bind to a particular PRO polypeptide encoded by a gene identified herein, its interaction with that polypeptide can be assayed by methods well known for detecting protein-protein interactions. Such assays include traditional approaches, such as, e.g., cross-linking, co-immunoprecipitation, and co-purification through gradients or chromatographic columns. In addition, protein-protein interactions can be monitored by using a yeast-based genetic system described by Fields and co-workers (Fields and Song, Nature (London), 340:245-246 (1989); Chien et al., Proc. Natl. Acad. Sci. USA, 88:9578-9582 (1991)) as disclosed by Chevray and Nathans, Proc. Natl. Acad. Sci. USA, 89: 5789-5793 (1991). Many transcriptional activators, such as yeast GAL4, consist of two physically discrete modular domains, one acting as the DNA-binding domain, the other one functioning as the transcription-activation domain. The yeast expression system described in the foregoing publications (generally referred to as the "two-hybrid system") takes advantage of this property, and employs two hybrid proteins, one in which the target protein is fused to the DNA-binding domain of GAL4, and another, in which candidate activating proteins are fused to the activation domain. The expression of a GAL1-*lac*Z reporter gene under control of a GAL4-activated promoter

depends on reconstitution of GAL4 activity via protein-protein interaction. Colonies containing interacting polypeptides are detected with a chromogenic substrate for β-galactosidase. A complete kit (MATCHMAKER™) for identifying protein-protein interactions between two specific proteins using the two-hybrid technique is commercially available from Clontech. This system can also be extended to map protein domains involved in specific protein interactions as well as to pinpoint amino acid residues that are crucial for these interactions.

Compounds that interfere with the interaction of a gene encoding a PRO polypeptide identified herein and other intra- or extracellular components can be tested as follows: usually a reaction mixture is prepared containing the product of the gene and the intra- or extracellular component under conditions and for a time allowing for the interaction and binding of the two products. To test the ability of a candidate compound to inhibit binding, the reaction is run in the absence and in the presence of the test compound. In addition, a placebo may be added to a third reaction mixture, to serve as positive control. The binding (complex formation) between the test compound and the intra- or extracellular component present in the mixture is monitored as described hereinabove. The formation of a complex in the control reaction(s) but not in the reaction mixture containing the test compound indicates that the test compound interferes with the interaction of the test compound and its reaction partner.

To assay for antagonists, the PRO polypeptide may be added to a cell along with the compound to be screened for a particular activity and the ability of the compound to inhibit the activity of interest in the presence of the PRO polypeptide indicates that the compound is an antagonist to the PRO polypeptide. Alternatively, antagonists may be detected by combining the PRO polypeptide and a potential antagonist with membrane-bound PRO polypeptide receptors or recombinant receptors under appropriate conditions for a competitive inhibition assay. The PRO polypeptide can be labeled, such as by radioactivity, such that the number of PRO polypeptide molecules bound to the receptor can be used to determine the effectiveness of the potential antagonist. The gene encoding the receptor can be identified by numerous methods known to those of skill in the art, for example, ligand panning and FACS sorting. Coligan et al., Current Protocols in Immun., 1(2): Chapter 5 (1991). Preferably, expression cloning is employed wherein polyadenylated RNA is prepared from a cell responsive to the PRO polypeptide and a cDNA library created from this RNA is divided into pools and used to transfect COS cells or other cells that are not responsive to the PRO polypeptide. Transfected cells that are grown on glass slides are exposed to labeled PRO polypeptide. The PRO polypeptide can be labeled by a variety of means including iodination or inclusion of a recognition site for a site-specific protein kinase. Following fixation and incubation, the slides are subjected to autoradiographic analysis. Positive pools are identified and sub-pools are prepared and re-transfected using an interactive sub-pooling and re-screening process, eventually yielding a single clone that encodes the putative receptor.

As an alternative approach for receptor identification, labeled PRO polypeptide can be photoaffinity-linked with cell membrane or extract preparations that express the receptor molecule. Cross-linked material is resolved by PAGE and exposed to X-ray film. The labeled complex containing the receptor can be excised, resolved into peptide fragments, and subjected to protein micro-sequencing. The amino acid sequence obtained from micro- sequencing would be used to design a set of degenerate oligonucleotide probes to screen a cDNA library to identify the gene encoding the putative receptor.

In another assay for antagonists, mammalian cells or a membrane preparation expressing the receptor would be incubated with labeled PRO polypeptide in the presence of the candidate compound. The ability of the compound to enhance or block this interaction could then be measured.

More specific examples of potential antagonists include an oligonucleotide that binds to the fusions of immunoglobulin with PRO polypeptide, and, in particular, antibodies including, without limitation, poly- and monoclonal antibodies and antibody fragments, single-chain antibodies, anti-idiotypic antibodies, and chimeric or humanized versions of such antibodies or fragments, as well as human antibodies and antibody fragments. Alternatively, a potential antagonist may be a closely related protein, for example, a mutated form of the PRO polypeptide that recognizes the receptor but imparts no effect, thereby competitively inhibiting the action of the PRO polypeptide.

Another potential PRO polypeptide antagonist is an antisense RNA or DNA construct prepared using antisense technology, where, e.g., an antisense RNA or DNA molecule acts to block directly the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, both of which methods are based on binding of a polynucleotide to DNA or RNA. For example, the 5' coding portion of the polynucleotide sequence, which encodes the mature PRO polypeptides herein, is used to design an antisense RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription (triple helix - see Lee et al., Nucl. Acids Res., 6:3073 (1979); Cooney et al., Science, 241: 456 (1988); Dervan et al., Science, 251:1360 (1991)), thereby preventing transcription and the production of the PRO polypeptide. The antisense RNA oligonucleotide hybridizes to the mRNA *in vivo* and blocks translation of the mRNA molecule into the PRO polypeptide (antisense - Okano, Neurochem., 56:560 (1991); Oligodeoxynucleotides as Antisense Inhibitors of Gene Expression (CRC Press: Boca Raton, FL, 1988). The oligonucleotides described above can also be delivered to cells such that the antisense RNA or DNA may be expressed *in vivo* to inhibit production of the PRO polypeptide. When antisense DNA is used, oligodeoxyribonucleotides derived from the translation-initiation site, e.g., between about -10 and +10 positions of the target gene nucleotide sequence, are preferred.

Potential antagonists include small molecules that bind to the active site, the receptor binding site, or growth factor or other relevant binding site of the PRO polypeptide, thereby blocking the normal biological activity of the PRO polypeptide. Examples of small molecules include, but are not limited to, small peptides or peptide-like molecules, preferably soluble peptides, and synthetic non-peptidyl organic or inorganic compounds.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozymes act by sequence-specific hybridization to the complementary target RNA, followed by endonucleolytic cleavage. Specific ribozyme cleavage sites within a potential RNA target can be identified by known techniques. For further details see, e.g., Rossi, Current Biology, 4:469-471 (1994), and PCT publication No. WO 97/33551 (published September 18, 1997).

Nucleic acid molecules in triple-helix formation used to inhibit transcription should be single-stranded and composed of deoxynucleotides. The base composition of these oligonucleotides is designed such that it promotes triple-helix formation via Hoogsteen base-pairing rules, which generally require sizeable stretches of

purines or pyrimidines on one strand of a duplex. For further details see, e.g., PCT publication No. WO 97/33551, *supra*.

These small molecules can be identified by any one or more of the screening assays discussed hereinabove and/or by any other screening techniques well known for those skilled in the art.

With regard to the PRO211 and PRO217 polypeptide, therapeutic indications include disorders associated with the preservation and maintenance of gastrointestinal mucosa and the repair of acute and chronic mucosal lesions (e.g., enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration and congenital microvillus atrophy), skin diseases associated with abnormal keratinocyte differentiation (e.g., psoriasis, epithelial cancers such as lung squamous cell carcinoma, epidermoid carcinoma of the vulva and gliomas.

Since the PRO232 polypeptide and nucleic acid encoding it possess sequence homology to a cell surface stem cell antigen and its encoding nucleic acid, probes based upon the PRO232 nucleotide sequence may be employed to identify other novel stem cell surface antigen proteins. Soluble forms of the PRO232 polypeptide may be employed as antagonists of membrane bound PRO232 activity both *in vitro* and *in vivo*. PRO232 polypeptides may be employed in screening assays designed to identify agonists or antagonists of the native PRO232 polypeptide, wherein such assays may take the form of any conventional cell-type or biochemical binding assay. Moreover, the PRO232 polypeptide may serve as a molecular marker for the tissues in which the polypeptide is specifically expressed.

With regard to the PRO187 polypeptides disclosed herein, FGF-8 has been implicated in cellular differentiation and embryogenesis, including the patterning which appears during limb formation. FGF-8 and the PRO187 molecules of the invention therefore are likely to have potent effects on cell growth and development. Diseases which relate to cellular growth and differentiation are therefore suitable targets for therapeutics based on functionality similar to FGF-8. For example, diseases related to growth or survival of nerve cells including Parkinson's disease, Alzheimer's disease, ALS, neuropathies. Additionally, disease related to uncontrolled cell growth, e.g., cancer, would also be expected therapeutic targets.

With regard to the PRO265 polypeptides disclosed herein, other methods for use with PRO265 are described in U.S. Patent 5,654,270 to Ruoslahti et al. In particular, PRO265 can be used in comparison with the fibromodulin disclosed therein to compare its effects on reducing dermal scarring and other properties of the fibromodulin described therein including where it is located and with what it binds and does not.

The PRO219 polypeptides of the present invention which play a regulatory role in the blood coagulation cascade may be employed *in vivo* for therapeutic purposes as well as for *in vitro* purposes. Those of ordinary skill in the art will well know how to employ PRO219 polypeptides for such uses.

The PRO246 polypeptides of the present invention which serve as cell surface receptors for one or more viruses will find other uses. For example, extracellular domains derived from these PRO246 polypeptides may be employed therapeutically *in vivo* for lessening the effects of viral infection. Those PRO246 polypeptides which serves as tumor specific antigens may be exploited as therapeutic targets for anti-tumor drugs, and the like. Those of ordinary skill in the art will well know how to employ PRO246 polypeptides for such uses.

Assays in which connective growth factor and other growth factors are usually used should be performed with PRO261. An assay to determine whether TGF beta induces PRO261, indicating a role in cancer is performed as known in the art. Wound repair and tissue growth assays are also performed with PRO261.

The results are applied accordingly.

PRO228 polypeptides should be used in assays in which EMR1, CD97 and latrophilin would be used in to determine their relative activities. The results can be applied accordingly. For example, a competitive binding assay with PRO228 and CD97 can be performed with the ligand for CD97, CD55.

Native PRO533 is a 216 amino acid polypeptide of which residues 1-22 are the signal sequence. Residues 3 to 216 have a Blast score of 509, corresponding to 53% homology to fibroblast growth factor. At the nucleotide level, DNA47412, the EST from which PCR oligos were generated to isolate the full length DNA49435-1219, has been observed to map to 11p15. Sequence homology to the 11p15 locus would indicate that PRO533 may have utility in the treatment of Usher Syndrome or Atrophia areata.

As mentioned previously, fibroblast growth factors can act upon cells in both a mitogenic and non-mitogenic manner. These factors are mitogenic for a wide variety of normal diploid mesoderm-derived and neural crest-derived cells, inducing granulosa cells, adrenal cortical cells, chrondrocytes, myoblasts, corneal and vascular endothelial cells (bovine or human), vascular smooth muscle cells, lens, retina and prostatic epithelial cells, oligodendrocytes, astrocytes, chrondocytes, myoblasts and osteoblasts.

Non-mitogenic actions of fibroblast growth factors include promotion of cell migration into a wound area (chemotaxis), initiation of new blood vessel formulation (angiogenesis), modulation of nerve regeneration and survival (neurotrophism), modulation of endocrine functions, and stimulation or suppression of specific cellular protein expression, extracellular matrix production and cell survival. Baird, A. & Bohlen, P., *Handbook of Exp. Phrmacol.* 95(1): 369-418 (1990). These properties provide a basis for using fibroblast growth factors in therapeutic approaches to accelerate wound healing, nerve repair, collateral blood vessel formation, and the like. For example, fibroblast growth factors, have been suggested to minimize myocardium damage in heart disease and surgery (U.S.P. 4,378,437).

Since the PRO245 polypeptide and nucleic acid encoding it possess sequence homology to a transmembrane protein tyrosine kinase protein and its encoding nucleic acid, probes based upon the PRO245 nucleotide sequence may be employed to identify other novel transmembrane tyrosine kinase proteins. Soluble forms of the PRO245 polypeptide may be employed as antagonists of membrane bound PRO245 activity both *in vitro* and *in vivo*. PRO245 polypeptides may be employed in screening assays designed to identify agonists or antagonists of the native PRO245 polypeptide, wherein such assays may take the form of any conventional cell-type or biochemical binding assay. Moreover, the PRO245 polypeptide may serve as a molecular marker for the tissues in which the polypeptide is specifically expressed.

PRO220, PRO221 and PRO227 all have leucine rich repeats. Additionally, PRO220 and PRO221 have homology to SLIT and leucine rich repeat protein. Therefore, these proteins are useful in assays described in the literature, supra, wherein the SLIT and leucine rich repeat protein are used. Regarding the SLIT protein, PRO227 can be used in an assay to determine the affect of PRO227 on neurodegenerative disease. Additionally, PRO227 has homology to human glycoprotein V. In the case of PRO227, this polypeptide is used in an assay to determine its affect on bleeding, clotting, tissue repair and scarring.

The PRO266 polypeptide can be used in assays to determine if it has a role in neurodegenerative diseases or their reversal.

PRO269 polypeptides and portions thereof which effect the activity of thrombin may also be useful for

in vivo therapeutic purposes, as well as for various *in vitro* applications. In addition, PRO269 polypeptides and portions thereof may have therapeutic use as an antithrombotic agent with reduced risk for hemorrhage as compared with heparin. Peptides having homology to thrombomodulin are particularly desirable.

PRO287 polypeptides and portions thereof which effect the activity of bone morphogenic protein "BMP1"/procollagen C-proteinase (PCP) may also be useful for *in vivo* therapeutic purposes, as well as for various *in vitro* applications. In addition, PRO287 polypeptides and portions thereof may have therapeutic applications in wound healing and tissue repair. Peptides having homology to procollagen C-proteinase enhancer protein and its precursor may also be used to induce bone and/or cartilage formation and are therefore of particular interest to the scientific and medical communities.

Therapeutic indications for PRO214 polypeptides include disorders associated with the preservation and maintenance of gastrointestinal mucosa and the repair of acute and chronic mucosal lesions (e.g., enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration and congenital microvillus atrophy), skin diseases associated with abnormal keratinocyte differentiation (e.g., psoriasis, epithelial cancers such as lung squamous cell carcinoma, epidermoid carcinoma of the vulva and gliomas.

Studies on the generation and analysis of mice deficient in members of the TGF- superfamily are reported in Matzuk, Trends in Endocrinol. and Metabol., 6: 120-127 (1995).

The PRO317 polypeptide, as well as PRO317-specific antibodies, inhibitors, agonists, receptors, or their analogs, herein are useful in treating PRO317-associated disorders. Hence, for example, they may be employed in modulating endometrial bleeding angiogenesis, and may also have an effect on kidney tissue. Endometrial bleeding can occur in gynecological diseases such as endometrial cancer as abnormal bleeding. Thus, the compositions herein may find use in diagnosing and treating abnormal bleeding conditions in the endometrium, as by reducing or eliminating the need for a hysterectomy. The molecules herein may also find use in angiogenesis applications such as anti-tumor indications for which the antibody against vascular endothelial growth factor is used, or, conversely, ischemic indications for which vascular endothelial growth factor is employed.

Bioactive compositions comprising PRO317 or agonists or antagonists thereof may be administered in a suitable therapeutic dose determined by any of several methodologies including clinical studies on mammalian species to determine maximal tolerable dose and on normal human subjects to determine safe dose. Additionally, the bioactive agent may be complexed with a variety of well established compounds or compositions which enhance stability or pharmacological properties such as half-life. It is contemplated that the therapeutic, bioactive composition may be delivered by intravenous infusion into the bloodstream or any other effective means which could be used for treating problems of the kidney, uterus, endometrium, blood vessels, or related tissue, *e.g.*, in the heart or genital tract.

Dosages and administration of PRO317, PRO317 agonist, or PRO317 antagonist in a pharmaceutical composition may be determined by one of ordinary skill in the art of clinical pharmacology or pharmacokinetics. See, for example, Mordenti and Rescigno, Pharmaceutical Research, 9:17-25 (1992); Morenti *et al.*, Pharmaceutical Research, 8:1351-1359 (1991); and Mordenti and Chappell, "The use of interspecies scaling in toxicokinetics" in Toxicokinetics and New Drug Development, Yacobi *et al.* (eds) (Pergamon Press: NY, 1989), pp. 42-96. An effective amount of PRO317, PRO317 agonist, or PRO317 antagonist to be employed

therapeutically will depend, for example, upon the therapeutic objectives, the route of administration, and the condition of the mammal. Accordingly, it will be necessary for the therapist to titer the dosage and modify the route of administration as required to obtain the optimal therapeutic effect. A typical daily dosage might range from about 10 ng/kg to up to 100 mg/kg of the mammal's body weight or more per day, preferably about 1 μg/kg/day to 10 mg/kg/day. Typically, the clinician will administer PRO317, PRO317 agonist, or PRO317 antagonist, until a dosage is reached that achieves the desired effect for treatment of the above mentioned disorders.

PRO317 or an PRO317 agonist or PRO317 antagonist may be administered alone or in combination with another to achieve the desired pharmacological effect. PRO317 itself, or agonists or antagonists of PRO317 can provide different effects when administered therapeutically. Such compounds for treatment will be formulated in a nontoxic, inert, pharmaceutically acceptable aqueous carrier medium preferably at a pH of about 5 to 8, more preferably 6 to 8, although the pH may vary according to the characteristics of the PRO317, agonist, or antagonist being formulated and the condition to be treated. Characteristics of the treatment compounds include solubility of the molecule, half-life, and antigenicity/immunogenicity; these and other characteristics may aid in defining an effective carrier.

PRO317 or PRO317 agonists or PRO317 antagonists may be delivered by known routes of administration including but not limited to topical creams and gels; transmucosal spray and aerosol, transdermal patch and bandage; injectable, intravenous, and lavage formulations; and orally administered liquids and pills, particularly formulated to resist stomach acid and enzymes. The particular formulation, exact dosage, and route of administration will be determined by the attending physician and will vary according to each specific situation.

Such determinations of administration are made by considering multiple variables such as the condition to be treated, the type of mammal to be treated, the compound to be administered, and the pharmacokinetic profile of the particular treatment compound. Additional factors which may be taken into account include disease state (*e.g.* severity) of the patient, age, weight, gender, diet, time of administration, drug combination, reaction sensitivities, and tolerance/response to therapy. Long-acting treatment compound formulations (such as liposomally encapsulated PRO317 or PEGylated PRO317 or PRO317 polymeric microspheres, such as polylactic acid-based microspheres) might be administered every 3 to 4 days, every week, or once every two weeks depending on half-life and clearance rate of the particular treatment compound.

Normal dosage amounts may vary from about 10 ng/kg to up to 100 mg/kg of mammal body weight or more per day, preferably about 1 μg/kg/day to 10 mg/kg/day, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature; see, for example, U.S. Pat. Nos. 4,657,760; 5,206,344; or 5,225,212. It is anticipated that different formulations will be effective for different treatment compounds and different disorders, that administration targeting the uterus, for example, may necessitate delivery in a manner different from that to another organ or tissue, such as cardiac tissue.

Where sustained-release administration of PRO317 is desired in a formulation with release characteristics suitable for the treatment of any disease or disorder requiring administration of PRO317, microencapsulation of PRO317 is contemplated. Microencapsulation of recombinant proteins for sustained release has been successfully performed with human growth hormone (rhGH), interferon- (rhIFN-), interleukin-2, and MN rgp120. Johnson *et al.*, Nat. Med., 2: 795-799 (1996); Yasuda, Biomed. Ther., 27:

1221-1223 (1993); Hora *et al.*, Bio/Technology, 8: 755-758 (1990); Cleland, "Design and Production of Single Immunization Vaccines Using Polylactide Polyglycolide Microsphere Systems," in Vaccine Design: The Subunit and Adjuvant Approach , Powell and Newman, eds, (Plenum Press: New York, 1995), pp. 439-462; WO 97/03692, WO 96/40072, WO 96/07399; and U.S Pat. No. 5,654,010.

It is contemplated that conditions or diseases of the uterus, endometrial tissue, or other genital tissues or cardiac tissues may precipitate damage that is treatable with PRO317 or PRO317 agonist where PRO317 expression is reduced in the diseased state; or with antibodies to PRO317 or other PRO317 antagonists where the expression of PRO317 is increased in the diseased state. These conditions or diseases may be specifically diagnosed by the probing tests discussed above for physiologic and pathologic problems which affect the function of the organ.

The PRO317, PRO317 agonist, or PRO317 antagonist may be administered to a mammal with another biologically active agent, either separately or in the same formulation to treat a common indication for which they are appropriate. For example, it is contemplated that PRO317 can be administered together with EBAF-1 for those indications on which they demonstrate the same qualitative biological effects. Alternatively, where they have opposite effects, EBAF-1 may be administered together with an antagonist to PRO317, such as an anti-PRO317 antibody. Further, PRO317 may be administered together with VEGF for coronary ischemia where such indication is warranted, or with an anti-VEGF for cancer as warranted, or, conversely, an antagonist to PRO317 may be administered with VEGF for coronary ischemia or with anti-VEGF to treat cancer as warranted. These administrations would be in effective amounts for treating such disorders.

Native PRO301 (SEQ ID NO:119) has a Blast score of 246 and 30% homology at residues 24 to 282 of Figure 44 with A33_HUMAN, an A33 antigen precursor. A33 antigen precursor, as explained in the Background is a tumor-specific antigen, and as such, is a recognized marker and therapeutic target for the diagnosis and treatment of colon cancer. The expression of tumor-specific antigens is often associated with the progression of neoplastic tissue disorders. Native PRO301 (SEQ ID NO:119) and A33_HUMAN also show a Blast score of 245 and 30% homology at residues 21 to 282 of Fig. 44 with A33_HUMAN, the variation dependent upon how spaces are inserted into the compared sequences. Native PRO301 (SEQ ID NO:119) also has a Blast score of 165 and 29% homology at residues 60 to 255 of Fig. 44 with HS46KDA_1, a human coxsackie and adenovirus receptor protein, also known as cell surface protein HCAR. This region of PRO301 also shows a similar Blast score and homology with HSU90716_1. Expression of such proteins is usually associated with viral infection and therapeutics for the prevention of such infection may be accordingly conceived. As mentioned in the Background, the expression of viral receptors is often associated with neoplastic tumors.

Therapeutic uses for the PRO234 polypeptides of the invention includes treatments associated with leukocyte homing or the interaction between leukocytes and the endothelium during an inflammatory response. Examples include asthma, rheumatoid arthritis, psoriasis and multiple sclerosis.

Since the PRO231 polypeptide and nucleic acid encoding it possess sequence homology to a putative acid phosphatase and its encoding nucleic acid, probes based upon the PRO231 nucleotide sequence may be employed to identify other novel phosphatase proteins. Soluble forms of the PRO231 polypeptide may be employed as antagonists of membrane bound PRO231 activity both *in vitro* and *in vivo*. PRO231 polypeptides

may be employed in screening assays designed to identify agonists or antagonists of the native PRO231 polypeptide, wherein such assays may take the form of any conventional cell-type or biochemical binding assay. Moreover, the PRO231 polypeptide may serve as a molecular marker for the tissues in which the polypeptide is specifically expressed.

PRO229 polypeptides can be fused with peptides of interest to determine whether the fusion peptide has an increased half-life over the peptide of interest. The PRO229 polypeptides can be used accordingly to increase the half-life of polypeptides of interest. Portions of PRO229 which cause the increase in half-life are an embodiment of the invention herein.

PRO238 can be used in assays which measure its ability to reduce substrates, including oxygen and Aceyl-CoA, and particularly, measure PRO238's ability to produce oxygen free radicals. This is done by using assays which have been previously described. PRO238 can further be used to assay for candidates which block, reduce or reverse its reducing abilities. This is done by performing side by side assays where candidates are added in one assay having PRO238 and a substrate to reduce, and not added in another assay, being the same but for the lack of the presence of the candidate.

PRO233 polypeptides and portions thereof which have homology to reductase may also be useful for *in vivo* therapeutic purposes, as well as for various other applications. The identification of novel reductase proteins and related molecules may be relevant to a number of human disorders such as inflammatory disease, organ failure, atherosclerosis, cardiac injury, infertility, birth defects, premature aging, AIDS, cancer, diabetic complications and mutations in general. Given that oxygen free radicals and antioxidants appear to play important roles in a number of disease processes, the identification of new reductase proteins and reductase-like molecules is of special importance in that such proteins may serve as potential therapeutics for a variety of different human disorders. Such polypeptides may also play important roles in biotechnological and medical research, as well as various industrial applications. As a result, there is particular scientific and medical interest in new molecules, such as PRO233.

The PRO223 polypeptides of the present invention which exhibit serine carboxypeptidease activity may be employed *in vivo* for therapeutic purposes as well as for *in vitro* purposes. Those of ordinary skill in the art will well know how to employ PRO223 polypeptides for such uses.

PRO235 polypeptides and portions thereof which may be involved in cell adhesion are also useful for *in vivo* therapeutic purposes, as well as for various *in vitro* applications. In addition, PRO235 polypeptides and portions thereof may have therapeutic applications in disease states which involve cell adhesion. Given the physiological importance of cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in cell adhesion. Therefore, peptides having homology to plexin are of particular interest to the scientific and medical communities.

Because the PRO236 and PRO262 polypeptides disclosed herein are homologous to various known β-galactosidase proteins, the PRO236 and PRO262 polypeptides disclosed herein will find use in conjugates of monoclonal antibodies and the polypeptide for specific killing of tumor cells by generation of active drug from a galactosylated prodrug (e.g., the generation of 5-fluorouridine from the prodrug β-D-galactosyl-5-fluorouridine). The PRO236 and PRO262 polypeptides disclosed herein may also find various uses both *in vivo* and *in vitro*, wherein those uses will be similar or identical to uses for which β-galactosidase proteins are now

employed. Those of ordinary skill in the art will well know how to employ PRO236 and PRO262 polypeptides for such uses.

PRO239 polypeptides and portions thereof which have homology to densin may also be useful for *in vivo* therapeutic purposes, as well as for various *in vitro* applications. In addition, PRO239 polypeptides and portions thereof may have therapeutic applications in disease states which involve synaptic mechanisms, regeneration or cell adhesion. Given the physiological importance of synaptic processes, regeneration and cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in synaptic machinery and cell adhesion. Therefore, peptides having homology to densin are of particular interest to the scientific and medical communities.

The PRO260 polypeptides described herein can be used in assays to determine their relation to fucosidase. In particular, the PRO260 polypeptides can be used in assays in determining their ability to remove fucose or other sugar residues from proteoglycans. The PRO260 polypeptides can be assayed to determine if they have any functional or locational similarities as fucosidase. The PRO260 polypeptides can then be used to regulate the systems in which they are integral.

PRO263 can be used in assays wherein CD44 antigen is generally used to determine PRO263 activity relative to that of CD44. The results can be used accordingly.

PRO270 polypeptides and portions thereof which effect reduction-oxidation (redox) state may also be useful for *in vivo* therapeutic purposes, as well as for various *in vitro* applications. More specifically, PRO270 polypeptides may affect the expression of a large variety of genes thought to be involved in the pathogenesis of AIDS, cancer, atherosclerosis, diabetic complications and in pathological conditions involving oxidative stress such as stroke and inflammation. In addition, PRO270 polypeptides and portions thereof may affect the expression of a genes which have a role in apoptosis. Therefore, peptides having homology to thioredoxin are particularly desirable to the scientific and medical communities.

PRO272 polypeptides and portions thereof which possess the ability to bind calcium may also have numerous *in vivo* therapeutic uses, as well as various *in vitro* applications. Therefore, peptides having homology to reticulocalbin are particularly desirable. Those with ordinary skill in the art will know how to employ PRO272 polypeptides and portions thereof for such purposes.

PRO294 polypeptides and portions thereof which have homology to collagen may also be useful for *in vivo* therapeutic purposes, as well as for various other applications. The identification of novel collagens and collage-like molecules may have relevance to a number of human disorders. Thus, the identification of new collagens and collage-like molecules is of special importance in that such proteins may serve as potential therapeutics for a variety of different human disorders. Such polypeptides may also play important roles in biotechnological and medical research as well as various industrial applications. Given the large number of uses for collagen, there is substantial interest in polypeptides with homology to the collagen molecule.

PRO295 polypeptides and portions thereof which have homology to integrin may also be useful for *in vivo* therapeutic purposes, as well as for various other applications. The identification of novel integrins and integrin-like molecules may have relevance to a number of human disorders such as modulating the binding or activity of cells of the immune system. Thus, the identification of new integrins and integrin-like molecules is

of special importance in that such proteins may serve as potential therapeutics for a variety of different human disorders. Such polypeptides may also play important roles in biotechnological and medical research as well as various industrial applications. As a result, there is particular scientific and medical interest in new molecules, such as PRO295.

As the PRO293 polypeptide is clearly a leucine rich repeat polypeptide homologue, the peptide can be used in all applications that the known NLRR-1 and NLRR-2 polypeptides are used. The activity can be compared between these peptides and thus applied accordingly.

The PRO247 polypeptides described herein can be used in assays in which densin is used to determine the activity of PRO247 relative to densin or these other proteins. The results can be used accordingly in diagnostics and/or therapeutic applications with PRO247.

PRO302, PRO303, PRO304, PRO307 and PRO343 polypeptides of the present invention which possess protease activity may be employed both *in vivo* for therapeutic purposes and *in vitro*. Those of ordinary skill in the art will well know how to employ the PRO302, PRO303, PRO304, PRO307 and PRO343 polypeptides of the present invention for such purposes.

PRO328 polypeptides and portions thereof which have homology to GLIP and CRISP may also be useful for *in vivo* therapeutic purposes, as well as for various other applications. The identification of novel GLIP and CRISP-like molecules may have relevance to a number of human disorders which involve transcriptional regulation or are over expressed in human tumors. Thus, the identification of new GLIP and CRISP-like molecules is of special importance in that such proteins may serve as potential therapeutics for a variety of different human disorders. Such polypeptides may also play important roles in biotechnological and medical research as well as in various industrial applications. As a result, there is particular scientific and medical interest in new molecules, such as PRO328.

Uses for PRO335, PRO331 or PRO326 including uses in competitive assays with LIG-1, ALS and decorin to determine their relative activities. The results can be used accordingly. PRO335, PRO331 or PRO326 can also be used in assays where LIG-1 would be used to determine if the same effects are incurred.

PRO332 contains GAG repeat (GKEK) at amino acid positions 625-628 in Fig. 108 (SEQ ID NO:310). Slippage in such repeats can be associated with human disease. Accordingly, PRO332 can use useful for the treatment of such disease conditions by gene therapy, i.e. by introduction of a gene containing the correct GKEK sequence motif.

Other uses of PRO334 include use in assays in which fibrillin or fibulin would be used to determine the relative activity of PRO334 to fibrillin or fibulin. In particular, PRO334 can be used in assays which require the mechanisms imparted by epidermal growth factor repeats.

Native PRO346 (SEQ ID NO:320) has a Blast score of 230, corresponding to 27% homology between amino acid residues 21 to 343 with residues 35 to 1040 CGM6_HUMAN, a carcinoembryonic antigen cgm6 precursor. This homology region includes nearly all but 2 N-terminal extracellular domain residues, including an immunoglobulin superfamily homology at residues 148 to 339 of PRO346 in addition to several transmembrane residues (340-343). Carcinoembryonic antigen precursor, as explained in the Background is a tumor-specific antigen, and as such, is a recognized marker and therapeutic target for the diagnosis and treatment of colon cancer. The expression of tumor-specific antigens is often associated with the progression of neoplastic

tissue disorders. Native PRO346 (SEQ ID NO:320) and P_W06874, a human carcinoembryonic antigen CEA-d have a Blast score of 224 and homology of 28% between residues 2 to 343 and 67 to 342, respectively. This homology includes the entire extracellular domain residues of native PRO346, minus the initiator methionine (residues 2 to 18) as well as several transmembrane residues (340-343).

PRO268 polypeptides which have protein disulfide isomerase activity will be useful for many applications where protein disulfide isomerase activity is desirable including, for example, for use in promoting proper disulfide bond formation in recombinantly produced proteins so as to increase the yield of correctly folded protein. Those of ordinary skill in the art will readily know how to employ such PRO268 polypeptides for such purposes.

PRO330 polypeptides of the present invention which possess biological activity related to that of the prolyl 4-hydroxylase alpha subunit protein may be employed both *in vivo* for therapeutic purposes and *in vitro*. Those of ordinary skill in the art will well know how to employ the PRO330 polypeptides of the present invention for such purposes.

Uses of the herein disclosed molecules may also be based upon the positive functional assay hits disclosed and described below.

F. Anti-PRO Antibodies

The present invention further provides anti-PRO antibodies. Exemplary antibodies include polyclonal, monoclonal, humanized, bispecific, and heteroconjugate antibodies.

1. Polyclonal Antibodies

The anti-PRO antibodies may comprise polyclonal antibodies. Methods of preparing polyclonal antibodies are known to the skilled artisan. Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. The immunizing agent may include the PRO polypeptide or a fusion protein thereof. It may be useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include but are not limited to keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. Examples of adjuvants which may be employed include Freund's complete adjuvant and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate). The immunization protocol may be selected by one skilled in the art without undue experimentation.

2. Monoclonal Antibodies

The anti-PRO antibodies may, alternatively, be monoclonal antibodies. Monoclonal antibodies may be prepared using hybridoma methods, such as those described by Kohler and Milstein, Nature, 256:495 (1975). In a hybridoma method, a mouse, hamster, or other appropriate host animal, is typically immunized with an immunizing agent to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized *in vitro*.

The immunizing agent will typically include the PRO polypeptide or a fusion protein thereof.

Generally, either peripheral blood lymphocytes ("PBLs") are used if cells of human origin are desired, or spleen cells or lymph node cells are used if non-human mammalian sources are desired. The lymphocytes are then fused with an immortalized cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell [Goding, Monoclonal Antibodies: Principles and Practice, Academic Press, (1986) pp. 59-103]. Immortalized cell lines are usually transformed mammalian cells, particularly myeloma cells of rodent, bovine and human origin. Usually, rat or mouse myeloma cell lines are employed. The hybridoma cells may be cultured in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, immortalized cells. For example, if the parental cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine ("HAT medium"), which substances prevent the growth of HGPRT-deficient cells.

Preferred immortalized cell lines are those that fuse efficiently, support stable high level expression of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. More preferred immortalized cell lines are murine myeloma lines, which can be obtained, for instance, from the Salk Institute Cell Distribution Center, San Diego, California and the American Type Culture Collection, Manassas, Virginia. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies [Kozbor, J. Immunol., 133:3001 (1984); Brodeur et al., Monoclonal Antibody Production Techniques and Applications, Marcel Dekker, Inc., New York, (1987) pp. 51-63].

The culture medium in which the hybridoma cells are cultured can then be assayed for the presence of monoclonal antibodies directed against PRO. Preferably, the binding specificity of monoclonal antibodies produced by the hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA). Such techniques and assays are known in the art. The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson and Pollard, Anal. Biochem., 107:220 (1980).

After the desired hybridoma cells are identified, the clones may be subcloned by limiting dilution procedures and grown by standard methods [Goding, supra]. Suitable culture media for this purpose include, for example, Dulbecco's Modified Eagle's Medium and RPMI-1640 medium. Alternatively, the hybridoma cells may be grown *in vivo* as ascites in a mammal.

The monoclonal antibodies secreted by the subclones may be isolated or purified from the culture medium or ascites fluid by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

The monoclonal antibodies may also be made by recombinant DNA methods, such as those described in U.S. Patent No. 4,816,567. DNA encoding the monoclonal antibodies of the invention can be readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells of the invention serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. The DNA also may be modified, for example, by substituting the

coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences [U.S. Patent No. 4,816,567; Morrison et al., supra] or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. Such a non-immunoglobulin polypeptide can be substituted for the constant domains of an antibody of the invention, or can be substituted for the variable domains of one antigen-combining site of an antibody of the invention to create a chimeric bivalent antibody.

The antibodies may be monovalent antibodies. Methods for preparing monovalent antibodies are well known in the art. For example, one method involves recombinant expression of immunoglobulin light chain and modified heavy chain. The heavy chain is truncated generally at any point in the Fc region so as to prevent heavy chain crosslinking. Alternatively, the relevant cysteine residues are substituted with another amino acid residue or are deleted so as to prevent crosslinking.

In vitro methods are also suitable for preparing monovalent antibodies. Digestion of antibodies to produce fragments thereof, particularly, Fab fragments, can be accomplished using routine techniques known in the art.

3. Human and Humanized Antibodies

The anti-PRO antibodies of the invention may further comprise humanized antibodies or human antibodies. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', $F(ab')_2$ or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies may also comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin [Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-329 (1988); and Presta, Curr. Op. Struct. Biol., 2:593-596 (1992)].

Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers [Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-327 (1988); Verhoeyen et al., Science, 239:1534-1536 (1988)], by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Patent

No. 4,816,567), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

Human antibodies can also be produced using various techniques known in the art, including phage display libraries [Hoogenboom and Winter, J. Mol. Biol., 227:381 (1991); Marks et al., J. Mol. Biol., 222:581 (1991)]. The techniques of Cole et al. and Boerner et al. are also available for the preparation of human monoclonal antibodies (Cole et al., Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985) and Boerner et al., J. Immunol., 147(1):86-95 (1991)]. Similarly, human antibodies can be made by introducing of human immunoglobulin loci into transgenic animals, e.g., mice in which the endogenous immunoglobulin genes have been partially or completely inactivated. Upon challenge, human antibody production is observed, which closely resembles that seen in humans in all respects, including gene rearrangement, assembly, and antibody repertoire. This approach is described, for example, in U.S. Patent Nos. 5,545,807; 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,661,016, and in the following scientific publications: Marks *et al.*, Bio/Technology 10, 779-783 (1992); Lonberg *et al.*, Nature 368 856-859 (1994); Morrison, Nature 368, 812-13 (1994); Fishwild *et al.*, Nature Biotechnology 14, 845-51 (1996); Neuberger, Nature Biotechnology 14, 826 (1996); Lonberg and Huszar, Intern. Rev. Immunol. 13 65-93 (1995).

4. Bispecific Antibodies

Bispecific antibodies are monoclonal, preferably human or humanized, antibodies that have binding specificities for at least two different antigens. In the present case, one of the binding specificities is for the PRO, the other one is for any other antigen, and preferably for a cell-surface protein or receptor or receptor subunit.

Methods for making bispecific antibodies are known in the art. Traditionally, the recombinant production of bispecific antibodies is based on the co-expression of two immunoglobulin heavy-chain/light-chain pairs, where the two heavy chains have different specificities [Milstein and Cuello, Nature, 305:537-539 (1983)]. Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of ten different antibody molecules, of which only one has the correct bispecific structure. The purification of the correct molecule is usually accomplished by affinity chromatography steps. Similar procedures are disclosed in WO 93/08829, published 13 May 1993, and in Traunecker et al., EMBO J., 10:3655-3659 (1991).

Antibody variable domains with the desired binding specificities (antibody-antigen combining sites) can be fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy-chain constant domain, comprising at least part of the hinge, CH2, and CH3 regions. It is preferred to have the first heavy-chain constant region (CH1) containing the site necessary for light-chain binding present in at least one of the fusions. DNAs encoding the immunoglobulin heavy-chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host organism. For further details of generating bispecific antibodies see, for example, Suresh et al., Methods in Enzymology, 121:210 (1986).

According to another approach described in WO 96/27011, the interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers which are recovered from recombinant cell culture. The preferred interface comprises at least a part of the CH3 region of an antibody constant domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (e.g. tyrosine or tryptophan). Compensatory "cavities" of identical or similar size to the large side chain(s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (e.g. alanine or threonine). This provides a mechanism for increasing the yield of the heterodimer over other unwanted end-products such as homodimers.

Bispecific antibodies can be prepared as full length antibodies or antibody fragments (e.g. $F(ab')_2$ bispecific antibodies). Techniques for generating bispecific antibodies from antibody fragments have been described in the literature. For example, bispecific antibodies can be prepared can be prepared using chemical linkage. Brennan *et al.*, Science 229:81 (1985) describe a procedure wherein intact antibodies are proteolytically cleaved to generate $F(ab')_2$ fragments. These fragments are reduced in the presence of the dithiol complexing agent sodium arsenite to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated are then converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives is then reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes.

Fab' fragments may be directly recovered from *E. coli* and chemically coupled to form bispecific antibodies. Shalaby *et al.*, J. Exp. Med. 175:217-225 (1992) describe the production of a fully humanized bispecific antibody $F(ab')_2$ molecule. Each Fab' fragment was separately secreted from *E. coli* and subjected to directed chemical coupling *in vitro* to form the bispecific antibody. The bispecific antibody thus formed was able to bind to cells overexpressing the ErbB2 receptor and normal human T cells, as well as trigger the lytic activity of human cytotoxic lymphocytes against human breast tumor targets.

Various technique for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. Kostelny *et al.*, J. Immunol. 148(5):1547-1553 (1992). The leucine zipper peptides from the Fos and Jun proteins were linked to the Fab' portions of two different antibodies by gene fusion. The antibody homodimers were reduced at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers. The "diabody" technology described by Hollinger *et al.*, Proc. Natl. Acad. Sci. USA 90:6444-6448 (1993) has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a heavy-chain variable domain (V_H) connected to a light-chain variable domain (V_L) by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the V_H and V_L domains of one fragment are forced to pair with the complementary V_L and V_H domains of another fragment, thereby forming two antigen-binding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (sFv) dimers has also been reported. See, Gruber *et al.*, J. Immunol. 152:5368 (1994).

Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be prepared. Tutt *et al.*, J. Immunol. 147:60 (1991).

Exemplary bispecific antibodies may bind to two different epitopes on a given PRO polypeptide herein. Alternatively, an anti-PRO polypeptide arm may be combined with an arm which binds to a triggering molecule on a leukocyte such as a T-cell receptor molecule (e.g. CD2, CD3, CD28, or B7), or Fc receptors for IgG (FcγR), such as FcγRI (CD64), FcγRII (CD32) and FcγRIII (CD16) so as to focus cellular defense mechanisms to the cell expressing the particular PRO polypeptide. Bispecific antibodies may also be used to localize cytotoxic agents to cells which express a particular PRO polypeptide. These antibodies possess a PRO-binding arm and an arm which binds a cytotoxic agent or a radionuclide chelator, such as EOTUBE, DPTA, DOTA, or TETA. Another bispecific antibody of interest binds the PRO polypeptide and further binds tissue factor (TF).

5. Heteroconjugate Antibodies

Heteroconjugate antibodies are also within the scope of the present invention. Heteroconjugate antibodies are composed of two covalently joined antibodies. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells [U.S. Patent No. 4,676,980], and for treatment of HIV infection [WO 91/00360; WO 92/200373; EP 03089]. It is contemplated that the antibodies may be prepared *in vitro* using known methods in synthetic protein chemistry, including those involving crosslinking agents. For example, immunotoxins may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents for this purpose include iminothiolate and methyl-4-mercaptobutyrimidate and those disclosed, for example, in U.S. Patent No. 4,676,980.

6. Effector Function Engineering

It may be desirable to modify the antibody of the invention with respect to effector function, so as to enhance, *e.g.*, the effectiveness of the antibody in treating cancer. For example, cysteine residue(s) may be introduced into the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron *et al.*, J. Exp Med., 176: 1191-1195 (1992) and Shopes, J. Immunol., 148: 2918-2922 (1992). Homodimeric antibodies with enhanced anti-tumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff *et al.* Cancer Research, 53: 2560-2565 (1993). Alternatively, an antibody can be engineered that has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson *et al.*, Anti-Cancer Drug Design, 3: 219-230 (1989).

7. Immunoconjugates

The invention also pertains to immunoconjugates comprising an antibody conjugated to a cytotoxic agent such as a chemotherapeutic agent, toxin (*e.g.*, an enzymatically active toxin of bacterial, fungal, plant, or animal origin, or fragments thereof), or a radioactive isotope (*i.e.*, a radioconjugate).

Chemotherapeutic agents useful in the generation of such immunoconjugates have been described above. Enzymatically active toxins and fragments thereof that can be used include diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain,

modeccin A chain, alpha-sarcin, *Aleurites fordii* proteins, dianthin proteins, *Phytolaca americana* proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, crotin, sapaonaria officinalis inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin, and the tricothecenes. A variety of radionuclides are available for the production of radioconjugated antibodies. Examples include ^{212}Bi, ^{131}I, ^{131}In, ^{90}Y, and ^{186}Re.

Conjugates of the antibody and cytotoxic agent are made using a variety of bifunctional protein-coupling agents such as N-succinimidyl-3-(2-pyridyldithiol) propionate (SPDP), iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutareldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared as described in Vitetta *et al.*, Science, 238: 1098 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody. See WO94/11026.

In another embodiment, the antibody may be conjugated to a "receptor" (such streptavidin) for utilization in tumor pretargeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of unbound conjugate from the circulation using a clearing agent and then administration of a "ligand" (*e.g.*, avidin) that is conjugated to a cytotoxic agent (*e.g.*, a radionucleotide).

8. Immunoliposomes

The antibodies disclosed herein may also be formulated as immunoliposomes. Liposomes containing the antibody are prepared by methods known in the art, such as described in Epstein *et al.*, Proc. Natl. Acad. Sci. USA, 82: 3688 (1985); Hwang *et al.*, Proc. Natl Acad. Sci. USA, 77: 4030 (1980); and U.S. Pat. Nos. 4,485,045 and 4,544,545. Liposomes with enhanced circulation time are disclosed in U.S. Patent No. 5,013,556.

Particularly useful liposomes can be generated by the reverse-phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol, and PEG-derivatized phosphatidylethanolamine (PEG-PE). Liposomes are extruded through filters of defined pore size to yield liposomes with the desired diameter. Fab' fragments of the antibody of the present invention can be conjugated to the liposomes as described in Martin *et al* ., J. Biol. Chem., 257: 286-288 (1982) via a disulfide-interchange reaction. A chemotherapeutic agent (such as Doxorubicin) is optionally contained within the liposome. See Gabizon *et al.*, J. National Cancer Inst., 81(19): 1484 (1989).

9. Pharmaceutical Compositions of Antibodies

Antibodies specifically binding a PRO polypeptide identified herein, as well as other molecules identified by the screening assays disclosed hereinbefore, can be administered for the treatment of various disorders in the form of pharmaceutical compositions.

If the PRO polypeptide is intracellular and whole antibodies are used as inhibitors, internalizing antibodies are preferred. However, lipofections or liposomes can also be used to deliver the antibody, or an antibody fragment, into cells. Where antibody fragments are used, the smallest inhibitory fragment that

specifically binds to the binding domain of the target protein is preferred. For example, based upon the variable-region sequences of an antibody, peptide molecules can be designed that retain the ability to bind the target protein sequence. Such peptides can be synthesized chemically and/or produced by recombinant DNA technology. See, *e.g.*, Marasco *et al.*, Proc. Natl. Acad. Sci. USA, 90: 7889-7893 (1993). The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. Alternatively, or in addition, the composition may comprise an agent that enhances its function, such as, for example, a cytotoxic agent, cytokine, chemotherapeutic agent, or growth-inhibitory agent. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles, and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences, *supra*.

The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, *e.g.*, films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and γ ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT ™ (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

G. Uses for anti-PRO Antibodies

The anti-PRO antibodies of the invention have various utilities. For example, anti-PRO antibodies may be used in diagnostic assays for PRO, *e.g.*, detecting its expression in specific cells, tissues, or serum. Various diagnostic assay techniques known in the art may be used, such as competitive binding assays, direct or indirect sandwich assays and immunoprecipitation assays conducted in either heterogeneous or homogeneous phases [Zola, Monoclonal Antibodies: A Manual of Techniques, CRC Press, Inc. (1987) pp. 147-158]. The antibodies used in the diagnostic assays can be labeled with a detectable moiety. The detectable moiety should be capable

of producing, either directly or indirectly, a detectable signal. For example, the detectable moiety may be a radioisotope, such as ^{3}H, ^{14}C, ^{32}P, ^{35}S, or ^{125}I, a fluorescent or chemiluminescent compound, such as fluorescein isothiocyanate, rhodamine, or luciferin, or an enzyme, such as alkaline phosphatase, beta-galactosidase or horseradish peroxidase. Any method known in the art for conjugating the antibody to the detectable moiety may be employed, including those methods described by Hunter et al., Nature, 144:945 (1962); David et al., Biochemistry, 13:1014 (1974); Pain et al., J. Immunol. Meth., 40:219 (1981); and Nygren, J. Histochem. and Cytochem., 30:407 (1982).

Anti-PRO antibodies also are useful for the affinity purification of PRO from recombinant cell culture or natural sources. In this process, the antibodies against PRO are immobilized on a suitable support, such a Sephadex resin or filter paper, using methods well known in the art. The immobilized antibody then is contacted with a sample containing the PRO to be purified, and thereafter the support is washed with a suitable solvent that will remove substantially all the material in the sample except the PRO, which is bound to the immobilized antibody. Finally, the support is washed with another suitable solvent that will release the PRO from the antibody.

The following examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way.

All patent and literature references cited in the present specification are hereby incorporated by reference in their entirety.

EXAMPLES

Commercially available reagents referred to in the examples were used according to manufacturer's instructions unless otherwise indicated. The source of those cells identified in the following examples, and throughout the specification, by ATCC accession numbers is the American Type Culture Collection, Rockville, Maryland.

EXAMPLE 1: Extracellular Domain Homology Screening to Identify Novel Polypeptides and cDNA Encoding Therefor

The extracellular domain (ECD) sequences (including the secretion signal sequence, if any) from about 950 known secreted proteins from the Swiss-Prot public database were used to search EST databases. The EST databases included public databases (e.g., Dayhoff, GenBank), and proprietary databases (e.g. LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA). The search was performed using the computer program BLAST or BLAST2 (Altschul, and Gish, Methods in Enzymology 266: 460-80 (1996); http://blast.wustl/edu/blast/README.html) as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequences. Those comparisons with a Blast score of 70 (or in some cases 90) or greater that did not encode known proteins were clustered and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington).

Using this extracellular domain homology screen, consensus DNA sequences were assembled relative to the other identified EST sequences. In addition, the consensus DNA sequences obtained were often (but not always) extended using repeated cycles of BLAST and phrap to extend the consensus sequence as far as possible

using the sources of EST sequences discussed above.

Based upon the consensus sequences obtained as described above, oligonucleotides were then synthesized and used to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for a PRO polypeptide. Forward (.f) and reverse (.r) PCR primers generally range from 20 to 30 nucleotides and are often designed to give a PCR product of about 100-1000 bp in length. The probe (.p) sequences are typically 40-55 bp in length. In some cases, additional oligonucleotides are synthesized when the consensus sequence is greater than about 1-1.5kbp. In order to screen several libraries for a full-length clone, DNA from the libraries was screened by PCR amplification, as per Ausubel et al., Current Protocols in Molecular Biology, with the PCR primer pair. A positive library was then used to isolate clones encoding the gene of interest using the probe oligonucleotide and one of the primer pairs.

The cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes et al., Science, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

EXAMPLE 2: Isolation of cDNA Clones Encoding PRO211 and PRO217

Consensus DNA sequences were assembled as described in Example 1 above and were designated as DNA28730 and DNA28760, respectively. Based on these consensus sequences, oligonucleotides were synthesized and used to identify by PCR a cDNA library that contained the sequences of interest and for use as probes to isolate a clone of the full-length coding sequence for the PRO211 and PRO217 polypeptides. The libraries used to isolate DNA32292-1131 and DNA33094-1131 were fetal lung libraries.

cDNA clones were sequenced in their entirety. The entire nucleotide sequences of PRO211 (DNA32292-1131) and PRO217 (UNQ191) are shown in Figure 1 (SEQ ID NO: 1) and Figure 3 (SEQ ID NO:3), respectively. The predicted polypeptides are 353 and 379 amino acid in length, respectively, with respective molecular weights of approximately 38,190 and 41,520 daltons.

The oligonucleotide sequences used in the above procedures were the following:

28730.p (OLI 516) (SEQ ID NO:5)

5'-AGGGAGCACGGACAGTGTGCAGATGTGGACGAGTGCTCACTAGCA-3'

28730.f (OLI 517) (SEQ ID NO:6)

5'-AGAGTGTATCTCTGGCTACGC-3'

28730.r (OLI 518) (SEQ ID NO:7)

5'-TAAGTCCGGCACATTACAGGTC-3'

28760.p (OLI 617) (SEQ ID NO:8)

5'-CCCACGATGTATGAATGGTGGACTTTGTGTGACTCCTGGTTTCTGCATC-3'

28760.f (OLI 618) (SEQ ID NO:9)

5'-AAAGACGCATCTGCGAGTGTCC-3'

28760.r (OLI 619) (SEQ ID NO:10)
5'-TGCTGATTTCACACTGCTCTCCC-3'

EXAMPLE 3: Isolation of cDNA Clones Encoding Human PRO230

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA30857. An EST proprietary to Genentech was employed in the consensus assembly. The EST is designated as DNA20088 and has the nucleotide sequence shown in Figure 7 (SEQ ID NO:13).

Based on the DNA30857 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO230.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-TTCGAGGCCTCTGAGAAGTGGCCC-3' (SEQ ID NO:14)
reverse PCR primer 5'-GGCGGTATCTCTCTGGCCTCCC-3' (SEQ ID NO:15)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30857 sequence which had the following nucleotide sequence

hybridization probe
5'-TTCTCCACAGCAGCTGTGGCATCCGATCGTGTCTCAATCCATTCTCTGGG-3' (SEQ ID NO:16)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO230 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO230 (herein designated as DNA33223-1136 and the derived protein sequence for PRO230.

The entire nucleotide sequence of DNA33223-1136 is shown in Figure 5 (SEQ ID NO:11). Clone DNA33223-1136 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 100-103 and ending at the stop codon at nucleotide positions 1501-1503 (Figure 5; SEQ ID NO:11). The predicted polypeptide precursor is 467 amino acids long (Figure 6).

EXAMPLE 4: Isolation of cDNA Clones Encoding Human PRO232

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA30935. Based on the DNA30935 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO232.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-TGCTGTGCTACTCCTGCAAAGCCC-3' (SEQ ID NO:19)
reverse PCR primer 5'-TGCACAAGTCGGTGTCACAGCACG-3' (SEQ ID NO:20)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30935

sequence which had the following nucleotide sequence

hybridization probe

5'-AGCAACGAGGACTGCCTGCAGGTGGAGAACTGCACCCAGCTGGG-3' (SEQ ID NO:21)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO232 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO232 [herein designated as DNA34435-1140] and the derived protein sequence for PRO232.

The entire nucleotide sequence of DNA34435-1140 is shown in Figure 8 (SEQ ID NO:17). Clone DNA34435-1140 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 17-19 and ending at the stop codon at nucleotide positions 359-361 (Fig. 8; SEQ ID NO:17). The predicted polypeptide precursor is 114 amino acids long (Fig. 9). Clone DNA34435-1140 has been deposited with ATCC on September 16, 1997 and is assigned ATCC deposit no. ATCC 209250.

Analysis of the amino acid sequence of the full-length PRO232 suggests that it possesses 35% sequence identity with a stem cell surface antigen from Gallus gallus.

EXAMPLE 5: Isolation of cDNA Clones Encoding PRO187

A proprietary expressed sequence tag (EST) DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) was searched and an EST (#843193) was identified which showed homology to fibroblast growth factor (FGF-8) also known as androgen-induced growth factor. mRNA was isolated from human fetal lung tissue using reagents and protocols from Invitrogen, San Diego, CA (Fast Track 2). The cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents (e.g., Invitrogen, San Diego, CA, Life Technologies, Gaithersburg, MD). The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into the cloning vector pRK5D using reagents and protocols from Life Technologies, Gaithersburg, MD (Super Script Plasmid System). The double-stranded cDNA was sized to greater than 1000 bp and the SalI/NotI linkered cDNA was cloned into XhoI/NotI cleaved vector. pRK5D is a cloning vector that has an sp6 transcription initiation site followed by an SfiI restriction enzyme site preceding the XhoI/NotI cDNA cloning sites.

Several libraries from various tissue sources were screened by PCR amplification with the following oligonucleotide probes:

IN843193.f (OLI315) (SEQ ID NO:24)

5'-CAGTACGTGAGGGACCAGGGCGCCATGA-3'

IN843193.r (OLI 317) (SEQ ID NO:25)

5'-CCGGTGACCTGCACGTGCTTGCCA-3'

A positive library was then used to isolate clones encoding the PRO187 gene using one of the above oligonucleotides and the following oligonucleotide probe:

IN843193.p (OLI 316) (SEQ ID NO:26)

5'-GCGGATCTGCCGCCTGCTCANCTGGTCGGTCATGGCGCCCT-3'

A cDNA clone was sequenced in entirety. The entire nucleotide sequence of PRO187 (DNA27864-1155) is shown in Figure 10 (SEQ ID NO:22). Clone DNA27864-1155 contains a single open reading frame with an apparent translational initiation site at nucleotide position 1 (Figure 10; SEQ ID NO:22). The predicted polypeptide precursor is 205 amino acids long. Clone DNA27864-1155 has been deposited with the ATCC (designation: DNA27864-1155) and is assigned ATCC deposit no. ATCC 209375.

Based on a BLAST and FastA sequence alignment analysis (using the ALIGN computer program) of the full-length sequence, the PRO187 polypeptide shows 74% amino acid sequence identity (Blast score 310) to human fibroblast growth factor-8 (androgen-induced growth factor).

EXAMPLE 6: Isolation of cDNA Clones Encoding PRO265

A consensus DNA sequence was assembled relative to other EST sequences as described in Example 1 above using phrap. This consensus sequence is herein designated DNA33679. Based on the DNA33679 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO265.

PCR primers (two forward and one reverse) were synthesized:

forward PCR primer A: 5'-CGGTCTACCTGTATGGCAACC-3' (SEQ ID NO:29);

forward PCR primer B: 5'-GCAGGACAACCAGATAAACCAC-3' (SEQ ID NO:30);

reverse PCR primer 5'-ACGCAGATTTGAGAAGGCTGTC-3' (SEQ ID NO:31)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA33679 sequence which had the following nucleotide sequence

hybridization probe

5'-TTCACGGGCTGCTCTTGCCCAGCTCTTGAAGCTTGAAGAGCTGCAC-3' (SEQ ID NO:32)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO265 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human a fetal brain library.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO265 [herein designated as DNA36350-1158] (SEQ ID NO:27) and the derived protein sequence for PRO265.

The entire nucleotide sequence of DNA36350-1158 is shown in Figure 12 (SEQ ID NO:27). Clone DNA36350-1158 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 352-354 and ending at the stop codon at positions 2332-2334 (Figure 12). The predicted polypeptide precursor is 660 amino acids long (Figure 13). Clone DNA36350-1158 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209378.

Analysis of the amino acid sequence of the full-length PRO265 polypeptide suggests that portions of it possess significant homology to the fibromodulin and the fibromodulin precursor, thereby indicating that PRO265 may be a novel member of the leucine rich repeat family, particularly related to fibromodulin.

EXAMPLE 7: Isolation of cDNA Clones Encoding Human PRO219

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA28729. Based on the DNA28729 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO219.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-GTGACCCTGGTTGTGAATACTCC-3' (SEQ ID NO:35)

reverse PCR primer 5'-ACAGCCATGGTCTATAGCTTGG-3' (SEQ ID NO:36)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28729 sequence which had the following nucleotide sequence

hybridization probe

5'-GCCTGTCAGTGTCCTGAGGGACACGTGCTCCGCAGCGATGGGAAG-3' (SEQ ID NO:37)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO219 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO219 [herein designated as DNA32290-1164] (SEQ ID NO:33) and the derived protein sequence for PRO219.

The entire nucleotide sequence of DNA32290-1164 is shown in Figure 14 (SEQ ID NO:33). Clone DNA32290-1164 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 204-206 and ending at the stop codon at nucleotide positions 2949-2951 (Figure 14). The predicted polypeptide precursor is 915 amino acids long (Figure 15). Clone DNA32290-1164 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209384.

Analysis of the amino acid sequence of the full-length PRO219 polypeptide suggests that portions of it possess significant homology to the mouse and human matrilin-2 precursor polypeptides.

EXAMPLE 8: Isolation of cDNA Clones Encoding Human PRO246

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA30955. Based on the DNA30955 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO246.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-AGGGTCTCCAGGAGAAAGACTC-3' (SEQ ID NO:40)

reverse PCR primer 5'-ATTGTGGGCCTTGCAGACATAGAC-3' (SEQ ID NO:41)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30955 sequence which had the following nucleotide sequence

hybridization probe

5'-GGCCACAGCATCAAAACCTTAGAACTCAATGTACTGGTTCCTCCAGCTCC-3' (SEQ ID NO:42)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO246 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO246 [herein designated as DNA35639-1172] (SEQ ID NO:38) and the derived protein sequence for PRO246.

The entire nucleotide sequence of DNA35639-1172 is shown in Figure 16 (SEQ ID NO:38). Clone DNA35639-1172 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 126-128 and ending at the stop codon at nucleotide positions 1296-1298 (Figure 16). The predicted polypeptide precursor is 390 amino acids long (Figure 17). Clone DNA35639-1172 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209396.

Analysis of the amino acid sequence of the full-length PRO246 polypeptide suggests that it possess significant homology to the human cell surface protein HCAR, thereby indicating that PRO246 may be a novel cell surface virus receptor.

EXAMPLE 9: Isolation of cDNA Clones Encoding Human PRO228

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA28758. An EST proprietary to Genentech was employed in the consensus assembly. This EST is shown in Figure 20 (SEQ ID NO:50) and is herein designated as DNA21951.

Based on the DNA28758 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO228.

PCR primers (forward and reverse) were synthesized:

forward PCR primer	5'-GGTAATGAGCTCCATTACAG-3' (SEQ ID NO:51)
forward PCR primer	5'-GGAGTAGAAAGCGCATGG-3' (SEQ ID NO:52)
forward PCR primer	5'-CACCTGATACCATGAATGGCAG-3' (SEQ ID NO:53)
reverse PCR primer	5'-CGAGCTCGAATTAATTCG-3' (SEQ ID NO:54)
reverse PCR primer	5'-GGATCTCCTGAGCTCAGG-3' (SEQ ID NO:55)
reverse PCR primer	5'-CCTAGTTGAGTGATCCTTGTAAG-3' (SEQ ID NO:56)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28758 sequence which had the following nucleotide sequence

hybridization probe

5'-ATGAGACCCACACCTCATGCCGCTGTAATCACCTGACACATTTGCAATT-3' (SEQ ID NO:57)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO228 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO228 [herein designated as DNA33092-1202] (SEQ ID NO:48) and the derived protein sequence for PRO228.

The entire nucleotide sequence of DNA33092-1202 is shown in Figure 18 (SEQ ID NO:48). Clone DNA33092-1202 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 24-26 of SEQ ID NO:48 and ending at the stop codon after nucleotide position 2093 of SEQ ID NO:48. The predicted polypeptide precursor is 690 amino acids long (Figure 19). Clone DNA33092-1202 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209420.

Analysis of the amino acid sequence of the full-length PRO228 polypeptide suggests that portions of it possess significant homology to the secretin-related proteins CD97 and EMR1 as well as the secretin member, latrophilin, thereby indicating that PRO228 may be a new member of the secretin related proteins.

EXAMPLE 10: Isolation of cDNA Clones Encoding Human PRO533

The EST sequence accession number AF007268, a murine fibroblast growth factor (FGF-15) was used to search various public EST databases (e.g., GenBank, Dayhoff, etc.). The search was performed using the computer program BLAST or BLAST2 [Altschul et al., Methods in Enzymology, 266:460-480 (1996); http://blast.wustl/edu/blast/README.html] as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequences. The search resulted in a hit with GenBank EST AA220994, which has been identified as stratagene NT2 neuronal precursor 937230.

Based on the Genbank EST AA220994 sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence. Forward and reverse PCR primers may range from 20 to 30 nucleotides (typically about 24), and are designed to give a PCR product of 100-1000 bp in length. The probe sequences are typically 40-55 bp (typically about 50) in length. In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification, as per *Ausubel et al., Current Protocols in Molecular Biology*, with the PCR primer pair. A positive library was then used to isolate clones encoding the gene of interest using the probe oligonucleotide and one of the PCR primers.

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified below. A positive library was then used to isolate clones encoding the PRO533 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal retina. The cDNA libraries used to isolated the cDNA clones were constructed by standard methods using commercially available reagents (e.g., Invitrogen, San Diego, CA; Clontech, etc.) The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes et al., *Science*, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

A cDNA clone was sequenced in its entirety. The full length nucleotide sequence of PRO533 is shown in Figure 21 (SEQ ID NO:58). Clone DNA49435-1219 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 459-461 (Figure 21; SEQ ID NO:58). The predicted polypeptide precursor is 216 amino acids long. Clone DNA47412-1219 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209480.

Based on a BLAST-2 and FastA sequence alignment analysis of the full-length sequence, PRO533 shows amino acid sequence identity to fibroblast growth factor (53%).

The oligonucleotide sequences used in the above procedure were the following:

FGF15.forward: 5'-ATCCGCCCAGATGGCTACAATGTGTA-3' (SEQ ID NO:60);

FGF15.probe: 5'-GCCTCCCGGTCTCCCTGAGCAGTGCCAAACAGCGGCAGTGTA-3' (SEQ ID NO:61);

FGF15.reverse: 5'-CCAGTCCGGTGACAAGCCCAAA-3' (SEQ ID NO:62).

EXAMPLE 11: Isolation of cDNA Clones Encoding Human PRO245

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA30954.

Based on the DNA30954 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO245.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-ATCGTTGTGAAGTTAGTGCCCC-3' (SEQ ID NO:65)

reverse PCR primer 5'-ACCTGCGATATCCAACAGAATTG-3' (SEQ ID NO:66)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30954 sequence which had the following nucleotide sequence

hybridization probe

5'-GGAAGAGGATACAGTCACTCTGGAAGTATTAGTGGCTCCAGCAGTTCC-3' (SEQ ID NO:67)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO245 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO245 [herein designated as DNA35638-1141] and the derived protein sequence for PRO245.

The entire nucleotide sequence of DNA35638-1141 is shown in Figure 23 (SEQ ID NO:63). Clone DNA35638-1141 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 89-91 and ending at the stop codon at nucleotide positions 1025-1027 (Fig. 23; SEQ ID NO:63). The predicted polypeptide precursor is 312 amino acids long (Fig. 24). Clone DNA35638-1141 has been deposited with ATCC on September 16, 1997 and is assigned ATCC deposit no. ATCC 209265.

Analysis of the amino acid sequence of the full-length PRO245 suggests that a portion of it possesses 60% amino acid identity with the human c-myb protein and, therefore, may be a new member of the transmembrane protein receptor tyrosine kinase family.

EXAMPLE 12: Isolation of cDNA Clones Encoding Human PRO220, PRO221 and PRO227

(a) PRO220

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28749. Based on the DNA28749 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO220.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-TCACCTGGAGCCTTTATTGGCC-3' (SEQ ID NO:74)

reverse PCR primer 5'-ATACCAGCTATAACCAGGCTGCG-3' (SEQ ID NO:75)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28749 sequence which had the following nucleotide sequence:

hybridization probe

5'-CAACAGTAAGTGGTTTGATGCTCTTCCAAATCTAGAGATTCTGATGATTGGG-3' (SEQ ID NO:76).

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO220 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO220 [herein designated as DNA32298-1132 and the derived protein sequence for PRO220.

The entire nucleotide sequence of DNA32298-1132 is shown in Figure 25 (SEQ ID NO:68). Clone DNA32298-1132 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 480-482 and ending at the stop codon at nucleotide positions 2604-2606 (Figure 25). The predicted polypeptide precursor is 708 amino acids long (Figure 26). Clone DNA32298-1132 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209257.

Analysis of the amino acid sequence of the full-length PRO220 shows it has homology to member of the leucine rich repeat protein superfamily, including the leucine rich repeat protein and the neuronal leucine-rich repeat protein 1.

(b) PRO221

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28756. Based on the DNA28756 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO221.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CCATGTGTCTCCTCCTACAAAG-3' (SEQ ID NO:77)

reverse PCR primer 5'-GGGAATAGATGTGATCTGATTGG-3' (SEQ ID NO:78)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28756 sequence which had the following nucleotide sequence:

hybridization probe

5'-CACCTGTAGCAATGCAAATCTCAAGGAAATACCTAGAGATCTTCCTCCTG-3' (SEQ ID NO:79)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO221 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO221 [herein designated as DNA33089-1132 and the derived protein sequence for PRO221.

The entire nucleotide sequence of DNA33089-1132 is shown in Figure 27 (SEQ ID NO:70). Clone DNA33089-1132 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 179-181 and ending at the stop codon at nucleotide positions 956-958 (Figure 27). The predicted polypeptide precursor is 259 amino acids long (Figure 28). PRO221 is believed to have a transmembrane region at amino acids 206-225. Clone DNA33089-1132 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209262.

Analysis of the amino acid sequence of the full-length PRO221 shows it has homology to member of the leucine rich repeat protein superfamily, including the SLIT protein.

(c) PRO227

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28740. Based on the DNA28740 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO227.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-AGCAACCGCCTGAAGCTCATCC-3' (SEQ ID NO:80)

reverse PCR primer 5'-AAGGCGCGGTGAAAGATGTAGACG-3' (SEQ ID NO:81)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28740 sequence which had the following nucleotide sequence:

hybridization probe

5'GACTACATGTTTCAGGACCTGTACAACCTCAAGTCACTGGAGGTTGGCGA-3' (SEQ ID NO:82).

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO227 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO227 [herein designated as DNA33786-1132 and the derived protein sequence for PRO227.

The entire nucleotide sequence of DNA33786-1132 is shown in Figure 29 (SEQ ID NO:72). Clone

DNA33786-1132 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 33-35 and ending at the stop codon at nucleotide positions 1893-1895 (Figure 29). The predicted polypeptide precursor is 620 amino acids long (Figure 30). PRO227 is believed to have a transmembrane region. Clone DNA33786-1132 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209253.

Analysis of the amino acid sequence of the full-length PRO221 shows it has homology to member of the leucine rich repeat protein superfamily, including the platelet glycoprotein V precursor and the human glycoprotein V.

EXAMPLE 13: Isolation of cDNA Clones Encoding Human PRO258

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA28746.

Based on the DNA28746 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO258.

PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-GCTAGGAATTCCACAGAAGCCC-3' (SEQ ID NO:85)
reverse PCR primer 5'-AACCTGGAATGTCACCGAGCTG-3' (SEQ ID NO:86)
reverse PCR primer 5'-CCTAGCACAGTGACGAGGGACTTGGC-3' (SEQ ID NO:87)

Additionally, synthetic oligonucleotide hybridization probes were constructed from the consensus DNA28740 sequence which had the following nucleotide sequence:

hybridization probe

5'-AAGACACAGCCACCCTAAACTGTCAGTCTTCTGGGAGCAAGCCTGCAGCC-3' (SEQ ID NO:88)
5'-GCCCTGGCAGACGAGGGCGAGTACACCTGCTCAATCTTCACTATGCCTGT-3' (SEQ ID NO:89)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO258 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO258 [herein designated as DNA35918-1174] (SEQ ID NO:83) and the derived protein sequence for PRO258.

The entire nucleotide sequence of DNA35918-1174 is shown in Figure 31 (SEQ ID NO:83). Clone DNA35918-1174 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 147-149 of SEQ ID NO:83 and ending at the stop codon after nucleotide position 1340 of SEQ ID NO:83 (Figure 31). The predicted polypeptide precursor is 398 amino acids long (Figure 32). Clone DNA35918-1174 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209402.

Analysis of the amino acid sequence of the full-length PRO258 polypeptide suggests that portions of it possess significant homology to the CRTAM and the poliovirus receptor and have an Ig domain, thereby indicating that PRO258 is a new member of the Ig superfamily.

EXAMPLE 14: Isolation of cDNA Clones Encoding Human PRO266

An expressed sequence tag database was searched for ESTs having homology to SLIT, resulting in the identification of a single EST sequence designated herein as T73996. Based on the T73996 EST sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO266.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-GTTGGATCTGGGCAACAATAAC-3' (SEQ ID NO:92)

reverse PCR primer 5'-ATTGTTGTGCAGGCTGAGTTTAAG-3' (SEQ ID NO:93)

Additionally, a synthetic oligonucleotide hybridization probe was constructed which had the following nucleotide sequence

hybridization probe

5'-GGTGGCTATACATGGATAGCAATTACCTGGACACGCTGTCCCGGG-3' (SEQ ID NO:94)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO266 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO266 [herein designated as DNA37150-1178] (SEQ ID NO:90) and the derived protein sequence for PRO266.

The entire nucleotide sequence of DNA37150-1178 is shown in Figure 33 (SEQ ID NO:90). Clone DNA37150-1178 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 167-169 and ending at the stop codon after nucleotide position 2254 of SEQ ID NO:90. The predicted polypeptide precursor is 696 amino acids long (Figure 34). Clone DNA37150-1178 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209401.

Analysis of the amino acid sequence of the full-length PRO266 polypeptide suggests that portions of it possess significant homology to the SLIT protein, thereby indicating that PRO266 may be a novel leucine rich repeat protein.

EXAMPLE 15: Isolation of cDNA Clones Encoding Human PRO269

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35705. Based on the DNA35705 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO269.

Forward and reverse PCR primers were synthesized:

forward PCR primer (.f1) 5'-TGGAAGGAGATGCGATGCCACCTG -3' (SEQ ID NO:97)

forward PCR primer (.f2) 5'-TGACCAGTGGGGAAGGACAG-3' (SEQ ID NO:98)

forward PCR primer (.f3) 5'-ACAGAGCAGAGGGTGCCTTG-3' (SEQ ID NO:99)

reverse PCR primer (.r1) 5'-TCAGGGACAAGTGGTGTCTCTCCC-3'

(SEQ ID NO:100)

reverse PCR primer (.r2) 5'-TCAGGGAAGGAGTGTGCAGTTCTG-3'

(SEQ ID NO:101)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35705 sequence which had the following nucleotide sequence:

hybridization probe

5'-ACAGCTCCCGATCTCAGTTACTTGCATCGCGGACGAAATCGGCGCTCGCT-3' (SEQ ID NO:102)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO269 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO269 [herein designated as DNA38260-1180] (SEQ ID NO:95) and the derived protein sequence for PRO269.

The entire nucleotide sequence of DNA38260-1180 is shown in Figure 35 (SEQ ID NO:95). Clone DNA38260-1180 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 314-316 and ending at the stop codon at nucleotide positions 1784-1786 (Fig. 35; SEQ ID NO:95). The predicted polypeptide precursor is 490 amino acids long (Fig. 36). Clone DNA38260-1180 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209397.

Analysis of the amino acid sequence of the full-length PRO269 suggests that portions of it possess significant homology to the human thrombomodulin proteins, thereby indicating that PRO269 may possess one or more thrombomodulin-like domains.

EXAMPLE 16: Isolation of cDNA Clones Encoding Human PRO287

A consensus DNA sequence encoding PRO287 was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28728. Based on the DNA28728 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO287.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CCGATTCATAGACCTCGAGAGT-3' (SEQ ID NO:105)

reverse PCR primer 5'-GTCAAGGAGTCCTCCACAATAC-3' (SEQ ID NO:106)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28728 sequence which had the following nucleotide sequence

hybridization probe

5'-GTGTACAATGGCCATGCCAATGGCCAGCGCATTGGCCGCTTCTGT-3'

(SEQ ID NO:107)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to

isolate clones encoding the PRO287 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO287 [herein designated as DNA39969-1185, SEQ ID NO:103] and the derived protein sequence for PRO287.

The entire nucleotide sequence of DNA39969-1185 is shown in Figure 37 (SEQ ID NO:103). Clone DNA39969-1185 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 307-309 and ending at the stop codon at nucleotide positions 1552-1554 (Fig. 37; SEQ ID NO:103). The predicted polypeptide precursor is 415 amino acids long (Fig. 38). Clone DNA39969-1185 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209400.

Analysis of the amino acid sequence of the full-length PRO287 suggests that it may possess one or more procollagen C-proteinase enhancer protein precursor or procollagen C-proteinase enhancer protein-like domains. Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO287 shows nucleic acid sequence identity to procollagen C-proteinase enhancer protein precursor and procollagen C-proteinase enhancer protein (47 and 54%, respectively).

EXAMPLE 17: Isolation of cDNA Clones Encoding Human PRO214

A consensus DNA sequence was assembled using phrap as described in Example 1 above. This consensus DNA sequence is designated herein as DNA28744. Based on this consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence.

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified below. A positive library was then used to isolate clones encoding the PRO214 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue.

A cDNA clone was sequenced in its entirety. The full length nucleotide sequence of DNA32286-1191 is shown in Figure 39 (SEQ ID NO:108). DNA32286-1191 contains a single open reading frame with an apparent translational initiation site at nucleotide position 103 (Fig. 39; SEQ ID NO:108). The predicted polypeptide precursor is 420 amino acids long (SEQ ID NO:109).

Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO214 polypeptide shows amino acid sequence identity to HT protein and/or Fibulin (49% and 38%, respectively).

The oligonucleotide sequences used in the above procedure were the following:

28744.p (OLI555)

5'-CCTGGCTATCAGCAGGTGGGCTCCAAGTGTCTCGATGTGGATGAGTGTGA-3' (SEQ ID NO:110)

28744.f (OLI556)

5'-ATTCTGCGTGAACACTGAGGGC-3' (SEQ ID NO:111)

28744.r (OLI557)

5'-ATCTGCTTGTAGCCCTCGGCAC-3' (SEQ ID NO:112)

EXAMPLE 18: Isolation of cDNA Clones Encoding Human PRO317

A consensus DNA sequence was assembled using phrap as described in Example 1 above, wherein the consensus sequence is herein designated as DNA28722. Based on this consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence. The forward and reverse PCR primers, respectively, synthesized for this purpose were:

5'-AGGACTGCCATAACTTGCCTG (OLI489) (SEQ ID NO:115) and

5'-ATAGGAGTTGAAGCAGCGCTGC (OLI490) (SEQ ID NO:116).

The probe synthesized for this purpose was:

5'-TGTGTGGACATAGACGAGTGCCGCTACCGCTACTGCCAGCACCGC (OLI488) (SEQ ID NO:117)

mRNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification, as per Ausubel *et al.*, *Current Protocols in Molecular Biology* (1989), with the PCR primer pair identified above. A positive library was then used to isolate clones containing the PRO317 gene using the probe oligonucleotide identified above and one of the PCR primers.

A cDNA clone was sequenced in its entirety. The entire nucleotide sequence of DNA33461-1199 (encoding PRO317) is shown in Figure 41 (SEQ ID NO:113). Clone DNA33461-1199 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 68-70 (Fig. 41; SEQ ID NO:113). The predicted polypeptide precursor is 366 amino acids long. The predicted signal sequence is amino acids 1-18 of Figure 42 (SEQ ID NO:114). There is one predicted N-linked glycosylation site at amino acid residue 160. Clone DNA33461-1199 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209367.

Based on BLAST™ and FastA™ sequence alignment analysis (using the ALIGN™ computer program) of the full-length PRO317sequence, PRO317 shows the most amino acid sequence identity to EBAF-1 (92%). The results also demonstrate a significant homology between human PRO317 and mouse LEFTY protein. The C-terminal end of the PRO317 protein contains many conserved sequences consistent with the pattern expected of a member of the TGF- superfamily.

In situ expression analysis in human tissues performed as described below evidences that there is distinctly strong expression of the PRO317 polypeptide in pancreatic tissue.

EXAMPLE 19: Isolation of cDNA clones Encoding Human PRO301

A consensus DNA sequence designated herein as DNA35936 was assembled using phrap as described in Example 1 above. Based on this consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence.

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified below. A positive library was then used to isolate clones encoding the PRO301 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney.

A cDNA clone was sequenced in its entirety. The full length nucleotide sequence of native sequence PRO301 is shown in Figure 43 (SEQ ID NO:118). Clone DNA40628-1216 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 52-54 (Fig. 43; SEQ ID NO:118). The predicted polypeptide precursor is 299 amino acids long with a predicted molecular weight of 32,583 daltons and pI of 8.29. Clone DNA40628-1216 has been deposited with ATCC and is assigned ATCC deposit No. ATCC 209432.

Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO301 shows amino acid sequence identity to A33 antigen precursor (30%) and coxsackie and adenovirus receptor protein (29%).

The oligonucleotide sequences used in the above procedure were the following:

OLI2162 (35936.f1) 5'-TCGCGGAGCTGTGTTCTGTTTCCC-3' (SEQ ID NO:120)
OLI2163 (35936.p1)
5'-TGATCGCGATGGGGACAAAGGCGCAAGCTCGAGAGGAAACTGTTGTGCCT-3' (SEQ ID NO:121)
OLI2164 (35936.f2)
5'-ACACCTGGTTCAAAGATGGG-3' (SEQ ID NO:122)
OLI2165 (35936.r1)
5'-TAGGAAGAGTTGCTGAAGGCACGG-3' (SEQ ID NO:123)
OLI2166 (35936.f3)
5'-TTGCCTTACTCAGGTGCTAC-3' (SEQ ID NO:124)
OLI2167 (35936.r2)
5'-ACTCAGCAGTGGTAGGAAAG-3' (SEQ ID NO:125)

EXAMPLE 20: Isolation of cDNA Clones Encoding Human PRO224

A consensus DNA sequence assembled relative to the other identified EST sequences as described in Example 1, wherein the consensus sequence is designated herein as DNA30845. Based on the DNA30845 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO224.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-AAGTTCCAGTGCCGCACCAGTGGC-3' (SEQ ID NO:128)
reverse PCR primer 5'-TTGGTTCCACAGCCGAGCTCGTCG-3' (SEQ ID NO:129)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30845 sequence which had the following nucleotide sequence

hybridization probe
5'-GAGGAGGAGTGCAGGATTGAGCCATGTACCCAGAAAGGGCAATGCCCACC-3' (SEQ ID NO:130)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO224 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO224 [herein designated as DNA33221-1133] and the derived protein sequence for PRO224.

The entire nucleotide sequence of DNA33221-1133 is shown in Figure 45 (SEQ ID NO:126). Clone DNA33221-1133 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 33-35 and ending at the stop codon at nucleotide positions 879-899 (Figure 45; SEQ ID NO:126). The start of a transmembrane region begins at nucleotide position 777. The predicted polypeptide precursor is 282 amino acids long (Figure 46). Clone DNA33221-1133 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209263.

Analysis of the amino acid sequence of the full-length PRO224 suggests that it has homology to very low-density lipoprotein receptors, apolipoprotein E receptor and chicken oocyte receptors P95. Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO224 has amino acid identity to portions of these proteins in the range from 28% to 45%, and overall identity with these proteins in the range from 33% to 39%.

EXAMPLE 21: Isolation of cDNA Clones Encoding Human PRO222

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28771. Based on the DNA28771 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO222.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-ATCTCCTATCGCTGCTTTCCCGG-3' (SEQ ID NO:133)

reverse PCR primer 5'-AGCCAGGATCGCAGTAAAACTCC-3' (SEQ ID NO:134)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28771 sequence which had the following nucleotide sequence:

hybridization probe

5'-ATTTAAACTTGATGGGTCTGCGTATCTTGAGTGCTTACAAAACCTTATCT-3' (SEQ ID NO:135)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO222 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO222 [herein designated as DNA33107-1135] and the derived protein sequence for PRO222.

The entire nucleotide sequence of DNA33107-1135 is shown in Figure 47 (SEQ ID NO:131). Clone DNA33107-1135 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 159-161 and ending at the stop codon at nucleotide positions 1629-1631 (Fig. 47; SEQ ID NO:131). The predicted polypeptide precursor is 490 amino acids long (Fig. 48). Clone DNA33107-1135 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209251.

Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO222 shows amino acid sequence identity to mouse complement factor h precursor (25-26%), complement receptor (27-29%),

mouse complement C3b receptor type 2 long form precursor (25-47%) and human hypothetical protein kiaa0247 (40%).

EXAMPLE 22: Isolation of cDNA clones Encoding PRO234

A consensus DNA sequence was assembled (DNA30926) using phrap as described in Example 1 above. Based on this consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence.

RNA for the construction of the cDNA libraries was isolated using standard isolation protocols, e.g., Ausubel *et al., Current Protocols in Molecular Biology*, from tissue or cell line sources or it was purchased from commercial sources (e.g., Clontech). The cDNA libraries used to isolate the cDNA clones were constructed by standard methods (e.g., Ausubel *et al.*) using commercially available reagents (e.g., Invitrogen). This library was derived from 22 week old fetal brain tissue.

A cDNA clone was sequenced in its entirety. The entire nucleotide sequence of PRO234 is shown in Figure 49 (SEQ ID NO:136). The predicted polypeptide precursor is 382 amino acids long and has a calculated molecular weight of approximately 43.1 kDa.

The oligonucleotide sequences used in the above procedure were the following:

30926.p (OLI826) (SEQ ID NO:138): 5'-GTTCATTGAAAACCTCTTGCCATCT GATGGTGACTTCTGGATTGGGCTCA-3'

30926.f (OLI827) (SEQ ID NO:139): 5'-AAGCCAAAGAAGCCTGCAGGAGGG-3'

30926.r (OLI828) (SEQ ID NO:140): 5'-CAGTCCAAGCATAAAGGTCCTGGC-3'

EXAMPLE 23: Isolation of cDNA Clones Encoding Human PRO231

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence was designated herein as DNA30933. Based on the DNA30933 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO231.

Three PCR primers (two forward and one reverse) were synthesized:

forward PCR primer 1 5'-CCAACTACCAAAGCTGCTGGAGCC-3' (SEQ ID NO:143)

forward PCR primer 2 5'-GCAGCTCTATTACCACGGGAAGGA-3' (SEQ ID NO:144)

reverse PCR primer 5'-TCCTTCCCGTGGTAATAGAGCTGC-3' (SEQ ID NO:145)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30933 sequence which had the following nucleotide sequence

hybridization probe

5'-GGCAGAGAACCAGAGGCCGGAGGAGACTGCCTCTTTACAGCCAGG-3' (SEQ ID NO:146)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO231 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO231 [herein designated as DNA34434-1139] and the derived protein sequence for PRO231.

The entire nucleotide sequence of DNA34434-1139 is shown in Figure 51 (SEQ ID NO:141). Clone DNA34434-1139 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 173-175 and ending at the stop codon at nucleotide positions 1457-1459 (Fig. 51; SEQ ID NO:141). The predicted polypeptide precursor is 428 amino acids long (Fig. 52). Clone DNA34434-1139 has been deposited with ATCC on September 16, 1997 and is assigned ATCC deposit no. ATCC 209252.

Analysis of the amino acid sequence of the full-length PRO231 suggests that it possesses 30% and 31% amino acid identity with the human and rat prostatic acid phosphatase precursor proteins, respectively.

EXAMPLE 24: Isolation of cDNA Clones Encoding Human PRO229

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA28762. Based on the DNA28762 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO229.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-TTCAGCTCATCACCTTCACCTGCC-3' (SEQ ID NO:149)

reverse PCR primer 5'-GGCTCATACAAAATACCACTAGGG-3' (SEQ ID NO:150)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28762 sequence which had the following nucleotide sequence

hybridization probe

5'-GGGCCTCCACCGCTGTGAAGGGCGGGTGGAGGTGGAACAGAAAGGCCAGT-3' (SEQ ID NO:151)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO229 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO229 [herein designated as DNA33100-1159] (SEQ ID NO:147) and the derived protein sequence for PRO229.

The entire nucleotide sequence of DNA33100-1159 is shown in Figure 53 (SEQ ID NO:147). Clone DNA33100-1159 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 98-100 and ending at the stop codon at nucleotide positions 1139-1141 (Figure 53). The predicted polypeptide precursor is 347 amino acids long (Figure 54). Clone DNA33100-1159 has been deposited with ATCC and is assigned ATCC deposit no.ATCC 209377

Analysis of the amino acid sequence of the full-length PRO229 polypeptide suggests that portions of it possess significant homology to antigen wc1.1, M130 antigen and CD6.

EXAMPLE 25: Isolation of cDNA Clones Encoding Human PRO238

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described above in Example 1. This consensus sequence is herein designated DNA30908. Based on the DNA30908 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO238.

PCR primers (forward and reverse) were synthesized:

forward PCR primer 1 5'-GGTGCTAAACTGGTGCTCTGTGGC-3' (SEQ ID NO:154)

forward PCR primer 2 5'-CAGGGCAAGATGAGCATTCC-3' (SEQ ID NO:155)

reverse PCR primer 5'-TCATACTGTTCCATCTCGGCACGC-3' (SEQ ID NO:156)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30908 sequence which had the following nucleotide sequence

hybridization probe

5'-AATGGTGGGGCCCTAGAAGAGCTCATCAGAGAACTCACCGCTTCTCATGC-3' (SEQ ID NO:157)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO238 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO238 and the derived protein sequence for PRO238.

The entire nucleotide sequence of DNA35600-1162 is shown in Figure 55 (SEQ ID NO:152). Clone DNA35600-1162 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 134-136 and ending prior to the stop codon at nucleotide positions 1064-1066 (Figure 55). The predicted polypeptide precursor is 310 amino acids long (Figure 56). Clone DNA35600-1162 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209370.

Analysis of the amino acid sequence of the full-length PRO238 polypeptide suggests that portions of it possess significant homology to reductase, particularly oxidoreductase, thereby indicating that PRO238 may be a novel reductase.

EXAMPLE 26: Isolation of cDNA Clones Encoding Human PRO233

The extracellular domain (ECD) sequences (including the secretion signal, if any) of from about 950 known secreted proteins from the Swiss-Prot public protein database were used to search expressed sequence tag (EST) databases. The EST databases included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA). The search was performed using the computer program BLAST or BLAST2 (Altshul et al., Methods in Enzymology 266:460-480 (1996)) as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequence. Those comparisons resulting in a BLAST score of 70 (or in some cases 90) or greater that did not encode known proteins were clustered and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington; http://bozeman.mbt.washington.edu/phrap.docs/phrap.html).

An expressed sequence tag (EST) was identified by the EST database search and a consensus DNA sequence was assembled relative to other EST sequences using phrap. This consensus sequence is herein designated DNA30945. Based on the DNA30945 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO233.

Forward and reverse PCR primers were synthesized:

forward PCR primer 5'-GGTGAAGGCAGAAATTGGAGATG-3' (SEQ ID NO:160)

reverse PCR primer 5'-ATCCCATGCATCAGCCTGTTTACC-3' (SEQ ID NO:161)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30945 sequence which had the following nucleotide sequence

hybridization probe

5'-GCTGGTGTAGTCTATACATCAGATTTGTTTGCTACACAAGATCCTCAG-3'

(SEQ ID NO:162)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO233 gene using the probe oligonucleotide.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO233 [herein designated as DNA34436-1238] (SEQ ID NO:158) and the derived protein sequence for PRO233.

The entire nucleotide sequence of DNA34436-1238 is shown in Figure 57 (SEQ ID NO:158). Clone DNA34436-1238 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 101-103 and ending at the stop codon at nucleotide positions 1001-1003 (Figure 57). The predicted polypeptide precursor is 300 amino acids long (Figure 58). The full-length PRO233 protein shown in Figure 58 has an estimated molecular weight of about 32,964 daltons and a pI of about 9.52. Clone DNA34436-1238 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209523.

Analysis of the amino acid sequence of the full-length PRO233 polypeptide suggests that portions of it possess significant homology to reductase proteins, thereby indicating that PRO233 may be a novel reductase.

EXAMPLE 27: Isolation of cDNA Clones Encoding Human PRO223

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA30836. Based on the DNA30836 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO223.

PCR primer pairs (one forward and two reverse) were synthesized:

forward PCR primer 5'-TTCCATGCCACCTAAGGGAGACTC-3' (SEQ ID NO:165)

reverse PCR primer 1 5'-TGGATGAGGTGTGCAATGGCTGGC-3' (SEQ ID NO:166)

reverse PCR primer 2 5'-AGCTCTCAGAGGCTGGTCATAGGG-3' (SEQ ID NO:167)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30836 sequence which had the following nucleotide sequence

hybridization probe

5'-GTCGGCCCTTTCCCAGGACTGAACATGAAGAGTTATGCCGGCTTCCTCAC-3' (SEQ ID NO:168)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO223 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO223 [herein designated as DNA33206-1165] (SEQ ID NO:163) and the derived protein sequence for PRO223.

The entire nucleotide sequence of DNA33206-1165 is shown in Figure 59 (SEQ ID NO:163). Clone DNA33206-1165 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 97-99 and ending at the stop codon at nucleotide positions 1525-1527 (Figure 59). The predicted polypeptide precursor is 476 amino acids long (Figure 60). Clone DNA33206-1165 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209372.

Analysis of the amino acid sequence of the full-length PRO223 polypeptide suggests that it possesses significant homology to various serine carboxypeptidase proteins, thereby indicating that PRO223 may be a novel serine carboxypeptidase.

EXAMPLE 28: Isolation of cDNA Clones Encoding Human PRO235

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated "DNA30927". Based on the DNA30927 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO235.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer	5'-TGGAATACCGCCTCCTGCAG-3'	(SEQ ID NO:171)
reverse PCR primer	5'-CTTCTGCCCTTTGGAGAAGATGGC-3'	(SEQ ID NO:172)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30927 sequence which had the following nucleotide sequence

hybridization probe

5'-GGACTCACTGGCCCAGGCCTTCAATATCACCAGCCAGGACGAT-3' (SEQ ID NO:173)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO235 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO235 [herein designated as DNA35558-1167] (SEQ ID NO:169) and the derived protein sequence for

PRO235.

The entire nucleotide sequence of DNA35558-1167 is shown in Figure 61 (SEQ ID NO:169). Clone DNA35558-1167 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 667-669 and ending at the stop codon at nucleotide positions 2323-2325 (Figure 61). The predicted polypeptide precursor is 552 amino acids long (Figure 62). Clone DNA35558-1167 has been deposited with ATCC and is assigned ATCC deposit no. 209374.

Analysis of the amino acid sequence of the full-length PRO235 polypeptide suggests that portions of it possess significant homology to the human, mouse and *Xenopus* plexin protein, thereby indicating that PRO235 may be a novel plexin protein.

EXAMPLE 29: Isolation of cDNA Clones Encoding Human PRO236 and Human PRO262

Consensus DNA sequences were assembled relative to other EST sequences using phrap as described in Example 1 above. These consensus sequences are herein designated DNA30901 and DNA30847. Based on the DNA30901 and DNA30847 consensus sequences, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO236 and PRO262, respectively.

Based upon the DNA30901 consensus sequence, a pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-TGGCTACTCCAAGACCCTGGCATG-3' (SEQ ID NO:178)

reverse PCR primer 5'-TGGACAAATCCCCTTGCTCAGCCC-3' (SEQ ID NO:179)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30901 sequence which had the following nucleotide sequence

hybridization probe

5'-GGGCTTCACCGAAGCAGTGGACCTTTATTTTGACCACCTGATGTCCAGGG-3' (SEQ ID NO:180)

Based upon the DNA30847 consensus sequence, a pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CCAGCTATGACTATGATGCACC-3' (SEQ ID NO:181)

reverse PCR primer 5'-TGGCACCCAGAATGGTGTTGGCTC-3' (SEQ ID NO:182)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30847 sequence which had the following nucleotide sequence

hybridization probe

5'-CGAGATGTCATCAGCAAGTTCCAGGAAGTTCCTTTGGGACCTTTACCTCC-3' (SEQ ID NO:183)

In order to screen several libraries for a source of full-length clones, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. Positive libraries were then used to isolate clones encoding the PRO236 and PRO262 genes using the probe oligonucleotides and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue for PRO236 and human fetal liver tissue for PRO262.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for

PRO236 [herein designated as DNA35599-1168] (SEQ ID NO:174), the derived protein sequence for PRO236, the full-length DNA sequence for PRO262 [herein designated as DNA36992-1168] (SEQ ID NO:176) and the derived protein sequence for PRO262.

The entire nucleotide sequence of DNA35599-1168 is shown in Figure 63 (SEQ ID NO:174). Clone DNA35599-1168 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 69-71 and ending at the stop codon at nucleotide positions 1977-1979 (Figure 63). The predicted polypeptide precursor is 636 amino acids long (Figure 64). Clone DNA35599-1168 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209373.

The entire nucleotide sequence of DNA36992-1168 is shown in Figure 65 (SEQ ID NO:176). Clone DNA36992-1168 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 240-242 and ending at the stop codon at nucleotide positions 2202-2204 (Figure 65). The predicted polypeptide precursor is 654 amino acids long (Figure 66). Clone DNA36992-1168 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209382.

Analysis of the amino acid sequence of the full-length PRO236 and PRO262 polypeptides suggests that portions of those polypeptides possess significant homology to β-galactosidase proteins derived from various sources, thereby indicating that PRO236 and PRO262 may be novel β-galactosidase homologs.

EXAMPLE 30: Isolation of cDNA Clones Encoding Human PRO239

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA30909. Based on the DNA30909 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO239.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CCTCCCTCTATTACCCATGTC-3' (SEQ ID NO:186)

reverse PCR primer 5'-GACCAACTTTCTCTGGGAGTGAGG-3' (SEQ ID NO:187)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30909 sequence which had the following nucleotide sequence

hybridization probe

5'-GTCACTTTATTTCTCTAACAACAAGCTCGAATCCTTACCAGTGGCAG-3'

(SEQ ID NO:188)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO239 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO239 [herein designated as DNA34407-1169] (SEQ ID NO:184) and the derived protein sequence for PRO239.

The entire nucleotide sequence of DNA34407-1169 is shown in Figure 67 (SEQ ID NO:184). Clone

DNA34407-1169 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 72-74 and ending at the stop codon at nucleotide positions 1575-1577 (Figure 67). The predicted polypeptide precursor is 501 amino acids long (Figure 68). Clone DNA34407-1169 has been deposited with ATCC and is assigned ATCC deposit no.ATCC 209383.

Analysis of the amino acid sequence of the full-length PRO239 polypeptide suggests that portions of it possess significant homology to the densin protein, thereby indicating that PRO239 may be a novel molecule in the densin family.

EXAMPLE 31: Isolation of cDNA Clones Encoding Human PRO257

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA28731. Based on the DNA28731 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO257.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-TCTCTATTCCAAACTGTGGCG-3' (SEQ ID NO:191)

reverse PCR primer 5'-TTTGATGACGATTCGAAGGTGG-3' (SEQ ID NO:192)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28731 sequence which had the following nucleotide sequence

hybridization probe

5'-GGAAGGATCCTTCACCAGCCCCAATTACCCAAAGCCGCATCCTGAGC-3' (SEQ ID NO:193)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO257 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO257 [herein designated as DNA35841-1173 (SEQ ID NO:189) and the derived protein sequence for PRO257.

The entire nucleotide sequence of DNA35841-1173 is shown in Figure 69 (SEQ ID NO:189). Clone DNA35841-1173 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 964-966 and ending at the stop codon at nucleotide positions 2785-2787 (Figure 69). The predicted polypeptide precursor is 607 amino acids long (Figure 70). Clone DNA35841-1173 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209403.

Analysis of the amino acid sequence of the full-length PRO257 polypeptide suggests that portions of it possess significant homology to the ebnerin protein, thereby indicating that PRO257 may be a novel protein member related to the ebnerin protein.

EXAMPLE 32: Isolation of cDNA Clones Encoding Human PRO260

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA30834. Based on the DNA30834 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO260.

PCR primers (forward and two reverse) were synthesized:

forward PCR primer: 5'-TGGTTTGACCAGGCCAAGTTCGG-3' (SEQ ID NO:196);

reverse PCR primer A: 5'-GGATTCATCCTCAAGGAAGAGCGG-3' (SEQ ID NO:197); and

reverse PCR primer B: 5'AACTTGCAGCATCAGCCACTCTGC-3' (SEQ ID NO:198)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30834 sequence which had the following nucleotide sequence:

hybridization probe:

5'-TTCCGTGCCCAGCTTCGGTAGCGAGTGGTTCTGGTGGTATTGGCA-3' (SEQ ID NO:199)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO260 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO260 [herein designated as DNA33470-1175] (SEQ ID NO:194) and the derived protein sequence for PRO260.

The entire nucleotide sequence of DNA33470-1175 is shown in Figure 71 (SEQ ID NO:194). Clone DNA33470-1175 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 67-69 and ending at the stop codon 1468-1470 (see Figure 71). The predicted polypeptide precursor is 467 amino acids long (Figure 72). Clone DNA33470-1175 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209398.

Analysis of the amino acid sequence of the full-length PRO260 polypeptide suggests that portions of it possess significant homology to the alpha-l-fucosidase precursor, thereby indicating that PRO260 may be a novel fucosidase.

EXAMPLE 33: Isolation of cDNA Clones Encoding Human PRO263

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA30914. Based on the DNA30914 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO263.

PCR primers (tow forward and one reverse) were synthesized:

forward PCR primer 1: 5'-GAGCTTTCCATCCAGGTGTCATGC-3' (SEQ ID NO:202);

forward PCR primer 2: 5'-GTCAGTGACAGTACCTACTCGG-3' (SEQ ID NO:203); reverse PCR primer:

5'-TGGAGCAGGAGGAGTAGTAGTAGG-3' (SEQ ID NO:204)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30914 sequence which had the following nucleotide sequence:

hybridization probe:

5'-AGGAGGCCTGTAGGCTGCTGGGACTAAGTTTGGCCGGCAAGGACCAAGTT-3' (SEQ ID NO:205)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO263 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO263 [herein designated as DNA34431-1177] (SEQ ID NO:200) and the derived protein sequence for PRO263.

The entire nucleotide sequence of DNA34431-1177 is shown in Figure 73 (SEQ ID NO:200). Clone DNA34431-1177 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 160-162 of SEQ ID NO:200 and ending at the stop codon after the nucleotide at position 1126-1128 of SEQ ID NO:200 (Figure 73). The predicted polypeptide precursor is 322 amino acids long (Figure 74). Clone DNA34431-1177 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209399.

Analysis of the amino acid sequence of the full-length PRO263 polypeptide suggests that portions of it possess significant homology to CD44 antigen, thereby indicating that PRO263 may be a novel cell surface adhesion molecule.

EXAMPLE 34: Isolation of cDNA Clones Encoding Human PRO270

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence was designated herein as DNA35712. Based on the DNA35712 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO270. Forward and reverse PCR primers were synthesized:

forward PCR primer (.f1) 5'-GCTTGGATATTCGCATGGGCCTAC-3' (SEQ ID NO:208)

forward PCR primer (.f2) 5'-TGGAGACAATATCCCTGAGG-3' (SEQ ID NO:209)

reverse PCR primer (.r1) 5'-AACAGTTGGCCACAGCATGGCAGG-3' (SEQ ID NO:210)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35712 sequence which had the following nucleotide sequence

hybridization probe

5'-CCATTGATGAGGAACTAGAACGGGACAAGAGGGTCACTTGGATTGTGGAG-3'

(SEQ ID NO:211)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO270 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO270 [herein designated as DNA39510-1181] (SEQ ID NO:206) and the derived protein sequence for PRO270.

The entire nucleotide sequence of DNA39510-1181 is shown in Figure 75 (SEQ ID NO:206). Clone DNA39510-1181 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 3-5 and ending at the stop codon at nucleotide positions 891-893 (Fig. 75; SEQ ID NO:206). The predicted polypeptide precursor is 296 amino acids long (Fig. 76). Clone DNA39510-1181 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209392.

Analysis of the amino acid sequence of the full-length PRO270 suggests that portions of it possess significant homology to the thioredoxin-protein, thereby indicating that the PRO270 protein may be a novel member of the thioredoxin family.

EXAMPLE 35: Isolation of cDNA Clones Encoding Human PRO271

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35737. Based on the DNA35737 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO271.

Forward and reverse PCR primers were synthesized:

forward PCR primer 1 5'-TGCTTCGCTACTGCCCTC-3' (SEQ ID NO:214)
forward PCR primer 2 5'-TTCCCTTGTGGGTTGGAG-3' (SEQ ID NO:215)
forward PCR primer 3 5'-AGGGCTGGAAGCCAGTTC-3' (SEQ ID NO:216)
reverse PCR primer 1 5'-AGCCAGTGAGGAAATGCG-3' (SEQ ID NO:217)
reverse PCR primer 2 5'-TGTCCAAAGTACACACACCTGAGG-3' (SEQ ID NO:218)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35737 sequence which had the following nucleotide sequence

hybridization probe

5'-GATGCCACGATCGCCAAGGTGGGACAGCTCTTTGCCGCCTGGAAG-3' (SEQ ID NO:219)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO271 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO271 [herein designated as DNA39423-1182] (SEQ ID NO:212) and the derived protein sequence for PRO271.

The entire nucleotide sequence of DNA39423-1182 is shown in Figure 77 (SEQ ID NO:212). Clone DNA39423-1182 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 101-103 and ending at the stop codon at nucleotide positions 1181-1183 (Figure 77). The predicted polypeptide precursor is 360 amino acids long (Figure 78). Clone DNA39423-1182 has been deposited with

ATCC and is assigned ATCC deposit no. ATCC 209387.

Analysis of the amino acid sequence of the full-length PRO271 polypeptide suggests that it possess significant homology to the proteoglycan link protein, thereby indicating that PRO271 may be a link protein homolog.

EXAMPLE 36: Isolation of cDNA Clones Encoding Human PRO272

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA36460. Based on the DNA36460 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO272.

Forward and reverse PCR primers were synthesized:

forward PCR primer (.f1)	5'-CGCAGGCCCTCATGGCCAGG-3'	(SEQ ID NO:222)
forward PCR primer (.f2)	5'-GAAATCCTGGGTAATTGG-3'	(SEQ ID NO:223)
reverse PCR primer	5'-GTGCGCGGTGCTCACAGCTCATC-3'	(SEQ ID NO:224)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA36460 sequence which had the following nucleotide sequence

hybridization probe

5'-CCCCCCTGAGCGACGCTCCCCCATGATGACGCCCACGGGAACTTC-3' (SEQ ID NO:225)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO272 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO272 [herein designated as DNA40620-1183] (SEQ ID NO:220) and the derived protein sequence for PRO272.

The entire nucleotide sequence of DNA40620-1183 is shown in Figure 79 (SEQ ID NO:220). Clone DNA40620-1183 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 35-37 and ending at the stop codon at nucleotide positions 1019-1021 (Figure 79). The predicted polypeptide precursor is 328 amino acids long (Figure 80). Clone DNA40620-1183 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209388.

Analysis of the amino acid sequence of the full-length PRO272 polypeptide suggests that portions of it possess significant homology to the human and mouse reticulocalbin proteins, respectively, thereby indicating that PRO272 may be a novel reticulocalbin protein.

EXAMPLE 37: Isolation of cDNA Clones Encoding Human PRO294

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35731. Based on the DNA35731 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained

the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO294.

Forward and reverse PCR primers were synthesized:

forward PCR primer (.f1)		5'-TGGTCTCGCACACCGATC-3'	(SEQ ID NO:228)
forward PCR primer (.f2)		5'-CTGCTGTCCACAGGGGAG-3'	(SEQ ID NO:229)
forward PCR primer (.f3)		5'-CCTTGAAGCATACTGCTC-3'	(SEQ ID NO:230)
forward PCR primer (.f4)		5'-GAGATAGCAATTTCCGCC-3'	(SEQ ID NO:231)
reverse PCR primer	(.r1)	5'-TTCCTCAAGAGGGCAGCC-3'	(SEQ ID NO:232)
reverse PCR primer	(.r2)	5'-CTTGGCACCAATGTCCGAGATTTC-3'	

(SEQ ID NO:233)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35731 sequence which had the following nucleotide sequence

hybridization probe

5'-GCTCTGAGGAAGGTGACGCGCGGGGCCTCCGAACCCTTGGCCTTG-3'

(SEQ ID NO:234)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO294 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO294 [herein designated as DNA40604-1187] (SEQ ID NO:226) and the derived protein sequence for PRO294.

The entire nucleotide sequence of DNA40604-1187 is shown in Figure 81 (SEQ ID NO:226). Clone DNA40604-1187 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 396-398 and ending at the stop codon at nucleotide positions 2046-2048 (Figure 81). The predicted polypeptide precursor is 550 amino acids long (Figure 82). Clone DNA40604-1187 has been deposited with ATCC and is assigned ATCC deposit no. 209394.

Analysis of the amino acid sequence of the full-length PRO294 polypeptide suggests that portions of it possess significant homology to portions of various collagen proteins, thereby indicating that PRO294 may be collagen-like molecule.

EXAMPLE 38: Isolation of cDNA Clones Encoding Human PRO295

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35814. Based on the DNA35814 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO295.

Forward and reverse PCR primers were synthesized:

forward PCR primer (.f1) 5'-GCAGAGCGGAGATGCAGCGGCTTG-3'

(SEQ ID NO:238)

forward PCR primer (.f2) 5'-CCCAGCATGTACTGCCAG-3' (SEQ ID NO:239)

forward PCR primer (.f3) 5'-TTGGCAGCTTCATGGAGG-3' (SEQ ID NO:240)

forward PCR primer (.f4) 5'-CCTGGGCAAAAATGCAAC-3' (SEQ ID NO:241)

reverse PCR primer (.r1) 5'-CTCCAGCTCCTGGCGCACCTCCTC-3' (SEQ ID NO:242)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35814 sequence which had the following nucleotide sequence

hybridization probe

5'-GGCTCTCAGCTACCGCGCAGGAGCGAGGCCACCCTCAATGAGATG-3'

(SEQ ID NO:243)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO295 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO295 [herein designated as DNA38268-1188] (SEQ ID NO:235) and the derived protein sequence for PRO295.

The entire nucleotide sequence of DNA38268-1188 is shown in Figure 83 (SEQ ID NO:235). Clone DNA38268-1188 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 153-155 and ending at the stop codon at nucleotide positions 1202-1204 (Figure 83). The predicted polypeptide precursor is 350 amino acids long (Figure 84). Clone DNA38268-1188 has been deposited with ATCC and is assigned ATCC deposit no. 209421.

Analysis of the amino acid sequence of the full-length PRO295 polypeptide suggests that portions of it possess significant homology to the integrin proteins, thereby indicating that PRO295 may be a novel integrin.

EXAMPLE 39: Isolation of cDNA Clones Encoding Human PRO293

The extracellular domain (ECD) sequences (including the secretion signal, if any) of from about 950 known secreted proteins from the Swiss-Prot public protein database were used to search expressed sequence tag (EST) databases. The EST databases included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA). The search was performed using the computer program BLAST or BLAST2 (Altshul et al., Methods in Enzymology 266:460-480 (1996)) as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequence. Those comparisons resulting in a BLAST score of 70 (or in some cases 90) or greater that did not encode known proteins were clustered and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington; http://bozeman.mbt.washington.edu/phrap.docs/phrap.html).

Based on an expression tag sequence designated herein as T08294 identified in the above analysis, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO293.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-AACAAGGTAAGATGCCATCCTG-3' (SEQ ID NO:246)
reverse PCR primer 5'-AAACTTGTCGATGGAGACCAGCTC-3' (SEQ ID NO:247)
Additionally, a synthetic oligonucleotide hybridization probe was constructed from the expression sequence tag which had the following nucleotide sequence
hybridization probe
5'-AGGGGCTGCAAAGCCTGGAGAGCCTCTCCTTCTATGACAACCAGC-3' (SEQ ID NO:248)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO293 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO293 [herein designated as DNA37151-1193] (SEQ ID NO:244) and the derived protein sequence for PRO293.

The entire nucleotide sequence of DNA37151-1193 is shown in Figure 85 (SEQ ID NO:244). Clone DNA37151-1193 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 881-883 and ending at the stop codon after nucleotide position 3019 of SEQ ID NO:244, Figure 85). The predicted polypeptide precursor is 713 amino acids long (Figure 86). Clone DNA37151-1193 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209393.

Analysis of the amino acid sequence of the full-length PRO293 polypeptide suggests that portions of it possess significant homology to the NLRR proteins, thereby indicating that PRO293 may be a novel NLRR protein.

EXAMPLE 40: Isolation of cDNA Clones Encoding Human PRO247

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA33480. Based on the DNA33480 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO247.

A pair of PCR primers (forward and reverse) were synthesized:
forward PCR primer 5'-CAACAATGAGGGCACCAAGC-3' (SEQ ID NO:251)
reverse PCR primer 5'-GATGGCTAGGTTCTGGAGGTTCTG-3' (SEQ ID NO:252)
Additionally, a synthetic oligonucleotide hybridization probe was constructed from the DNA33480 expression sequence tag which had the following nucleotide sequence
hybridization probe
5'-CAACCTGCAGGAGATTGACCTCAAGGACAACAACCTCAAGACCATCG-3' (SEQ ID NO:253)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO247 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO247 [herein designated as DNA35673-1201] (SEQ ID NO:249) and the derived protein sequence for PRO247.

The entire nucleotide sequence of DNA35673-1201 is shown in Figure 89 (SEQ ID NO:249). Clone DNA35673-1201 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 80-82 of SEQ ID NO:249 and ending at the stop codon after nucleotide position 1717 of SEQ ID NO:249 (Figure 89). The predicted polypeptide precursor is 546 amino acids long (Figure 88). Clone DNA35673-1201 has been deposited with ATCC and is assigned ATCC deposit no. 209418.

Analysis of the amino acid sequence of the full-length PRO247 polypeptide suggests that portions of it possess significant homology to the densin molecule and KIAA0231, thereby indicating that PRO247 may be a novel leucine rich repeat protein.

EXAMPLE 41: Isolation of cDNA Clones Encoding Human PRO302, PRO303, PRO304, PRO307 and PRO343

Consensus DNA sequences were assembled relative to other EST sequences using phrap as described in Example 1 above. These consensus sequences are herein designated DNA35953, DNA35955, DNA35958, DNA37160 and DNA30895. Based on the DNA35953 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO302.

PCR primers (forward and reverse) were synthesized:

forward PCR primer 1 5'-GTCCGCAAGGATGCCTACATGTTC-3' (SEQ ID NO:264)

forward PCR primer 2 5'-GCAGAGGTGTCTAAGGTTG-3' (SEQ ID NO:265)

reverse PCR primer 5'-AGCTCTAGACCAATGCCAGCTTCC-3' (SEQ ID NO:266)

Also, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35953 sequence which had the following nucleotide sequence

hybridization probe

5'-GCCACCAACTCCTGCAAGAACTTCTCAGAACTGCCCCTGGTCATG-3' (SEQ ID NO:267)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO302 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue (LIB228).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO302 [herein designated as DNA40370-1217] (SEQ ID NO:254) and the derived protein sequence for PRO302.

The entire nucleotide sequence of DNA40370-1217 is shown in Figure 89 (SEQ ID NO:254). Clone DNA40370-1217 contains a single open reading frame with an apparent translational initiation site at nucleotide

positions 34-36 and ending at the stop codon at nucleotide positions 1390-1392 (Figure 89). The predicted polypeptide precursor is 452 amino acids long (Figure 90). Various unique aspects of the PRO302 protein are shown in Figure 90. Clone DNA40370-1217 has been deposited with the ATCC on November 21, 1997 and is assigned ATCC deposit no. ATCC 209485.

Based on the DNA35955 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO303.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-GGGGAATTCACCCTATGACATTGCC-3' (SEQ ID NO:268)

reverse PCR primer 5'-GAATGCCCTGCAAGCATCAACTGG-3' (SEQ ID NO:269)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35955 sequence which had the following nucleotide sequence:

hybridization probe

5'-GCACCTGTCACCTACACTAAACACATCCAGCCCATCTGTCTCCAGGCCTC-3' (SEQ ID NO:270)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO303 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue (LIB25).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO303 [herein designated as DNA42551-1217] (SEQ ID NO:256) and the derived protein sequence for PRO303.

The entire nucleotide sequence of DNA42551-1217 is shown in Figure 91 (SEQ ID NO:256). Clone DNA42551-1217 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 20-22 and ending at the stop codon at nucleotide positions 962-964 (Figure 91). The predicted polypeptide precursor is 314 amino acids long (Figure 92). Various unique aspects of the PRO303 protein are shown in Figure 92. Clone DNA42551-1217 has been deposited on November 21, 1997 with the ATCC and is assigned ATCC deposit no. ATCC 209483.

Based on the DNA35958 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO304.

Pairs of PCR primers (forward and reverse) were synthesized:

forward PCR primer 1 5'-GCGGAAGGGCAGAATGGGACTCCAAG-3' (SEQ ID NO:271)

forward PCR primer 2 5'-CAGCCCTGCCACATGTGC-3' (SEQ ID NO:272)

forward PCR primer 3 5'-TACTGGGTGGTCAGCAAC-3' (SEQ ID NO:273)

reverse PCR primer 5'-GGCGAAGAGCAGGGTGAGACCCCG-3' (SEQ ID NO:274)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35958 sequence which had the following nucleotide sequence

hybridization probe

5'-GCCCTCATCCTCTCTGGCAAATGCAGTTACAGCCCGGAGCCCGAC-3' (SEQ ID NO:275)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO304 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from 22 week human fetal brain tissue (LIB153).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO304 [herein designated as DNA39520-1217] (SEQ ID NO:258) and the derived protein sequence for PRO304.

The entire nucleotide sequence of DNA39520-1217 is shown in Figure 93 (SEQ ID NO:258). Clone DNA39520-1217 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 34-36 and ending at the stop codon at nucleotide positions 1702-1704 (Figure 93). The predicted polypeptide precursor is 556 amino acids long (Figure 94). Various unique aspects of the PRO304 protein are shown in Figure 94. Clone DNA39520-1217 has been deposited with ATCC on November 21, 1997 and is assigned ATCC deposit no. ATCC 209482.

Based on the DNA37160 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO307.

Pairs of PCR primers (forward and reverse) were synthesized:

forward PCR primer 1 5'-GGGCAGGGATTCCAGGGCTCC-3' (SEQ ID NO:276)

forward PCR primer 2 5'-GGCTATGACAGCAGGTTC-3' (SEQ ID NO:277)

forward PCR primer 3 5'-TGACAATGACCGACCAGG-3' (SEQ ID NO:278)

reverse PCR primer 5'-GCATCGCATTGCTGGTAGAGCAAG-3' (SEQ ID NO:279)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA37160 sequence which had the following nucleotide sequence

hybridization probe

5'-TTACAGTGCCCCCTGGAAACCCACTTGGCCTGCATACCGCCTCCC-3' (SEQ ID NO:280)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO307 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue (LIB229).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO307 [herein designated as DNA41225-1217] (SEQ ID NO:260) and the derived protein sequence for PRO307.

The entire nucleotide sequence of DNA41225-1217 is shown in Figure 95 (SEQ ID NO:260). Clone DNA41225-1217 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 92-94 and ending at the stop codon at nucleotide positions 1241-1243 (Figure 95). The predicted polypeptide precursor is 383 amino acids long (Figure 96). Various unique aspects of the PRO307 protein are shown in Figure 96. Clone DNA41225-1217 has been deposited with ATCC on November 21, 1997 and is assigned ATCC deposit no. ATCC 209491.

Based on the DNA30895 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO343.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CGTCTCGAGCGCTCCATACAGTTCCCTTGCCCCA-3' (SEQ ID NO:281)

reverse PCR primer

5'-TGGAGGGGGAGCGGGATGCTTGTCTGGGCGACTCCGGGGGCC

CCCTCATGTGCCAGGTGGA-3' (SEQ ID NO:282)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30895 sequence which had the following nucleotide sequence

hybridization probe

5'-CCCTCAGACCCTGCAGAAGCTGAAGGTTCCTATCATCGACTCGGAAGTCTGCAGCCATCTG

TACTGGCGGGGAGCAGGACAGGGACCCATCACTGAGGACATGCTGTGTGCCGGCTACT-3' (SEQ ID NO:283)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO343 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue (LIB26).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO343 [herein designated as DNA43318-1217] (SEQ ID NO:262) and the derived protein sequence for PRO343.

The entire nucleotide sequence of DNA43318-1217 is shown in Figure 97 (SEQ ID NO:262). Clone DNA43318-1217 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 53-55 and ending at the stop codon at nucleotide positions 1004-1006 (Figure 97). The predicted polypeptide precursor is 317 amino acids long (Figure 98). Various unique aspects of the PRO343 protein are shown in Figure 98. Clone DNA43318-1217 has been deposited with ATCC on November 21, 1997 and is assigned ATCC deposit no. ATCC 209481.

EXAMPLE 42: Isolation of cDNA Clones Encoding Human PRO328

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35615. Based on the DNA35615 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO328.

Forward and reverse PCR primers were synthesized:

forward PCR primer	5'-TCCTGCAGTTTCCTGATGC-3'	(SEQ ID NO:286)
reverse PCR primer	5'-CTCATATTGCACACCAGTAATTCG-3'	(SEQ ID NO:287)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35615 sequence which had the following nucleotide sequence

hybridization probe

5'-ATGAGGAGAAACGTTTGATGGTGGAGCTGCACAACCTCTACCGGG-3'

(SEQ ID NO:288)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO328 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO328 [herein designated as DNA40587-1231] (SEQ ID NO:284) and the derived protein sequence for PRO328.

The entire nucleotide sequence of DNA40587-1231 is shown in Figure 99 (SEQ ID NO:284). Clone DNA40587-1231 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 15-17 and ending at the stop codon at nucleotide positions 1404-1406 (Figure 99). The predicted polypeptide precursor is 463 amino acids long (Figure 100). Clone DNA40587-1231 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209438.

Analysis of the amino acid sequence of the full-length PRO328 polypeptide suggests that portions of it possess significant homology to the human glioblastoma protein and to the cysteine rich secretory protein thereby indicating that PRO328 may be a novel glioblastoma protein or cysteine rich secretory protein.

EXAMPLE 43: Isolation of cDNA Clones Encoding Human PRO335, PRO331 or PRO326

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA36685. Based on the DNA36685 consensus sequence, and Incyte EST sequence no. 2228990, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO335, PRO331 or PRO326.

Forward and reverse PCR primers were synthesized for the determination of PRO335:

forward PCR primer	5'-GGAACCGAATCTCAGCTA-3'	(SEQ ID NO:295)
forward PCR primer	5'-CCTAAACTGAACTGGACCA-3'	(SEQ ID NO:296)
forward PCR primer	5'-GGCTGGAGACACTGAACCT-3'	(SEQ ID NO:297)
forward PCR primer	5'-ACAGCTGCACAGCTCAGAACAGTG-3'	(SEQ ID NO:298)
reverse PCR primer	5'-CATTCCCAGTATAAAAATTTTC-3'	(SEQ ID NO:299)
reverse PCR primer	5'-GGGTCTTGGTGAATGAGG-3'	(SEQ ID NO:300)
reverse PCR primer	5'-GTGCCTCTCGGTTACCACCAATGG-3'	(SEQ ID NO:301)

Additionally, a synthetic oligonucleotide hybridization probe was constructed for the determination of PRO335 which had the following nucleotide sequence

hybridization probe

5'-GCGGCCACTGTTGGACCGAACTGTAACCAAGGGAGAAACAGCCGTCCTAC-3'

(SEQ ID NO:302)

Forward and reverse PCR primers were synthesized for the determination of PRO331:

forward PCR primer 5'-GCCTTTGACAACCTTCAGTCACTAGTGG-3' (SEQ ID NO:303)

reverse PCR primer 5'-CCCCATGTGTCCATGACTGTTCCC-3' (SEQ ID NO:304)

Additionally, a synthetic oligonucleotide hybridization probe was constructed for the determination of PRO331 which had the following nucleotide sequence

hybridization probe

5'-TACTGCCTCATGACCTCTTCACTCCCTTGCATCATCTTAGAGCGG-3'

(SEQ ID NO:305)

Forward and reverse PCR primers were synthesized for the determination of PRO326:

forward PCR primer 5'-ACTCCAAGGAAATCGGATCCGTTC-3' (SEQ ID NO:306)

reverse PCR primer 5'-TTAGCAGCTGAGGATGGGCACAAC-3' (SEQ ID NO:307)

Additionally, a synthetic oligonucleotide hybridization probe was constructed for the determination of PRO331 which had the following nucleotide sequence

hybridization probe

5'-GCCTTCACTGGTTTGGATGCATTGGAGCATCTAGACCTGAGTGACAACGC-3'

(SEQ ID NO:308)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO335, PRO331 or PRO326 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue (PRO335 and PRO326) and human fetal brain (PRO331).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO335, PRO331 or PRO326 [herein designated as SEQ ID NOS:289, 291 and 293, respectively; see Figures 101, 103 and 105, respectively], and the derived protein sequence for PRO335, PRO331 or PRO326 (see Figures 102, 104 and 106, respectively; SEQ ID NOS:290, 292 and 294, respectively).

The entire nucleotide sequences are shown in Figures 101, 103 and 105, deposited with the ATCC on June 2, 1998, November 7, 1997 and November 21, 1997, respectively.

Analysis of the amino acid sequence of the full-length PRO335, PRO331 or PRO326 polypeptide suggests that portions of it possess significant homology to the LIG-1 protein, thereby indicating that PRO335, PRO331 and PRO326 may be a novel LIG-1-related protein.

EXAMPLE 44: Isolation of cDNA clones Encoding Human PRO332

Based upon an ECD homology search performed as described in Example 1 above, a consensus DNA sequence designated herein as DNA36688 was assembled. Based on the DNA36688 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO332.

A pair of PCR primers (forward and reverse) were synthesized:

5'-GCATTGGCCGCGAGACTTTGCC-3' (SEQ ID NO:311)

5'-GCGGCCACGGTCCTTGGAAATG-3' (SEQ ID NO:312)

A probe was also synthesized:

5'-TGGAGGAGCTCAACCTCAGCTACAACCGCATCACCAGCCCACAGG-3'

(SEQ ID NO:313)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO332 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from a human fetal liver library (LIB229).

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for DNA40982-1235 and the derived protein sequence for PRO332.

The entire nucleotide sequence of DNA40982-1235 is shown in Figure 107 (SEQ ID NO:309). Clone DNA40982-1235 contains a single open reading frame (with an apparent translational initiation site at nucleotide positions 342-344, as indicated in Figure 107). The predicted polypeptide precursor is 642 amino acids long, and has a calculated molecular weight of 72,067 (pI: 6.60). Clone DNA40982-1235 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209433.

Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO332 shows about 30-40% amino acid sequence identity with a series of known proteoglycan sequences, including, for example, fibromodulin and fibromodulin precursor sequences of various species (FMOD_BOVIN, FMOD CHICK, FMOD_RAT, FMOD_MOUSE, FMOD_HUMAN, P_R36773), osteomodulin sequences (AB000114 1, AB007848_1), decorin sequences (CFU83141_1, OCU03394_1, P_R42266, P_R42267, P_R42260, P_R89439), keratan sulfate proteoglycans (BTU48360_1, AF022890_1), corneal proteoglycan (AF022256_1), and bone/cartilage proteoglycans and proteoglycane precursors (PGS1_BOVIN, PGS2_ MOUSE, PGS2_HUMAN).

EXAMPLE 45: Isolation of cDNA clones Encoding Human PRO334

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. Based on the consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO334.

Forward and reverse PCR primers were synthesized for the determination of PRO334:

forward PCR primer 5'-GATGGTTCCTGCTCAAGTGCCCTG-3' (SEQ ID NO:316)

reverse PCR primer 5'-TTGCACTTGTAGGACCCACGTACG-3' (SEQ ID NO:317)

Additionally, a synthetic oligonucleotide hybridization probe was constructed for the determination of PRO334 which had the following nucleotide sequence

hybridization probe

5'-CTGATGGGAGGACCTGTGTAGATGTTGATGAATGTGCTACAGGAAGAGCC-3'

(SEQ ID NO:318)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was

screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO334 gene using the probe oligonucleotide and one of the PCR primers.

Human fetal kidney cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO334 [herein designated as DNA41379-1236] (SEQ ID NO:314) and the derived protein sequence for PRO334.

The entire nucleotide sequence of DNA41379-1236 (also referred to as UNQ295) is shown in Figure 109 (SEQ ID NO:314). Clone DNA41379-1236 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 203-205 and ending at the stop codon at nucleotide positions 1730-1732 (Figure 109). The predicted polypeptide precursor is 509 amino acids long (Figure 110). Clone DNA41379-1236 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209488.

Analysis of the amino acid sequence of the full-length PRO334 polypeptide suggests that portions of it possess significant homology to the fibulin and fibrillin proteins, thereby indicating that PRO334 may be a novel member of the EGF protein family.

EXAMPLE 46: Isolation of cDNA Clones Encoding Human PRO346

A consensus DNA sequence was identified using phrap as described in Example I above. Specifically, this consensus sequence is herein designated DNA38240. Based on the DNA38240 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length PRO346 coding sequence.

RNA for construction of the cDNA libraries was isolated from human fetal liver. The cDNA libraries used to isolated the cDNA clones were constructed by standard methods using commercially available reagents (e.g., Invitrogen, San Diego, CA; Clontech, etc.) The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes et al., *Science*, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

A cDNA clone was sequenced in entirety. The entire nucleotide sequence of DNA44167-1243 is shown in Figure 111 (SEQ ID NO:319). Clone DNA44167-1243 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 64-66 (Fig. 111; SEQ ID NO:319). The predicted polypeptide precursor is 450 amino acids long. Clone DNA44167-1243 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209434 (designation DNA44167-1243).

Based on a BLAST, BLAST-2 and FastA sequence alignment analysis (using the ALIGN computer program) of the full-length sequence, PRO346 shows amino acid sequence identity to carcinoembryonic antigen (28%).

The oligonucleotide sequences used in the above procedure were the following:

OLI2691 (38240.f1)

5'-GATCCTGTCACAAAGCCAGTGGTGC-3' (SEQ ID NO:321)

OLI2693 (38240.r1)

5'-CACTGACAGGGTTCCTCACCCAGG-3' (SEQ ID NO:322)

OLI2692 (38240.p1)

5'-CTCCCTCTGGGCTGTGGAGTATGTGGGGAACATGACCCTGACATG-3' (SEQ ID NO:323)

EXAMPLE 47: Isolation of cDNA Clones Encoding Human PRO268

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35698. Based on the DNA35698 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO268.

Forward and reverse PCR primers were synthesized:

forward PCR primer 1 5'-TGAGGTGGGCAAGCGGCGAAATG-3' (SEQ ID NO:326)

forward PCR primer 2 5'-TATGTGGATCAGGACGTGCC-3' (SEQ ID NO:327)

forward PCR primer 3 5'-TGCAGGGTTCAGTCTAGATTG-3' (SEQ ID NO:328)

reverse PCR primer 5'-TTGAAGGACAAAGGCAATCTGCCAC-3' (SEQ ID NO:329)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35698 sequence which had the following nucleotide sequence

hybridization probe

5'-GGAGTCTTGCAGTTCCCCTGGCAGTCCTGGTGCTGTTGCTTTGGG-3' (SEQ ID NO:330)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO268 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO268 [herein designated as DNA39427-1179] (SEQ ID NO:324) and the derived protein sequence for PRO268.

The entire nucleotide sequence of DNA39427-1179 is shown in Figure 113 (SEQ ID NO:324). Clone DNA39427-1179 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 13-15 and ending at the stop codon at nucleotide positions 853-855 (Figure 113). The predicted polypeptide precursor is 280 amino acids long (Figure 114). Clone DNA39427-1179 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209395.

Analysis of the amino acid sequence of the full-length PRO268 polypeptide suggests that it possess significant homology to protein disulfide isomerase, thereby indicating that PRO268 may be a novel protein disulfide isomerase.

EXAMPLE 48: Isolation of cDNA Clones Encoding Human PRO330

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA35730. Based on the DNA35730 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO330.

Forward and reverse PCR primers were synthesized:

forward PCR primer 1 5'-CCAGGCACAATTTCCAGA-3' (SEQ ID NO:333)
forward PCR primer 2 5'-GGACCCTTCTGTGTGCCAG-3' (SEQ ID NO:334)
reverse PCR primer 1 5'-GGTCTCAAGAACTCCTGTC-3' (SEQ ID NO:335)
reverse PCR primer 2 5'-ACACTCAGCATTGCCTGGTACTTG-3' (SEQ ID NO:336)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus sequence which had the following nucleotide sequence

hybridization probe
5'-GGGCACATGACTGACCTGATTTATGCAGAGAAAGAGCTGGTGCAG-3' (SEQ ID NO:337)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO330 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO330 [herein designated as DNA40603-1232] (SEQ ID NO:331) and the derived protein sequence for PRO330.

The entire nucleotide sequence of DNA40603-1232 is shown in Figure 115 (SEQ ID NO:331). Clone DNA40603-1232 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 167-169 and ending at the stop codon at nucleotide positions 1766-1768 (Figure 115). The predicted polypeptide precursor is 533 amino acids long (Figure 116). Clone DNA40603-1232 has been deposited with ATCC and is assigned ATCC deposit no.ATCC 209486 on November 21, 1997.

Analysis of the amino acid sequence of the full-length PRO330 polypeptide suggests that portions of it possess significant homology to the mouse prolyl 4-hydroxylase alpha subunit protein, thereby indicating that PRO330 may be a novel prolyl 4-hydroxylase alpha subunit polypeptide.

EXAMPLE 49: Isolation of cDNA Clones Encoding Human PRO310

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA40553. Based on the DNA40553 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO310.

Forward and reverse PCR primers were synthesized:

forward PCR primer 1 5'-TCCCCAAGCCGTTCTAGACGCGG-3' (SEQ ID NO:342)

forward PCR primer 2 5'-CTGGTTCTTCCTTGCACG-3' (SEQ ID NO:343)

reverse PCR primer 5'-GCCCAAATGCCCTAAGGCGGTATACCCC-3' (SEQ ID NO:344)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus sequence which had the following nucleotide sequence

hybridization probe

5'-GGGTGTGATGCTTGGAAGCATTTTCTGTGCTTTGATCACTATGCTAGGAC-3' (SEQ ID NO:345)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO310 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO310 [herein designated as DNA43046-1225 (SEQ ID NO:340) and the derived protein sequence for PRO310 (SEQ ID NO:341).

The entire nucleotide sequence of DNA43046-1225 is shown in Figure 119 (SEQ ID NO:340). Clone DNA43046-1225 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 81-83 and ending at the stop codon at nucleotide positions 1035-1037 (Figure 119). The predicted polypeptide precursor is 318 amino acids long (Figure 120) and has a calculated molecular weight of approximately 36,382 daltons. Clone DNA43046-1225 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209484.

Analysis of the amino acid sequence of the full-length PRO310 polypeptide suggests that portions of it possess homology to *C. elegans* proteins and to fringe, thereby indicating that PRO310 may be involved in development.

EXAMPLE 50: Isolation of cDNA clones Encoding Human PRO339

An expressed sequence tag (EST) DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) was searched and ESTs were identified. An assembly of Incyte clones and a consensus sequence was formed using phrap as described in Example 1 above.

Forward and reverse PCR primers were synthesized based upon the assembly-created consensus sequence:

forward PCR primer 1 5'-GGGATGCAGGTGGTGTCTCATGGGG-3' (SEQ ID NO:346)

forward PCR primer 2 5'-CCCTCATGTACCGGCTCC-3' (SEQ ID NO:347)

forward PCR primer 3 5'-GTGTGACACAGCGTGGGC-3' (SEQ ID NO:43)

forward PCR primer 4 5'-GACCGGCAGGCTTCTGCG-3' (SEQ ID NO:44)

reverse PCR primer 1 5'-CAGCAGCTTCAGCCACCAGGAGTGG-3' (SEQ ID NO:45)

reverse PCR primer 2 5'-CTGAGCCGTGGGCTGCAGTCTCGC-3' (SEQ ID NO:46)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus sequence which had the following nucleotide sequence

hybridization probe

5'-CCGACTACGACTGGTTCTTCATCATGCAGGATGACACATATGTGC-3' (SEQ ID NO:47)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO339 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue.

A cDNA clone was sequenced in entirety. The entire nucleotide sequence of DNA43466-1225 is shown in Figure 117 (SEQ ID NO:338). Clone DNA43466-1225 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 333-335 and ending at the stop codon found at nucleotide positions 2649-2651 (Figure 117; SEQ ID NO:338). The predicted polypeptide precursor is 772 amino acids long and has a calculated molecular weight of approximately 86,226 daltons. Clone DNA43466-1225 has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209490.

Based on a BLAST and FastA sequence alignment analysis (using the ALIGN computer program) of the full-length sequence, PRO339 has homology to *C. elegans* proteins and collagen-like polymer sequences as well as to fringe, thereby indicating that PRO339 may be involved in development or tissue growth.

EXAMPLE 51: Isolation of cDNA Clones Encoding Human PRO244

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. Based on this consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO244.

A pair of PCR primers (forward and reverse) were synthesized:

5'-TTCAGCTTCTGGGATGTAGGG-3' (30923.f1) (SEQ ID NO:378)

5'-TATTCCTACCATTTCACAAATCCG-3' (30923.r1) (SEQ ID NO:379)

A probe was also synthesized:

5'-GGAGGACTGTGCCACCATGAGAGACTCTTCAAACCCAAGGCAAAATTGG-3' (30923.p1) (SEQ ID NO:380)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO244 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from a human fetal kidney library. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence and the derived protein sequence for PRO244.

The entire nucleotide sequence of PRO244 is shown in Figure 121 (SEQ ID NO:376). Clone DNA35668-1171 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 106-108 (Fig. 121). The predicted polypeptide precursor is 219 amino acids long. Clone DNA35668-1171 has been deposited with ATCC (designated as DNA35663-1171) and is assigned ATCC deposit no. ATCC209371. The protein has a cytoplasmic domain (aa 1-20), a transmembrane domain (aa 21-46), and an extracellular domain (aa 47-219), with a C-lectin domain at aa 55-206.

Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO244 shows

notable amino acid sequence identity to hepatic lectin gallus gallus (43%), HIC hp120-binding C-type lectin (42%), macrophage lectin 2 (HUMHML2-1, 41%), and sequence PR32188 (44%).

EXAMPLE 52: Use of PRO Polypeptide-Encoding Nucleic Acid as Hybridization Probes

The following method describes use of a nucleotide sequence encoding a PRO polypeptide as a hybridization probe.

DNA comprising the coding sequence of of a PRO polypeptide of interest as disclosed herein may be employed as a probe or used as a basis from which to prepare probes to screen for homologous DNAs (such as those encoding naturally-occurring variants of the PRO polypeptide) in human tissue cDNA libraries or human tissue genomic libraries.

Hybridization and washing of filters containing either library DNAs is performed under the following high stringency conditions. Hybridization of radiolabeled PRO polypeptide-encoding nucleic acid-derived probe to the filters is performed in a solution of 50% formamide, 5x SSC, 0.1% SDS, 0.1% sodium pyrophosphate, 50 mM sodium phosphate, pH 6.8, 2x Denhardt's solution, and 10% dextran sulfate at 42°C for 20 hours. Washing of the filters is performed in an aqueous solution of 0.1x SSC and 0.1% SDS at 42°C.

DNAs having a desired sequence identity with the DNA encoding full-length native sequence PRO polypeptide can then be identified using standard techniques known in the art.

EXAMPLE 53: Expression of PRO Polypeptides in *E. coli*

This example illustrates preparation of an unglycosylated form of a desired PRO polypeptide by recombinant expression in *E. coli*.

The DNA sequence encoding the desired PRO polypeptide is initially amplified using selected PCR primers. The primers should contain restriction enzyme sites which correspond to the restriction enzyme sites on the selected expression vector. A variety of expression vectors may be employed. An example of a suitable vector is pBR322 (derived from *E. coli*; see Bolivar et al., Gene, 2:95 (1977)) which contains genes for ampicillin and tetracycline resistance. The vector is digested with restriction enzyme and dephosphorylated. The PCR amplified sequences are then ligated into the vector. The vector will preferably include sequences which encode for an antibiotic resistance gene, a trp promoter, a polyhis leader (including the first six STII codons, polyhis sequence, and enterokinase cleavage site), the specific PRO polypeptide coding region, lambda transcriptional terminator, and an argU gene.

The ligation mixture is then used to transform a selected *E. coli* strain using the methods described in Sambrook et al., supra. Transformants are identified by their ability to grow on LB plates and antibiotic resistant colonies are then selected. Plasmid DNA can be isolated and confirmed by restriction analysis and DNA sequencing.

Selected clones can be grown overnight in liquid culture medium such as LB broth supplemented with antibiotics. The overnight culture may subsequently be used to inoculate a larger scale culture. The cells are then grown to a desired optical density, during which the expression promoter is turned on.

After culturing the cells for several more hours, the cells can be harvested by centrifugation. The cell

pellet obtained by the centrifugation can be solubilized using various agents known in the art, and the solubilized PRO polypeptide can then be purified using a metal chelating column under conditions that allow tight binding of the protein.

PRO187, PRO317, PRO301, PRO224 and PRO238 were successfully expressed in *E. coli* in a poly-His tagged form, using the following procedure. The DNA encoding PRO187, PRO317, PRO301, PRO224 or PRO238 was initially amplified using selected PCR primers. The primers contained restriction enzyme sites which correspond to the restriction enzyme sites on the selected expression vector, and other useful sequences providing for efficient and reliable translation initiation, rapid purification on a metal chelation column, and proteolytic removal with enterokinase. The PCR-amplified, poly-His tagged sequences were then ligated into an expression vector, which was used to transform an *E. coli* host based on strain 52 (W3110 fuhA(tonA) lon galE rpoHts(htpRts) clpP(lacIq). Transformants were first grown in LB containing 50 mg/ml carbenicillin at 30°C with shaking until an O.D.600 of 3-5 was reached. Cultures were then diluted 50-100 fold into CRAP media (prepared by mixing 3.57 g $(NH_4)_2SO_4$, 0.71 g sodium citrate·2H2O, 1.07 g KCl, 5.36 g Difco yeast extract, 5.36 g Sheffield hycase SF in 500 mL water, as well as 110 mM MPOS, pH 7.3, 0.55% (w/v) glucose and 7 mM $MgSO_4$) and grown for approximately 20-30 hours at 30°C with shaking. Samples were removed to verify expression by SDS-PAGE analysis, and the bulk culture is centrifuged to pellet the cells. Cell pellets were frozen until purification and refolding.

E. coli paste from 0.5 to 1 L fermentations (6-10 g pellets) was resuspended in 10 volumes (w/v) in 7 M guanidine, 20 mM Tris, pH 8 buffer. Solid sodium sulfite and sodium tetrathionate is added to make final concentrations of 0.1M and 0.02 M, respectively, and the solution was stirred overnight at 4°C. This step results in a denatured protein with all cysteine residues blocked by sulfitolization. The solution was centrifuged at 40,000 rpm in a Beckman Ultracentifuge for 30 min. The supernatant was diluted with 3-5 volumes of metal chelate column buffer (6 M guanidine, 20 mM Tris, pH 7.4) and filtered through 0.22 micron filters to clarify. Depending the clarified extract was loaded onto a 5 ml Qiagen Ni-NTA metal chelate column equilibrated in the metal chelate column buffer. The column was washed with additional buffer containing 50 mM imidazole (Calbiochem, Utrol grade), pH 7.4. The protein was eluted with buffer containing 250 mM imidazole. Fractions containing the desired protein were pooled and stored at 4°C. Protein concentration was estimated by its absorbance at 280 nm using the calculated extinction coefficient based on its amino acid sequence.

The proteins were refolded by diluting sample slowly into freshly prepared refolding buffer consisting of: 20 mM Tris, pH 8.6, 0.3 M NaCl, 2.5 M urea, 5 mM cysteine, 20 mM glycine and 1 mM EDTA. Refolding volumes were chosen so that the final protein concentration was between 50 to 100 micrograms/ml. The refolding solution was stirred gently at 4°C for 12-36 hours. The refolding reaction was quenched by the addition of TFA to a final concentration of 0.4% (pH of approximately 3). Before further purification of the protein, the solution was filtered through a 0.22 micron filter and acetonitrile was added to 2-10% final concentration. The refolded protein was chromatographed on a Poros R1/H reversed phase column using a mobile buffer of 0.1% TFA with elution with a gradient of acetonitrile from 10 to 80%. Aliquots of fractions with A280 absorbance were analyzed on SDS polyacrylamide gels and fractions containing homogeneous refolded protein were pooled. Generally, the properly refolded species of most proteins are eluted at the lowest concentrations of acetonitrile since those species are the most compact with their hydrophobic interiors shielded

from interaction with the reversed phase resin. Aggregated species are usually eluted at higher acetonitrile concentrations. In addition to resolving misfolded forms of proteins from the desired form, the reversed phase step also removes endotoxin from the samples.

Fractions containing the desired folded PRO187, PRO317, PRO301, PRO224 and PRO238 proteins, respectively, were pooled and the acetonitrile removed using a gentle stream of nitrogen directed at the solution. Proteins were formulated into 20 mM Hepes, pH 6.8 with 0.14 M sodium chloride and 4% mannitol by dialysis or by gel filtration using G25 Superfine (Pharmacia) resins equilibrated in the formulation buffer and sterile filtered.

EXAMPLE 54: Expression of PRO Polypeptides in Mammalian Cells

This example illustrates preparation of a glycosylated form of a desired PRO polypeptide by recombinant expression in mammalian cells.

The vector, pRK5 (see EP 307,247, published March 15, 1989), is employed as the expression vector. Optionally, the PRO polypeptide-encoding DNA is ligated into pRK5 with selected restriction enzymes to allow insertion of the PRO polypeptide DNA using ligation methods such as described in Sambrook et al., supra. The resulting vector is called pRK5-PRO polypeptide.

In one embodiment, the selected host cells may be 293 cells. Human 293 cells (ATCC CCL 1573) are grown to confluence in tissue culture plates in medium such as DMEM supplemented with fetal calf serum and optionally, nutrient components and/or antibiotics. About 10 μg pRK5-PRO polypeptide DNA is mixed with about 1 μg DNA encoding the VA RNA gene [Thimmappaya et al., Cell, 31:543 (1982)] and dissolved in 500 μl of 1 mM Tris-HCl, 0.1 mM EDTA, 0.227 M $CaCl_2$. To this mixture is added, dropwise, 500 μl of 50 mM HEPES (pH 7.35), 280 mM NaCl, 1.5 mM $NaPO_4$, and a precipitate is allowed to form for 10 minutes at 25°C. The precipitate is suspended and added to the 293 cells and allowed to settle for about four hours at 37°C. The culture medium is aspirated off and 2 ml of 20% glycerol in PBS is added for 30 seconds. The 293 cells are then washed with serum free medium, fresh medium is added and the cells are incubated for about 5 days.

Approximately 24 hours after the transfections, the culture medium is removed and replaced with culture medium (alone) or culture medium containing 200 μCi/ml ^{35}S-cysteine and 200 μCi/ml ^{35}S-methionine. After a 12 hour incubation, the conditioned medium is collected, concentrated on a spin filter, and loaded onto a 15% SDS gel. The processed gel may be dried and exposed to film for a selected period of time to reveal the presence of PRO polypeptide. The cultures containing transfected cells may undergo further incubation (in serum free medium) and the medium is tested in selected bioassays.

In an alternative technique, PRO polypeptide may be introduced into 293 cells transiently using the dextran sulfate method described by Somparyrac et al., Proc. Natl. Acad. Sci., 12:7575 (1981). 293 cells are grown to maximal density in a spinner flask and 700 μg pRK5-PRO polypeptide DNA is added. The cells are first concentrated from the spinner flask by centrifugation and washed with PBS. The DNA-dextran precipitate is incubated on the cell pellet for four hours. The cells are treated with 20% glycerol for 90 seconds, washed with tissue culture medium, and re-introduced into the spinner flask containing tissue culture medium, 5 μg/ml bovine insulin and 0.1 μg/ml bovine transferrin. After about four days, the conditioned media is centrifuged and filtered to remove cells and debris. The sample containing expressed PRO polypeptide can then be

concentrated and purified by any selected method, such as dialysis and/or column chromatography.

In another embodiment, PRO polypeptides can be expressed in CHO cells. The pRK5-PRO polypeptide can be transfected into CHO cells using known reagents such as $CaPO_4$ or DEAE-dextran. As described above, the cell cultures can be incubated, and the medium replaced with culture medium (alone) or medium containing a radiolabel such as ^{35}S-methionine. After determining the presence of PRO polypeptide, the culture medium may be replaced with serum free medium. Preferably, the cultures are incubated for about 6 days, and then the conditioned medium is harvested. The medium containing the expressed PRO polypeptide can then be concentrated and purified by any selected method.

Epitope-tagged PRO polypeptide may also be expressed in host CHO cells. The PRO polypeptide may be subcloned out of the pRK5 vector. The subclone insert can undergo PCR to fuse in frame with a selected epitope tag such as a poly-his tag into a Baculovirus expression vector. The poly-his tagged PRO polypeptide insert can then be subcloned into a SV40 driven vector containing a selection marker such as DHFR for selection of stable clones. Finally, the CHO cells can be transfected (as described above) with the SV40 driven vector. Labeling may be performed, as described above, to verify expression. The culture medium containing the expressed poly-His tagged PRO polypeptide can then be concentrated and purified by any selected method, such as by Ni^{2+}-chelate affinity chromatography.

PRO211, PRO217, PRO230, PRO219, PRO245, PRO221, PRO258, PRO301, PRO224, PRO222, PRO234, PRO229, PRO223, PRO328 and PRO332 were successfully expressed in CHO cells by both a transient and a stable expression procedure. In addition, PRO232, PRO265, PRO246, PRO228, PRO227, PRO220, PRO266, PRO269, PRO287, PRO214, PRO231, PRO233, PRO238, PRO244, PRO235, PRO236, PRO262, PRO239, PRO257, PRO260, PRO263, PRO270, PRO271, PRO272, PRO294, PRO295, PRO293, PRO247, PRO303 and PRO268 were successfully transiently expressed in CHO cells.

Stable expression in CHO cells was performed using the following procedure. The proteins were expressed as an IgG construct (immunoadhesin), in which the coding sequences for the soluble forms (e.g. extracellular domains) of the respective proteins were fused to an IgG1 constant region sequence containing the hinge, CH2 and CH2 domains and/or is a poly-His tagged form.

Following PCR amplification, the respective DNAs were subcloned in a CHO expression vector using standard techniques as described in Ausubel et al., *Current Protocols of Molecular Biology*, Unit 3.16, John Wiley and Sons (1997). CHO expression vectors are constructed to have compatible restriction sites 5' and 3' of the DNA of interest to allow the convenient shuttling of cDNA's. The vector used expression in CHO cells is as described in Lucas *et al., Nucl. Acids Res.* 24: 9 (1774-1779 (1996), and uses the SV40 early promoter/enhancer to drive expression of the cDNA of interest and dihydrofolate reductase (DHFR). DHFR expression permits selection for stable maintenance of the plasmid following transfection.

Twelve micrograms of the desired plasmid DNA were introduced into approximately 10 million CHO cells using commercially available transfection reagents Superfect® (Quiagen), Dosper® or Fugene® (Boehringer Mannheim). The cells were grown and described in Lucas *et al.*, supra. Approximately 3×10^{-7} cells are frozen in an ampule for further growth and production as described below.

The ampules containing the plasmid DNA were thawed by placement into water bath and mixed by

vortexing. The contents were pipetted into a centrifuge tube containing 10 mLs of media and centrifuged at 1000 rpm for 5 minutes. The supernatant was aspirated and the cells were resuspended in 10 mL of selective media (0.2 μm filtered PS20 with 5% 0.2 μm diafiltered fetal bovine serum). The cells were then aliquoted into a 100 mL spinner containing 90 mL of selective media. After 1-2 days, the cells were transferred into a 250 mL spinner filled with 150 mL selective growth medium and incubated at 37°C. After another 2-3 days, a 250 mL, 500 mL and 2000 mL spinners were seeded with 3 x 10^5 cells/mL. The cell media was exchanged with fresh media by centrifugation and resuspension in production medium. Although any suitable CHO media may be employed, a production medium described in US Patent No. 5,122,469, issued June 16, 1992 was actually used. 3L production spinner is seeded at 1.2 x 10^6 cells/mL. On day 0, the cell number pH were determined. On day 1, the spinner was sampled and sparging with filtered air was commenced. On day 2, the spinner was sampled, the temperature shifted to 33°C, and 30 mL of 500 g/L glucose and 0.6 mL of 10% antifoam (e.g., 35% polydimethylsiloxane emulsion, Dow Corning 365 Medical Grade Emulsion). Throughout the production, pH was adjusted as necessary to keep at around 7.2. After 10 days, or until viability dropped below 70%, the cell culture was harvested by centrifugtion and filtering through a 0.22 μm filter. The filtrate was either stored at 4°C or immediately loaded onto columns for purification.

For the poly-His tagged constructs, the proteins were purified using a Ni-NTA column (Qiagen). Before purification, imidazole was added to the conditioned media to a concentration of 5 mM. The conditioned media was pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs of were purified from the conditioned media as follows. The conditioned medium was pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 μL of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity was assessed by SDS polyacrylamide gels and by N-terminal amino acid sequencing by Edman degradation.

PRO211, PRO217, PRO230, PRO232, PRO187, PRO265, PRO219, PRO246, PRO228, PRO533, PRO245, PRO221, PRO227, PRO220, PRO258, PRO266, PRO269, PRO287, PRO214, PRO317, PRO301, PRO224, PRO222, PRO234, PRO231, PRO229, PRO233, PRO238, PRO223, PRO235, PRO236, PRO262, PRO239, PRO257, PRO260, PRO263, PRO270, PRO271, PRO272, PRO294, PRO295, PRO293, PRO247, PRO304, PRO302, PRO307, PRO303, PRO343, PRO328, PRO326, PRO331, PRO332, PRO334, PRO346, PRO268, PRO330, PRO310 and PRO339 were also successfully transiently expressed in COS cells.

EXAMPLE 55: Expression of PRO Polypeptides in Yeast

The following method describes recombinant expression of a desired PRO polypeptide in yeast.

First, yeast expression vectors are constructed for intracellular production or secretion of PRO polypeptides from the ADH2/GAPDH promoter. DNA encoding a desired PRO polypeptide, a selected signal peptide and the promoter is inserted into suitable restriction enzyme sites in the selected plasmid to direct intracellular expression of the PRO polypeptide. For secretion, DNA encoding the PRO polypeptide can be cloned into the selected plasmid, together with DNA encoding the ADH2/GAPDH promoter, the yeast alpha-factor secretory signal/leader sequence, and linker sequences (if needed) for expression of the PRO polypeptide.

Yeast cells, such as yeast strain AB110, can then be transformed with the expression plasmids described above and cultured in selected fermentation media. The transformed yeast supernatants can be analyzed by precipitation with 10% trichloroacetic acid and separation by SDS-PAGE, followed by staining of the gels with Coomassie Blue stain.

Recombinant PRO polypeptide can subsequently be isolated and purified by removing the yeast cells from the fermentation medium by centrifugation and then concentrating the medium using selected cartridge filters. The concentrate containing the PRO polypeptide may further be purified using selected column chromatography resins.

EXAMPLE 56: Expression of PRO Polypeptides in Baculovirus-Infected Insect Cells

The following method describes recombinant expression of PRO polypeptides in Baculovirus-infected insect cells.

The desired PRO polypeptide is fused upstream of an epitope tag contained with a baculovirus expression vector. Such epitope tags include poly-his tags and immunoglobulin tags (like Fc regions of IgG). A variety of plasmids may be employed, including plasmids derived from commercially available plasmids such as pVL1393 (Novagen). Briefly, the PRO polypeptide or the desired portion of the PRO polypeptide (such as the sequence encoding the extracellular domain of a transmembrane protein) is amplified by PCR with primers complementary to the 5' and 3' regions. The 5' primer may incorporate flanking (selected) restriction enzyme sites. The product is then digested with those selected restriction enzymes and subcloned into the expression vector.

Recombinant baculovirus is generated by co-transfecting the above plasmid and BaculoGold™ virus DNA (Pharmingen) into *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711) using lipofectin (commercially available from GIBCO-BRL). After 4-5 days of incubation at 28°C, the released viruses are harvested and used for further amplifications. Viral infection and protein expression is performed as described by O'Reilley et al., Baculovirus expression vectors: A laboratory Manual, Oxford: Oxford University Press (1994).

Expressed poly-his tagged PRO polypeptide can then be purified, for example, by Ni^{2+}-chelate affinity chromatography as follows. Extracts are prepared from recombinant virus-infected Sf9 cells as described by Rupert *et al.*, *Nature*, 362:175-179 (1993). Briefly, Sf9 cells are washed, resuspended in sonication buffer (25 mL Hepes, pH 7.9; 12.5 mM $MgCl_2$; 0.1 mM EDTA; 10% Glycerol; 0.1% NP-40; 0.4 M KCl), and sonicated twice for 20 seconds on ice. The sonicates are cleared by centrifugation, and the supernatant is diluted 50-fold in loading buffer (50 mM phosphate, 300 mM NaCl, 10% Glycerol, pH 7.8) and filtered through a 0.45 μm filter. A Ni^{2+}-NTA agarose column (commercially available from Qiagen) is prepared with a bed volume of 5 mL, washed with 25 mL of water and equilibrated with 25 mL of loading buffer. The filtered cell extract is

loaded onto the column at 0.5 mL per minute. The column is washed to baseline A_{280} with loading buffer, at which point fraction collection is started. Next, the column is washed with a secondary wash buffer (50 mM phosphate; 300 mM NaCl, 10% Glycerol, pH 6.0), which elutes nonspecifically bound protein. After reaching A_{280} baseline again, the column is developed with a 0 to 500 mM Imidazole gradient in the secondary wash buffer. One mL fractions are collected and analyzed by SDS-PAGE and silver staining or western blot with Ni^{2+}-NTA-conjugated to alkaline phosphatase (Qiagen). Fractions containing the eluted His_{10}-tagged PRO polypeptide are pooled and dialyzed against loading buffer.

Alternatively, purification of the IgG tagged (or Fc tagged) PRO polypeptide can be performed using known chromatography techniques, including for instance, Protein A or protein G column chromatography.

PRO211, PRO217, PRO230, PRO187, PRO265, PRO246, PRO228, PRO533, PRO245, PRO221, PRO220, PRO258, PRO266, PRO269, PRO287, PRO214, PRO301, PRO224, PRO222, PRO234, PRO231, PRO229, PRO235, PRO239, PRO257, PRO272, PRO294, PRO295, PRO328, PRO326, PRO331, PRO334, PRO346 and PRO310 were successfully expressed in baculovirus infected Sf9 or high5 insect cells. While the expression was actually performed in a 0.5-2 L scale, it can be readily scaled up for larger (e.g. 8 L) preparations. The proteins were expressed as an IgG construct (immunoadhesin), in which the protein extracellular region was fused to an IgG1 constant region sequence containing the hinge, CH2 and CH3 domains and/or in poly-His tagged forms.

Following PCR amplification, the respective coding sequences were subcloned into a baculovirus expression vector (pb.PH.IgG for IgG fusions and pb.PH.His.c for poly-His tagged proteins), and the vector and Baculogold® baculovirus DNA (Pharmingen) were co-transfected into 105 *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711), using Lipofectin (Gibco BRL). pb.PH.IgG and pb.PH.His are modifications of the commercially available baculovirus expression vector pVL1393 (Pharmingen), with modified polylinker regions to include the His or Fc tag sequences. The cells were grown in Hink's TNM-FH medium supplemented with 10% FBS (Hyclone). Cells were incubated for 5 days at 28°C. The supernatant was harvested and subsequently used for the first viral amplification by infecting Sf9 cells in Hink's TNM-FH medium supplemented with 10% FBS at an approximate multiplicity of infection (MOI) of 10. Cells were incubated for 3 days at 28°C. The supernatant was harvested and the expression of the constructs in the baculovirus expression vector was determined by batch binding of 1 ml of supernatant to 25 mL of Ni-NTA beads (QIAGEN) for histidine tagged proteins or Protein-A Sepharose CL-4B beads (Pharmacia) for IgG tagged proteins followed by SDS-PAGE analysis comparing to a known concentration of protein standard by Coomassie blue staining.

The first viral amplification supernatant was used to infect a spinner culture (500 ml) of Sf9 cells grown in ESF-921 medium (Expression Systems LLC) at an approximate MOI of 0.1. Cells were incubated for 3 days at 28°C. The supernatant was harvested and filtered. Batch binding and SDS-PAGE analysis was repeated, as necessary, until expression of the spinner culture was confirmed.

The conditioned medium from the transfected cells (0.5 to 3 L) was harvested by centrifugation to remove the cells and filtered through 0.22 micron filters. For the poly-His tagged constructs, the protein construct were purified using a Ni-NTA column (Qiagen). Before purification, imidazole was added to the conditioned media to a concentration of 5 mM. The conditioned media were pumped onto a 6 ml Ni-NTA

column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs of proteins were purified from the conditioned media as follows. The conditioned media were pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 mL of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity of the proteins was verified by SDS polyacrylamide gel (PEG) electrophoresis and N-terminal amino acid sequencing by Edman degradation.

EXAMPLE 57: Preparation of Antibodies that Bind to PRO Polypeptides

This example illustrates preparation of monoclonal antibodies which can specifically bind to a PRO polypeptide.

Techniques for producing the monoclonal antibodies are known in the art and are described, for instance, in Goding, supra. Immunogens that may be employed include purified PRO polypeptide, fusion proteins containing the PRO polypeptide, and cells expressing recombinant PRO polypeptide on the cell surface. Selection of the immunogen can be made by the skilled artisan without undue experimentation.

Mice, such as Balb/c, are immunized with the PRO polypeptide immunogen emulsified in complete Freund's adjuvant and injected subcutaneously or intraperitoneally in an amount from 1-100 micrograms. Alternatively, the immunogen is emulsified in MPL-TDM adjuvant (Ribi Immunochemical Research, Hamilton, MT) and injected into the animal's hind foot pads. The immunized mice are then boosted 10 to 12 days later with additional immunogen emulsified in the selected adjuvant. Thereafter, for several weeks, the mice may also be boosted with additional immunization injections. Serum samples may be periodically obtained from the mice by retro-orbital bleeding for testing in ELISA assays to detect anti-PRO polypeptide antibodies.

After a suitable antibody titer has been detected, the animals "positive" for antibodies can be injected with a final intravenous injection of PRO polypeptide. Three to four days later, the mice are sacrificed and the spleen cells are harvested. The spleen cells are then fused (using 35% polyethylene glycol) to a selected murine myeloma cell line such as P3X63AgU.1, available from ATCC, No. CRL 1597. The fusions generate hybridoma cells which can then be plated in 96 well tissue culture plates containing HAT (hypoxanthine, aminopterin, and thymidine) medium to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

The hybridoma cells will be screened in an ELISA for reactivity against the PRO polypeptide. Determination of "positive" hybridoma cells secreting the desired monoclonal antibodies against the PRO polypeptide is within the skill in the art.

The positive hybridoma cells can be injected intraperitoneally into syngeneic Balb/c mice to produce

ascites containing the anti-PRO polypeptide monoclonal antibodies. Alternatively, the hybridoma cells can be grown in tissue culture flasks or roller bottles. Purification of the monoclonal antibodies produced in the ascites can be accomplished using ammonium sulfate precipitation, followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can be employed.

EXAMPLE 58: Chimeric PRO Polypeptides

PRO polypeptides may be expressed as chimeric proteins with one or more additional polypeptide domains added to facilitate protein purification. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS™ extension/affinity purification system (Immunex Corp., Seattle Wash.). The inclusion of a cleavable linker sequence such as Factor XA or enterokinase (Invitrogen, San Diego Calif.) between the purification domain and the PRO polypeptide sequence may be useful to facilitate expression of DNA encoding the PRO polypeptide.

EXAMPLE 59: Purification of PRO Polypeptides Using Specific Antibodies

Native or recombinant PRO polypeptides may be purified by a variety of standard techniques in the art of protein purification. For example, pro-PRO polypeptide, mature PRO polypeptide, or pre-PRO polypeptide is purified by immunoaffinity chromatography using antibodies specific for the PRO polypeptide of interest. In general, an immunoaffinity column is constructed by covalently coupling the anti-PRO polypeptide antibody to an activated chromatographic resin.

Polyclonal immunoglobulins are prepared from immune sera either by precipitation with ammonium sulfate or by purification on immobilized Protein A (Pharmacia LKB Biotechnology, Piscataway, N.J.). Likewise, monoclonal antibodies are prepared from mouse ascites fluid by ammonium sulfate precipitation or chromatography on immobilized Protein A. Partially purified immunoglobulin is covalently attached to a chromatographic resin such as CnBr-activated SEPHAROSE™ (Pharmacia LKB Biotechnology). The antibody is coupled to the resin, the resin is blocked, and the derivative resin is washed according to the manufacturer's instructions.

Such an immunoaffinity column is utilized in the purification of PRO polypeptide by preparing a fraction from cells containing PRO polypeptide in a soluble form. This preparation is derived by solubilization of the whole cell or of a subcellular fraction obtained via differential centrifugation by the addition of detergent or by other methods well known in the art. Alternatively, soluble PRO polypeptide containing a signal sequence may be secreted in useful quantity into the medium in which the cells are grown.

A soluble PRO polypeptide-containing preparation is passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of PRO polypeptide (*e.g.*, high ionic strength buffers in the presence of detergent). Then, the column is eluted under conditions that disrupt antibody/PRO polypeptide binding (*e.g.*, a low pH buffer such as approximately pH 2-3, or a high concentration of a chaotrope such as urea or thiocyanate ion), and PRO polypeptide is collected.

EXAMPLE 60: Drug Screening

This invention is particularly useful for screening compounds by using PRO polypeptides or binding fragment thereof in any of a variety of drug screening techniques. The PRO polypeptide or fragment employed in such a test may either be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. One method of drug screening utilizes eukaryotic or prokaryotic host cells which are stably transformed with recombinant nucleic acids expressing the PRO polypeptide or fragment. Drugs are screened against such transformed cells in competitive binding assays. Such cells, either in viable or fixed form, can be used for standard binding assays. One may measure, for example, the formation of complexes between PRO polypeptide or a fragment and the agent being tested. Alternatively, one can examine the diminution in complex formation between the PRO polypeptide and its target cell or target receptors caused by the agent being tested.

Thus, the present invention provides methods of screening for drugs or any other agents which can affect a PRO polypeptide-associated disease or disorder. These methods comprise contacting such an agent with an PRO polypeptide or fragment thereof and assaying (I) for the presence of a complex between the agent and the PRO polypeptide or fragment, or (ii) for the presence of a complex between the PRO polypeptide or fragment and the cell, by methods well known in the art. In such competitive binding assays, the PRO polypeptide or fragment is typically labeled. After suitable incubation, free PRO polypeptide or fragment is separated from that present in bound form, and the amount of free or uncomplexed label is a measure of the ability of the particular agent to bind to PRO polypeptide or to interfere with the PRO polypeptide/cell complex.

Another technique for drug screening provides high throughput screening for compounds having suitable binding affinity to a polypeptide and is described in detail in WO 84/03564, published on September 13, 1984. Briefly stated, large numbers of different small peptide test compounds are synthesized on a solid substrate, such as plastic pins or some other surface. As applied to a PRO polypeptide, the peptide test compounds are reacted with PRO polypeptide and washed. Bound PRO polypeptide is detected by methods well known in the art. Purified PRO polypeptide can also be coated directly onto plates for use in the aforementioned drug screening techniques. In addition, non-neutralizing antibodies can be used to capture the peptide and immobilize it on the solid support.

This invention also contemplates the use of competitive drug screening assays in which neutralizing antibodies capable of binding PRO polypeptide specifically compete with a test compound for binding to PRO polypeptide or fragments thereof. In this manner, the antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with PRO polypeptide.

EXAMPLE 61: Rational Drug Design

The goal of rational drug design is to produce structural analogs of biologically active polypeptide of interest (*i.e.*, a PRO polypeptide) or of small molecules with which they interact, *e.g.*, agonists, antagonists, or inhibitors. Any of these examples can be used to fashion drugs which are more active or stable forms of the PRO polypeptide or which enhance or interfere with the function of the PRO polypeptide *in vivo* (*c.f.*, Hodgson, Bio/Technology, 9: 19-21 (1991)).

In one approach, the three-dimensional structure of the PRO polypeptide, or of an PRO polypeptide-inhibitor complex, is determined by x-ray crystallography, by computer modeling or, most typically,

by a combination of the two approaches. Both the shape and charges of the PRO polypeptide must be ascertained to elucidate the structure and to determine active site(s) of the molecule. Less often, useful information regarding the structure of the PRO polypeptide may be gained by modeling based on the structure of homologous proteins. In both cases, relevant structural information is used to design analogous PRO polypeptide-like molecules or to identify efficient inhibitors. Useful examples of rational drug design may include molecules which have improved activity or stability as shown by Braxton and Wells, Biochemistry, 31:7796-7801 (1992) or which act as inhibitors, agonists, or antagonists of native peptides as shown by Athauda *et al.*, J. Biochem., 113:742-746 (1993).

It is also possible to isolate a target-specific antibody, selected by functional assay, as described above, and then to solve its crystal structure. This approach, in principle, yields a pharmacore upon which subsequent drug design can be based. It is possible to bypass protein crystallography altogether by generating anti-idiotypic antibodies (anti-ids) to a functional, pharmacologically active antibody. As a mirror image of a mirror image, the binding site of the anti-ids would be expected to be an analog of the original receptor. The anti-id could then be used to identify and isolate peptides from banks of chemically or biologically produced peptides. The isolated peptides would then act as the pharmacore.

By virtue of the present invention, sufficient amounts of the PRO polypeptide may be made available to perform such analytical studies as X-ray crystallography. In addition, knowledge of the PRO polypeptide amino acid sequence provided herein will provide guidance to those employing computer modeling techniques in place of or in addition to x-ray crystallography.

EXAMPLE 62: Diagnostic Test Using PRO317 Polypeptide-Specific Antibodies

Particular anti-PRO317 polypeptide antibodies are useful for the diagnosis of prepathologic conditions, and chronic or acute diseases such as gynecological diseases or ischemic diseases which are characterized by differences in the amount or distribution of PRO317. PRO317 has been found to be expressed in human kidney and is thus likely to be associated with abnormalities or pathologies which affect this organ. Further, since it is so closely related to EBAF-1, it is likely to affect the endometrium and other genital tissues. Further, due to library sources of certain ESTs, it appears that PRO317 may be involved as well in forming blood vessels and hence to be a modulator of angiogenesis.

Diagnostic tests for PRO317 include methods utilizing the antibody and a label to detect PRO317 in human body fluids, tissues, or extracts of such tissues. The polypeptide and antibodies of the present invention may be used with or without modification. Frequently, the polypeptide and antibodies will be labeled by joining them, either covalently or noncovalently, with a substance which provides for a detectable signal. A wide variety of labels and conjugation techniques are known and have been reported extensively in both the scientific and patent literature. Suitable labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent agents, chemiluminescent agents, magnetic particles, and the like. Patents teaching the use of such labels include U.S. Pat. Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345; 4,277,437; 4,275,149; and 4,366,241. Also, recombinant immunoglobulins may be produced as shown in U.S. Pat. No. 4,816,567.

A variety of protocols for measuring soluble or membrane-bound PRo317, using either polyclonal or monoclonal antibodies specific for that PRO317, are known in the art. Examples include enzyme-linked

immunosorbent assay (ELISA), radioimmunoassay (RIA), radioreceptor assay (RRA), and fluorescent activated cell sorting (FACS). A two-site monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on PRO317 is preferred, but a competitive binding assay may be employed. These assays are described, among other places, in Maddox *et al.* J Exp. Med., 158:1211 (1983).

EXAMPLE 63: Identification of PRO317 Receptors

Purified PRO317 is useful for characterization and purification of specific cell surface receptors and other binding molecules. Cells which respond to PRO317 by metabolic changes or other specific responses are likely to express a receptor for PRO317. Such receptors include, but are not limited to, receptors associated with and activated by tyrosine and serine/threonine kinases. See Kolodziejczyk and Hall, *supra*, for a review on known receptors for the TGF- superfamily. Candidate receptors for this superfamily fall into two primary groups, termed type I and type II receptors. Both types are serine/threonine kinases. Upon activation by the appropriate ligand, type I and type II receptors physically interact to form hetero-oligomers and subsequently activate intracellular signaling cascades, ultimately regulating gene transcription and expression. In addition, TGF- binds to a third receptor class, type III, a membrane-anchored proteoglycan lacking the kinase activity typical of signal transducing molecules.

PRO317 receptors or other PRO317-binding molecules may be identified by interaction with radiolabeled PRO317. Radioactive labels may be incorporated into PRO317 by various methods known in the art. A preferred embodiment is the labeling of primary amino groups in PRO317 with ^{125}I Bolton-Hunter reagent (Bolton and Hunter, Biochem. J., 133:529 (1973)), which has been used to label other polypeptides without concomitant loss of biological activity (Hebert *et al.*, J. Biol. Chem., 266:18989 (1991); McColl *et al.*, J. Immunol., 150:4550-4555 (1993)). Receptor-bearing cells are incubated with labeled PRO317. The cells are then washed to removed unbound PRO317, and receptor-bound PRO317 is quantified. The data obtained using different concentrations of PRO317 are used to calculate values for the number and affinity of receptors.

Labeled PRO317 is useful as a reagent for purification of its specific receptor. In one embodiment of affinity purification, PRO317 is covalently coupled to a chromatography column. Receptor-bearing cells are extracted, and the extract is passed over the column. The receptor binds to the column by virtue of its biological affinity for PRO317. The receptor is recovered from the column and subjected to N-terminal protein sequencing. This amino acid sequence is then used to design degenerate oligonucleotide probes for cloning the receptor gene.

In an alternative method, mRNA is obtained from receptor-bearing cells and made into a cDNA library. The library is transfected into a population of cells, and those cells expressing the receptor are selected using fluorescently labeled PRO317. The receptor is identified by recovering and sequencing recombinant DNA from highly labeled cells.

In another alternative method, antibodies are raised against the surface of receptor bearing cells, specifically monoclonal antibodies. The monoclonal antibodies are screened to identify those which inhibit the binding of labeled PRO317. These monoclonal antibodies are then used in affinity purification or expression cloning of the receptor.

Soluble receptors or other soluble binding molecules are identified in a similar manner. Labeled PRO317 is incubated with extracts or other appropriate materials derived from the uterus. After incubation,

PRO317 complexes larger than the size of purified PRO317 are identified by a sizing technique such as size-exclusion chromatography or density gradient centrifugation and are purified by methods known in the art. The soluble receptors or binding protein(s) are subjected to N-terminal sequencing to obtain information sufficient for database identification, if the soluble protein is known, or for cloning, if the soluble protein is unknown.

EXAMPLE 64: Determination of PRO317-Induced Cellular Response

The biological activity of PRO317 is measured, for example, by binding of an PRO317 of the invention to an PRO317 receptor. A test compound is screened as an antagonist for its ability to block binding of PRO317 to the receptor. A test compound is screened as an agonist of the PRO317 for its ability to bind an PRO317 receptor and influence the same physiological events as PRO317 using, for example, the KIRA-ELISA assay described by Sadick *et al.*, Analytical Biochemistry, 235:207-214 (1996) in which activation of a receptor tyrosine kinase is monitored by immuno-capture of the activated receptor and quantitation of the level of ligand-induced phosphorylation. The assay may be adapted to monitor PRO317-induced receptor activation through the use of an PRO317 receptor-specific antibody to capture the activated receptor. These techniques are also applicable to other PRO polypeptides described herein.

EXAMPLE 65: Use of PRO224 for Screening Compounds

PRO224 is expressed in a cell stripped of membrane proteins and capable of expressing PRO224. Low density lipoproteins having a detectable label are added to the cells and incubated for a sufficient time for endocytosis. The cells are washed. The cells are then analysed for label bound to the membrane and within the cell after cell lysis. Detection of the low density lipoproteins within the cell determines that PRO224 is within the family of low density lipoprotein receptor proteins. Members found within this family are then used for screening compounds which affect these receptors, and particularly the uptake of cholesterol via these receptors.

EXAMPLE 66: Ability of PRO Polypeptides to Inhibit Vascular Endothelial Growth Factor (VEGF) Stimulated Proliferation of Endothelial Cell Growth (Assay 9)

The ability of various PRO polypeptides to inhibit VEGF stimulated proliferation of endothelial cells was tested. Polypeptides testing positive in this assay are useful for inhibiting endothelial cell growth in mammals where such an effect would be beneficial, e.g., for inhibiting tumor growth.

Specifically, bovine adrenal cortical capillary endothelial cells (ACE) (from primary culture, maximum of 12-14 passages) were plated in 96-well plates at 500 cells/well per 100 microliter. Assay media included low glucose DMEM, 10% calf serum, 2 mM glutamine, and 1X penicillin/streptomycin/fungizone. Control wells included the following: (1) no ACE cells added; (2) ACE cells alone; (3) ACE cells plus 5 ng/ml FGF; (4) ACE cells plus 3 ng/ml VEGF; (5) ACE cells plus 3 ng/ml VEGF plus 1 ng/ml TGF-beta; and (6) ACE cells plus 3 ng/ml VEGF plus 5 ng/ml LIF. The test samples, poly-his tagged PRO polypeptides (in 100 microliter volumes), were then added to the wells (at dilutions of 1%, 0.1% and 0.01%, respectively). The cell cultures were incubated for 6-7 days at 37°C/5% CO_2. After the incubation, the media in the wells was aspirated, and the cells were washed 1X with PBS. An acid phosphatase reaction mixture (100 microliter; 0.1M sodium acetate, pH 5.5, 0.1% Triton X-100, 10 mM p-nitrophenyl phosphate) was then added to each well. After a 2

hour incubation at 37°C, the reaction was stopped by addition of 10 microliters 1N NaOH. Optical density (OD) was measured on a microplate reader at 405 nm.

The activity of PRO polypeptides was calculated as the percent inhibition of VEGF (3 ng/ml) stimulated proliferation (as determined by measuring acid phosphatase activity at OD 405 nm) relative to the cells without stimulation. TGF-beta was employed as an activity reference at 1 ng/ml, since TGF-beta blocks 70-90% of VEGF-stimulated ACE cell proliferation. The results are indicative of the utility of the PRO polypeptides in cancer therapy and specifically in inhibiting tumor angiogenesis. Numerical values (relative inhibition) are determined by calculating the percent inhibition of VEGF stimulated proliferation by the PRO polypeptides relative to cells without stimulation and then dividing that percentage into the percent inhibition obtained by TGF-β at 1 ng/ml which is known to block 70-90% of VEGF stimulated cell proliferation. The results are considered positive if the PRO polypeptide exhibits 30% or greater inhibition of VEGF stimulation of endothelial cell growth (relative inhibition 30% or greater).

The following polypeptides tested positive in this assay: PRO211, PRO217, PRO187, PRO219, PRO246, PRO228, PRO245, PRO221, PRO258, PRO301, PRO224, PRO272, PRO328, PRO331, PRO224, PRO328, PRO272, PRO301, PRO331 and PRO214.

EXAMPLE 67: Retinal Neuron Survival (Assay 52)

This example demonstrates that certain PRO polypeptides have efficacy in enhancing the survival of retinal neuron cells and, therefore, are useful for the therapeutic treatment of retinal disorders or injuries including, for example, treating sight loss in mammals due to retinitis pigmentosum, AMD, etc.

Sprague Dawley rat pups at postnatal day 7 (mixed population: glia and retinal neuronal types) are killed by decapitation following CO_2 anesthesia and the eyes are removed under sterile conditions. The neural retina is dissected away from the pigment epithelium and other ocular tissue and then dissociated into a single cell suspension using 0.25% trypsin in Ca^{2+}, Mg^{2+}-free PBS. The retinas are incubated at 37°C for 7-10 minutes after which the trypsin is inactivated by adding 1 ml soybean trypsin inhibitor. The cells are plated at 100,000 cells per well in 96 well plates in DMEM/F12 supplemented with N2 and with or without the specific test PRO polypeptide. Cells for all experiments are grown at 37°C in a water saturated atmosphere of 5% CO_2. After 2-3 days in culture, cells are stained with calcein AM then fixed using 4% paraformaldehyde and stained with DAPI for determination of total cell count. The total cells (fluorescent) are quantified at 20X objective magnification using CCD camera and NIH image software for MacIntosh. Fields in the well are chosen at random.

The effect of various concentration of PRO polypeptides are reported herein where percent survival is calculated by dividing the total number of calcein AM positive cells at 2-3 days in culture by the total number of DAPI-labeled cells at 2-3 days in culture. Anything above 30% survival is considered positive.

The following PRO polypeptides tested positive in this assay using polypeptide concentrations within the range of 0.01% to 1.0% in the assay: PRO220 and PRO346.

EXAMPLE 68: Rod Photoreceptor Cell Survival (Assay 56)

This assay shows that certain polypeptides of the invention act to enhance the survival/proliferation of

rod photoreceptor cells and, therefore, are useful for the therapeutic treatment of retinal disorders or injuries including, for example, treating sight loss in mammals due to retinitis pigmentosum, AMD, etc. Sprague Dawley rat pups at 7 day postnatal (mixed population: glia and retinal neuronal cell types) are killed by decapitation following CO_2 anesthesis and the eyes are removed under sterile conditions. The neural retina is dissected away form the pigment epithelium and other ocular tissue and then dissociated into a single cell suspension using 0.25% trypsin in Ca^{2+}, Mg^{2+}-free PBS. The retinas are incubated at 37°C for 7-10 minutes after which the trypsin is inactivated by adding 1 ml soybean trypsin inhibitor. The cells are plated at 100,000 cells per well in 96 well plates in DMEM/F12 supplemented with N_2. Cells for all experiments are grown at 37°C in a water saturated atmosphere of 5% CO_2. After 2-3 days in culture, cells are fixed using 4% paraformaldehyde, and then stained using CellTracker Green CMFDA. Rho 4D2 (ascites or IgG 1:100), a monoclonal antibody directed towards the visual pigment rhodopsin is used to detect rod photoreceptor cells by indirect immunofluorescence. The results are calculated as % survival: total number of calcein - rhodopsin positive cells at 2-3 days in culture, divided by the total number of rhodopsin positive cells at time 2-3 days in culture. The total cells (fluorescent) are quantified at 20x objective magnification using a CCD camera and NIH image software for MacIntosh. Fields in the well are chosen at random.

The following polypeptides tested positive in this assay: PRO220 and PRO346.

EXAMPLE 69: Induction of Endothelial Cell Apoptosis (Assay 73)

The ability of PRO polypeptides to induce apoptosis in endothelial cells was tested in human venous umbilical vein endothelial cells (HUVEC, Cell Systems). A positive test in the assay is indicative of the usefulness of the polypeptide in therapeutically treating tumors as well as vascular disorders where inducing apoptosis of endothelial cells would be beneficial.

The cells were plated on 96-well microtiter plates (Amersham Life Science, cytostar-T scintillating microplate, RPNQ160, sterile, tissue-culture treated, individually wrapped), in 10% serum (CSG-medium, Cell Systems), at a density of 2 x 10^4 cells per well in a total volume of 100 μl. On day 2, test samples containing the PRO polypeptide were added in triplicate at dilutions of 1%, 0.33% and 0.11%. Wells without cells were used as a blank and wells with cells only were used as a negative control. As a positive control 1:3 serial dilutions of 50 μl of a 3x stock of staurosporine were used. The ability of the PRO polypeptide to induce apoptosis was determined by processing of the 96 well plates for detection of Annexin V, a member of the calcium and phospholipid binding proteins, to detect apoptosis.

0.2 ml Annexin V - Biotin stock solution (100 μg/ml) was diluted in 4.6 ml 2 x Ca^{2+} binding buffer and 2.5% BSA (1:25 dilution). 50 μl of the diluted Annexin V - Biotin solution was added to each well (except controls) to a final concentration of 1.0 μg/ml. The samples were incubated for 10-15 minutes with Annexin-Biotin prior to direct addition of ^{35}S-Streptavidin. ^{35}S-Streptavidin was diluted in 2x Ca^{2+} Binding buffer, 2.5% BSA and was added to all wells at a final concentration of 3 x 10^4 cpm/well. The plates were then sealed, centrifuged at 1000 rpm for 15 minutes and placed on orbital shaker for 2 hours. The analysis was performed on a 1450 Microbeta Trilux (Wallac). Percent above background represents the percentage amount of counts per minute above the negative controls. Percents greater than or equal to 30% above background are considered positive.

The following PRO polypeptides tested positive in this assay: PRO228, PRO217 and PRO301.

EXAMPLE 70: PDB12 Cell Inhibition (Assay 40)

This example demonstrates that various PRO polypeptides have efficacy in inhibiting protein production by PDB12 pancreatic ductal cells and are, therefore, useful in the therapeutic treatment of disorders which involve protein secretion by the pancreas, including diabetes, and the like.

PDB12 pancreatic ductal cells are plated on fibronectin coated 96 well plates at $1.5x10^3$ cells per well in 100 μL/180 μL of growth media. 100 μL of growth media with the PRO polypeptide test sample or negative control lacking the PRO polypeptide is then added to well, for a final volume of 200 μL. Controls contain growth medium containing a protein shown to be inactive in this assay. Cells are incubated for 4 days at 37°C. 20 μL of Alamar Blue Dye (AB) is then added to each well and the flourescent reading is measured at 4 hours post addition of AB, on a microtiter plate reader at 530 nm excitation and 590 nm emission. The standard employed is cells without Bovine Pituitary Extract (BPE) and with various concentrations of BPE. Buffer or CM controls from unknowns are run 2 times on each 96 well plate.

These assays allow one to calculate a percent decrease in protein production by comparing the Alamar Blue Dye calculated protein concentration produced by the PRO polypeptide-treated cells with the Alamar Blue Dye calculated protein concentration produced by the negative control cells. A percent decrease in protein production of greater than or equal to 25% as compared to the negative control cells is considered positive.

The following polypeptides tested positive in this assay: PRO211, PRO287, PRO301 and PRO293.

EXAMPLE 71: Stimulation of Adult Heart Hypertrophy (Assay 2)

This assay is designed to measure the ability of various PRO polypeptides to stimulate hypertrophy of adult heart. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of various cardiac insufficiency disorders.

Ventricular myocytes freshly isolated from adult (250g) Sprague Dawley rats are plated at 2000 cell/well in 180 μl volume. Cells are isolated and plated on day 1, the PRO polypeptide-containing test samples or growth medium only (negative control) (20 μl volume) is added on day 2 and the cells are then fixed and stained on day 5. After staining, cell size is visualized wherein cells showing no growth enhancement as compared to control cells are given a value of 0.0, cells showing small to moderate growth enhancement as compared to control cells are given a value of 1.0 and cells showing large growth enhancement as compared to control cells are given a value of 2.0. Any degree of growth enhancement as compared to the negative control cells is considered positive for the assay.

The following PRO polypeptides tested positive in this assay: PRO287, PRO301, PRO293 and PRO303.

EXAMPLE 72: PDB12 Cell Proliferation (Assay 29)

This example demonstrates that various PRO polypeptides have efficacy in inducing proliferation of PDB12 pancreatic ductal cells and are, therefore, useful in the therapeutic treatment of disorders which involve protein secretion by the pancreas, including diabetes, and the like.

PDB12 pancreatic ductal cells are plated on fibronectin coated 96 well plates at $1.5x10^3$ cells per well

in 100 μL/180 μL of growth media. 100 μL of growth media with the PRO polypeptide test sample or negative control lacking the PRO polypeptide is then added to well, for a final volume of 200 μL. Controls contain growth medium containing a protein shown to be inactive in this assay. Cells are incubated for 4 days at 37°C. 20 μL of Alamar Blue Dye (AB) is then added to each well and the flourescent reading is measured at 4 hours post addition of AB, on a microtiter plate reader at 530 nm excitation and 590 nm emission. The standard employed is cells without Bovine Pituitary Extract (BPE) and with various concentrations of BPE. Buffer or growth medium only controls from unknowns are run 2 times on each 96 well plate.

Percent increase in protein production is calculated by comparing the Alamar Blue Dye calculated protein concentration produced by the PRO polypeptide-treated cells with the Alamar Blue Dye calculated protein concentration produced by the negative control cells. A percent increase in protein production of greater than or equal to 25% as compared to the negative control cells is considered positive.

The following PRO polypeptides tested positive in this assay: PRO301 and PRO303.

EXAMPLE 73: Enhancement of Heart Neonatal Hypertrophy (Assay 1)

This assay is designed to measure the ability of PRO polypeptides to stimulate hypertrophy of neonatal heart. PRO polypeptides testing positive in this assay are expected to be useful for the therapeutic treatment of various cardiac insufficiency disorders.

Cardiac myocytes from 1-day old Harlan Sprague Dawley rats were obtained. Cells (180 μl at 7.5 x 10^4/ml, serum <0.1%, freshly isolated) are added on day 1 to 96-well plates previously coated with DMEM/F12 + 4% FCS. Test samples containing the test PRO polypeptide or growth medium only (negative control) (20 μl/well) are added directly to the wells on day 1. PGF (20 μl/well) is then added on day 2 at final concentration of 10^{-6} M. The cells are then stained on day 4 and visually scored on day 5, wherein cells showing no increase in size as compared to negative controls are scored 0.0, cells showing a small to moderate increase in size as compared to negative controls are scored 1.0 and cells showing a large increase in size as compared to negative controls are scored 2.0. A positive result in the assay is a score of 1.0 or greater.

The following polypeptides tested positive in this assay: PRO224 and PRO231.

EXAMPLE 74: Stimulatory Activity in Mixed Lymphocyte Reaction (MLR) Assay (Assay 24)

This example shows that certain polypeptides of the invention are active as a stimulator of the proliferation of stimulated T-lymphocytes. Compounds which stimulate proliferation of lymphocytes are useful therapeutically where enhancement of an immune response is beneficial. A therapeutic agent may take the form of antagonists of the polypeptide of the invention, for example, murine-human chimeric, humanized or human antibodies against the polypeptide.

The basic protocol for this assay is described in Current Protocols in Immunology, unit 3.12; edited by J E Coligan, A M Kruisbeek, D H Marglies, E M Shevach, W Strober, National Insitutes of Health, Published by John Wiley & Sons, Inc.

More specifically, in one assay variant, peripheral blood mononuclear cells (PBMC) are isolated from mammalian individuals, for example a human volunteer, by leukopheresis (one donor will supply stimulator PBMCs, the other donor will supply responder PBMCs). If desired, the cells are frozen in fetal bovine serum

and DMSO after isolation. Frozen cells may be thawed overnight in assay media (37°C, 5% CO_2) and then washed and resuspended to $3x10^6$ cells/ml of assay media (RPMI; 10% fetal bovine serum, 1% penicillin/streptomycin, 1% glutamine, 1% HEPES, 1% non-essential amino acids, 1% pyruvate). The stimulator PBMCs are prepared by irradiating the cells (about 3000 Rads).

The assay is prepared by plating in triplicate wells a mixture of:

100:1 of test sample diluted to 1% or to 0.1%,

50 :1 of irradiated stimulator cells, and

50 :1 of responder PBMC cells.

100 microliters of cell culture media or 100 microliter of CD4-IgG is used as the control. The wells are then incubated at 37°C, 5% CO_2 for 4 days. On day 5, each well is pulsed with tritiated thymidine (1.0 mC/well; Amersham). After 6 hours the cells are washed 3 times and then the uptake of the label is evaluated.

In another variant of this assay, PBMCs are isolated from the spleens of Balb/c mice and C57B6 mice. The cells are teased from freshly harvested spleens in assay media (RPMI; 10% fetal bovine serum, 1% penicillin/streptomycin, 1% glutamine, 1% HEPES, 1% non-essential amino acids, 1% pyruvate) and the PBMCs are isolated by overlaying these cells over Lympholyte M (Organon Teknika), centrifuging at 2000 rpm for 20 minutes, collecting and washing the mononuclear cell layer in assay media and resuspending the cells to $1x10^7$ cells/ml of assay media. The assay is then conducted as described above.

Positive increases over control are considered positive with increases of greater than or equal to 180% being preferred. However, any value greater than control indicates a stimulatory effect for the test protein.

The following PRO polypeptides tested positive in this assay: PRO245, PRO269, PRO217, PRO301, PRO266, PRO335, PRO331, PRO533 and PRO326.

EXAMPLE 75: Pericyte c-Fos Induction (Assay 93)

This assay shows that certain polypeptides of the invention act to induce the expression of c-fos in pericyte cells and, therefore, are useful not only as diagnostic markers for particular types of pericyte-associated tumors but also for giving rise to antagonists which would be expected to be useful for the therapeutic treatment of pericyte-associated tumors. Specifically, on day 1, pericytes are received from VEC Technologies and all but 5 ml of media is removed from flask. On day 2, the pericytes are trypsinized, washed, spun and then plated onto 96 well plates. On day 7, the media is removed and the pericytes are treated with 100 μl of PRO polypeptide test samples and controls (positive control = DME+5% serum +/- PDGF at 500 ng/ml; negative control = protein 32). Replicates are averaged and SD/CV are determined. Fold increase over Protein 32 (buffer control) value indicated by chemiluminescence units (RLU) luminometer reading verses frequency is plotted on a histogram. Two-fold above Protein 32 value is considered positive for the assay. ASY Matrix: Growth media = low glucose DMEM = 20% FBS + 1X pen strep + 1X fungizone. Assay Media = low glucose DMEM +5% FBS.

The following polypeptides tested positive in this assay: PRO214, PRO219, PRO221 and PRO224.

EXAMPLE 76: Ability of PRO Polypeptides to Stimulate the Release of Proteoglycans from Cartilage (Assay

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The ability of various PRO polypeptides to stimulate the release of proteoglycans from cartilage tissue was tested as follows.

The metacarphophalangeal joint of 4-6 month old pigs was aseptically dissected, and articular cartilage was removed by free hand slicing being careful to avoid the underlying bone. The cartilage was minced and cultured in bulk for 24 hours in a humidified atmosphere of 95% air, 5% CO_2 in serum free (SF) media (DME/F12 1:1) woth 0.1% BSA and 100U/ml penicillin and 100μg/ml streptomycin. After washing three times, approximately 100 mg of articular cartilage was aliquoted into micronics tubes and incubated for an additional 24 hours in the above SF media. PRO polypeptides were then added at 1% either alone or in combination with 18 ng/ml interleukin-1α, a known stimulator of proteoglycan release from cartilage tissue. The supernatant was then harvested and assayed for the amount of proteoglycans using the 1,9-dimethyl-methylene blue (DMB) colorimetric assay (Farndale and Buttle, Biochem. Biophys. Acta 883:173-177 (1985)). A positive result in this assay indicates that the test polypeptide will find use, for example, in the treatment of sports-related joint problems, articular cartilage defects, osteoarthritis or rheumatoid arthritis.

When various PRO polypeptides were tested in the above assay, the polypeptides demonstrated a marked ability to stimulate release of proteoglycans from cartilage tissue both basally and after stimulation with interleukin-1α and at 24 and 72 hours after treatment, thereby indicating that these PRO polypeptides are useful for stimulating proteoglycan release from cartilage tissue. As such, these PRO polypeptides are useful for the treatment of sports-related joint problems, articular cartilage defects, osteoarthritis or rheumatoid arthritis. The polypeptides testing positive in this assay are : PRO211.

EXAMPLE 77: Skin Vascular Permeability Assay (Assay 64)

This assay shows that certain polypeptides of the invention stimulate an immune response and induce inflammation by inducing mononuclear cell, eosinophil and PMN infiltration at the site of injection of the animal. Compounds which stimulate an immune response are useful therapeutically where stimulation of an immune response is beneficial. This skin vascular permeability assay is conducted as follows. Hairless guinea pigs weighing 350 grams or more are anesthetized with ketamine (75-80 mg/Kg) and 5 mg/Kg xylazine intramuscularly (IM). A sample of purified polypeptide of the invention or a conditioned media test sample is injected intradermally onto the backs of the test animals with 100 μl per injection site. It is possible to have about 10-30, preferably about 16-24, injection sites per animal. One μl of Evans blue dye (1% in physiologic buffered saline) is injected intracardially. Blemishes at the injection sites are then measured (mm diameter) at 1 hr and 6 hr post injection. Animals were sacrificed at 6 hrs after injection. Each skin injection site is biopsied and fixed in formalin. The skins are then prepared for histopathologic evaluation. Each site is evaluated for inflammatory cell infiltration into the skin. Sites with visible inflammatory cell inflammation are scored as positive. Inflammatory cells may be neutrophilic, eosinophilic, monocytic or lymphocytic. At least a minimal perivascular infiltrate at the injection site is scored as positve, no infiltrate at the site of injection is scored as negative.

The following polypeptides tested positive in this assay: PRO245, PRO217, PRO326, PRO266,

PRO272, PRO301, PRO331 and PRO335.

EXAMPLE 78: Enhancement of Heart Neonatal Hypertrophy Induced by F2a (Assay 37)

This assay is designed to measure the ability of PRO polypeptides to stimulate hypertrophy of neonatal heart. PRO polypeptides testing positive in this assay are expected to be useful for the therapeutic treatment of various cardiac insufficiency disorders.

Cardiac myocytes from 1-day old Harlan Sprague Dawley rats were obtained. Cells (180 μl at 7.5 x 10^4/ml, serum $<0.1\%$, freshly isolated) are added on day 1 to 96-well plates previously coated with DMEM/F12 + 4% FCS. Test samples containing the test PRO polypeptide (20 μl/well) are added directly to the wells on day 1. PGF (20 μl/well) is then added on day 2 at a final concentration of 10^{-6} M. The cells are then stained on day 4 and visually scored on day 5. Visual scores are based on cell size, wherein cells showing no increase in size as compared to negative controls are scored 0.0, cells showing a small to moderate increase in size as compared to negative controls are scored 1.0 and cells showing a large increase in size as compared to negative controls are scored 2.0. A score of 1.0 or greater is considered positive.

No PBS is included, since calcium concentration is critical for assay response. Plates are coated with DMEM/F12 plus 4% FCS (200 μl/well). Assay media included: DMEM/F12 (with 2.44 gm bicarbonate), 10 μg/ml transferrin, 1 μg/ml insulin, 1 μg/ml aprotinin, 2 mmol/L glutamine, 100 U/ml penicillin G, 100 μg/ml streptomycin. Protein buffer containing mannitol (4%) gave a positive signal (score 3.5) at 1/10 (0.4%) and 1/100 (0.04%), but not at 1/1000 (0.004%). Therefore the test sample buffer containing mannitol is not run.

The following PRO polypeptides tested positive in this assay: PRO224.

EXAMPLE 79: Inhibitory Activity in Mixed Lymphocyte Reaction (MLR) Assay (Assay 67)

This example shows that one or more of the polypeptides of the invention are active as inhibitors of the proliferation of stimulated T-lymphocytes. Compounds which inhibit proliferation of lymphocytes are useful therapeutically where suppression of an immune response is beneficial.

The basic protocol for this assay is described in Current Protocols in Immunology, unit 3.12; edited by J E Coligan, A M Kruisbeek, D H Marglies, E M Shevach, W Strober, National Insitutes of Health, Published by John Wiley & Sons, Inc.

More specifically, in one assay variant, peripheral blood mononuclear cells (PBMC) are isolated from mammalian individuals, for example a human volunteer, by leukopheresis (one donor will supply stimulator PBMCs, the other donor will supply responder PBMCs). If desired, the cells are frozen in fetal bovine serum and DMSO after isolation. Frozen cells may be thawed overnight in assay media (37°C, 5% CO_2) and then washed and resuspended to $3x10^6$ cells/ml of assay media (RPMI; 10% fetal bovine serum, 1% penicillin/streptomycin, 1% glutamine, 1% HEPES, 1% non-essential amino acids, 1% pyruvate). The stimulator PBMCs are prepared by irradiating the cells (about 3000 Rads).

The assay is prepared by plating in triplicate wells a mixture of:

100:1 of test sample diluted to 1% or to 0.1%,

50 :1 of irradiated stimulator cells, and

50 :1 of responder PBMC cells.

100 microliters of cell culture media or 100 microliter of CD4-IgG is used as the control. The wells are then incubated at 37°C, 5% CO_2 for 4 days. On day 5, each well is pulsed with tritiated thymidine (1.0 mC/well; Amersham). After 6 hours the cells are washed 3 times and then the uptake of the label is evaluated.

In another variant of this assay, PBMCs are isolated from the spleens of Balb/c mice and C57B6 mice. The cells are teased from freshly harvested spleens in assay media (RPMI; 10% fetal bovine serum, 1% penicillin/streptomycin, 1% glutamine, 1% HEPES, 1% non-essential amino acids, 1% pyruvate) and the PBMCs are isolated by overlaying these cells over Lympholyte M (Organon Teknika), centrifuging at 2000 rpm for 20 minutes, collecting and washing the mononuclear cell layer in assay media and resuspending the cells to $1x10^7$ cells/ml of assay media. The assay is then conducted as described above.

Any decreases below control is considered to be a positive result for an inhibitory compound, with decreases of less than or equal to 80% being preferred. However, any value less than control indicates an inhibitory effect for the test protein.

The following polypeptide tested positive in this assay: PRO235, PRO245 and PRO332.

EXAMPLE 80: Induction of Endothelial Cell Apoptosis (ELISA) (Assay 109)

The ability of PRO polypeptides to induce apoptosis in endothelial cells was tested in human venous umbilical vein endothelial cells (HUVEC, Cell Systems) using a 96-well format, in 0% serum media supplemented with 100 ng/ml VEGF, 0.1% BSA, 1X penn/strep. A positive result in this assay indicates the usefulness of the polypeptide for therapeutically treating any of a variety of conditions associated with undesired endothelial cell growth including, for example, the inhibition of tumor growth. The 96-well plates used were manufactured by Falcon (No. 3072). Coating of 96 well plates were prepared by allowing gelatinization to occur for >30 minutes with 100 μl of 0.2% gelatin in PBS solution. The gelatin mix was aspirated thoroughly before plating HUVEC cells at a final concentration of 2×10^4 cells/ml in 10% serum containing medium - 100 μl volume per well. The cells were grown for 24 hours before adding test samples containing the PRO polypeptide of interest.

To all wells, 100 μl of 0% serum media (Cell Systems) complemented with 100 ng/ml VEGF, 0.1% BSA, 1X penn/strep was added. Test samples containing PRO polypeptides were added in triplicate at dilutions of 1%, 0.33% and 0.11%. Wells without cells were used as a blank and wells with cells only were used as a negative control. As a positive control, 1:3 serial dilutions of 50 μl of a 3x stock of staurosporine were used. The cells were incubated for 24 to 35 hours prior to ELISA.

ELISA was used to determine levels of apoptosis preparing solutions according to the Boehringer Manual [Boehringer, Cell Death Detection ELISA plus, Cat No. 1 920 685]. Sample preparations: 96 well plates were spun down at 1 krpm for 10 minutes (200g); the supernatant was removed by fast inversion, placing the plate upside down on a paper towel to remove residual liquid. To each well, 200 μl of 1X Lysis buffer was added and incubation allowed at room temperature for 30 minutes without shaking. The plates were spun down for 10 minutes at 1 krpm, and 20 μl of the lysate (cytoplasmic fraction) was transferred into streptavidin coated MTP. 80 μl of immunoreagent mix was added to the 20 μl lystate in each well. The MTP was covered with adhesive foil and incubated at room tempearature for 2 hours by placing it on an orbital shaker (200 rpm). After two hours, the supernatant was removed by suction and the wells rinsed three times with 250 μl of 1X incubation

buffer per well (removed by suction). Substrate solution was added (100 μl) into each well and incubated on an orbital shaker at room temperature at 250 rpm until color development was sufficient for a photometric analysis (approx. after 10-20 minutes). A 96 well reader was used to read the plates at 405 nm, reference wavelength, 492 nm. The levels obtained for PIN 32 (control buffer) was set to 100%. Samples with levels >130% were considered positive for induction of apoptosis.

The following PRO polypeptides tested positive in this assay: PRO235.

EXAMPLE 81: Human Venous Endothelial Cell Calcium Flux Assay (Assay 68)

This assay is designed to determine whether PRO polypeptides of the present invention show the ability to stimulate calcium flux in human umbilical vein endothelial cells (HUVEC, Cell Systems). Calcium influx is a well documented response upon binding of certain ligands to their receptors. A test compound that results in a positive response in the present calcium influx assay can be said to bind to a specific receptor and activate a biological signaling pathway in human endothelial cells. This could ultimately lead, for example, to endothelial cell division, inhibition of endothelial cell proliferation, endothelial tube formation, cell migration, apoptosis, etc.

Human venous umbilical vein endothelial cells (HUVEC, Cell Systems) in growth media (50:50 without glycine, 1% glutamine, 10mM Hepes, 10% FBS, 10 ng/ml bFGF), were plated on 96-well microtiter ViewPlates-96 (Packard Instrument Company Part #6005182) microtiter plates at a cell density of 2 x 10^4 cells/well. The day after plating, the cells were washed three times with buffer (HBSS plus 10 mM Hepes), leaving 100 μl/well. Then 100 μl/well of 8 μM Fluo-3 (2x) was added. The cells were incubated for 1.5 hours at 37°C/5% CO_2. After incubation, the cells were then washed 3x with buffer (described above) leaving 100 μl/well. Test samples of the PRO polypeptides were prepared on different 96-well plates at 5x concentration in buffer. The positive control corresponded to 50 μM ionomycin (5x); the negative control corresponded to Protein 32. Cell plate and sample plates were run on a FLIPR (Molecular Devices) machine. The FLIPR machine added 25 μl of test sample to the cells, and readings were taken every second for one minute, then every 3 seconds for the next three minutes.

The fluorescence change from baseline to the maximum rise of the curve (Δ change) was calculated, and replicates averaged. The rate of fluorescence increase was monitored, and only those samples which had a Δ change greater than 1000 and a rise within 60 seconds, were considered positive.

The following PRO polypeptides tested positive in the present assay: PRO245.

EXAMPLE 82: Fibroblast (BHK-21) Proliferation (Assay 98)

This assay shows that certain PRO polypeptides of the invention act to induce proliferation of mammalian fibroblast cells in culture and, therefore, function as useful growth factors in mammalian systems. The assay is performed as follows. BHK-21 fibroblast cells plated in standard growth medium at 2500 cells/well in a total volume of 100 μl. The PRO polypeptide, β-FGF (positive control) or nothing (negative control) are then added to the wells in the presence of 1μg/ml of heparin for a total final volume of 200 μl. The cells are then incubated at 37°C for 6 to 7 days. After incubation, the media is removed, the cells are washed with PBS and then an acid phosphatase substrate reaction mixture (100 μl/well) is added. The cells are then incubated at

37°C for 2 hours. 10 μl per well of 1N NaOH is then added to stop the acid phosphatase reaction. The plates are then read at OD 405nm. A positive in the assay is acid phosphatase activity which is at least 50% above the negative control.

The following PRO polypeptide tested positive in this assay: PRO258.

EXAMPLE 83: Inhibition of Heart Adult Hypertrophy (Assay 42)

This assay is designed to measure the inhibition of heart adult hypertrophy. PRO polypeptides testing positive in this assay may find use in the therapeutic treatment of cardiac disorders associated with cardiac hypertrophy.

Ventricular myocytes are freshly isolated from adult (250g) Harlan Sprague Dawley rats and the cells are plated at 2000/well in 180 μl volume. On day two, test samples (20 μl) containing the test PRO polypeptide are added. On day five, the cells are fixed and then stained. An increase in ANP message can also be measured by PCR from cells after a few hours. Results are based on a visual score of cell size: 0 = no inhibition, -1 =small inhibition, -2 = large inhibition. A score of less than 0 is considered positive. Activity reference corresponds to phenylephrin (PE) at 0.1 mM, as a positive control. Assay media included: M199 (modified)-glutamine free, $NaHCO_3$, phenol red, supplemented with 100 nM insulin, 0.2% BSA, 5 mM cretine, 2 mM L-carnitine, 5 mM taurine, 100 U/ml penicillin G, 100 μg/ml streptomycin (CCT medium). Only inner 60 wells are used in 96 well plates. Of these, 6 wells are reserved for negative and positive (PE) controls.

The following PRO polypeptides provided a score of less than 0 in the above assay: PRO269.

EXAMPLE 84: Induction of c-fos in Endothelial Cells (Assay 34)

This assay is designed to determine whether PRO polypeptides show the ability to induce c-fos in endothelial cells. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of conditions or disorders where angiogenesis would be beneficial including, for example, wound healing, and the like (as would agonists of these PRO polypeptides). Antagonists of the PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of cancerous tumors.

Human venous umbilical vein endothelial cells (HUVEC, Cell Systems) in growth media (50% Ham's F12 w/o GHT: low glucose, and 50% DMEM without glycine: with NaHCO3, 1% glutamine, 10 mM HEPES, 10% FBS, 10 ng/ml bFGF) were plated on 96-well microtiter plates at a cell density of $1x10^4$ cells/well. The day after plating, the cells were starved by removing the growth media and treating the cells with 100 μl/well test samples and controls (positive control = growth media; negative control = Protein 32 buffer = 10 mM HEPES, 140 mM NaCl, 4% (w/v) mannitol, pH 6.8). The cells were incubated for 30 minutes at 37°C, in 5% CO_2. The samples were removed, and the first part of the bDNA kit protocol (Chiron Diagnostics, cat. #6005-037) was followed, where each capitalized reagent/buffer listed below was available from the kit.

Briefly, the amounts of the TM Lysis Buffer and Probes needed for the tests were calculated based on information provided by the manufacturer. The appropriate amounts of thawed Probes were added to the TM Lysis Buffer. The Capture Hybridization Buffer was warmed to room temperature. The bDNA strips were set up in the metal strip holders, and 100 μl of Capture Hybridization Buffer was added to each b-DNA well needed,

followed by incubation for at least 30 minutes. The test plates with the cells were removed from the incubator, and the media was gently removed using the vacuum manifold. 100 μl of Lysis Hybridization Buffer with Probes were quickly pipetted into each well of the microtiter plates. The plates were then incubated at 55°C for 15 minutes. Upon removal from the incubator, the plates were placed on the vortex mixer with the microtiter adapter head and vortexed on the #2 setting for one minute. 80 μl of the lysate was removed and added to the bDNA wells containing the Capture Hybridization Buffer, and pipetted up and down to mix. The plates were incubated at 53°C for at least 16 hours.

On the next day, the second part of the bDNA kit protocol was followed. Specifically, the plates were removed from the incubator and placed on the bench to cool for 10 minutes. The volumes of additions needed were calculated based upon information provided by the manufacturer. An Amplifier Working Solution was prepared by making a 1:100 dilution of the Amplifier Concentrate (20 fm/μl) in AL Hybridization Buffer. The hybridization mixture was removed from the plates and washed twice with Wash A. 50 μl of Amplifier Working Solution was added to each well and the wells were incubated at 53°C for 30 minutes. The plates were then removed from the incubator and allowed to cool for 10 minutes. The Label Probe Working Solution was prepared by making a 1:100 dilution of Label Concentrate (40 pmoles/μl) in AL Hybridization Buffer. After the 10-minute cool-down period, the amplifier hybridization mixture was removed and the plates were washed twice with Wash A. 50 μl of Label Probe Working Solution was added to each well and the wells were incubated at 53°C for 15 minutes. After cooling for 10 minutes, the Substrate was warmed to room temperature. Upon addition of 3 μl of Substrate Enhancer to each ml of Substrate needed for the assay, the plates were allowed to cool for 10 minutes, the label hybridization mixture was removed, and the plates were washed twice with Wash A and three times with Wash D. 50 μl of the Substrate Solution with Enhancer was added to each well. The plates were incubated for 30 minutes at 37°C and RLU was read in an appropriate luminometer.

The replicates were averaged and the coefficient of variation was determined. The measure of activity of the fold increase over the negative control (Protein 32/HEPES buffer described above) value was indicated by chemiluminescence units (RLU). The results are considered positive if the PRO polypeptide exhibits at least a two-fold value over the negative buffer control. Negative control = 1.00 RLU at 1.00% dilution. Positive control = 8.39 RLU at 1.00% dilution.

The following PRO polypeptides tested positive in this assay: PRO287.

EXAMPLE 85: Guinea Pig Vascular Leak (Assays 32 and 51)

This assay is designed to determine whether PRO polypeptides of the present invention show the ability to induce vascular permeability. Polypeptides testing positive in this assay are expected to be useful for the therapeutic treatment of conditions which would benefit from enhanced vascular permeability including, for example, conditions which may benefit from enhanced local immune system cell infiltration.

Hairless guinea pigs weighing 350 grams or more were anesthetized with Ketamine (75-80 mg/kg) and 5 mg/kg Xylazine intramuscularly. Test samples containing the PRO polypeptide or a physiological buffer without the test polypeptide are injected into skin on the back of the test animals with 100 μl per injection site intradermally. There were approximately 16-24 injection sites per animal. One ml of Evans blue dye (1% in PBS) is then injected intracardially. Skin vascular permeability responses to the compounds (*i.e.*, blemishes at

the injection sites of injection) are visually scored by measuring the diameter (in mm) of blue-colored leaks from the site of injection at 1 and 6 hours post administration of the test materials. The mm diameter of blueness at the site of injection is observed and recorded as well as the severity of the vascular leakage. Blemishes of at least 5 mm in diameter are considered positive for the assay when testing purified proteins, being indicative of the ability to induce vascular leakage or permeability. A response greater than 7 mm diameter is considered positive for conditioned media samples. Human VEGF at 0.1 μg/100 μl is used as a positive control, inducing a response of 15-23 mm diameter.

The following PRO polypeptides tested positive in this assay: PRO302 and PRO533.

EXAMPLE 86: Detection of Endothelial Cell Apoptosis (FACS) (Assay 96)

The ability of PRO polypeptides of the present invention to induce apoptosis in endothelial cells was tested in human venous umbilical vein endothelial cells (HUVEC, Cell Systems) in gelatinized T175 flasks using HUVEC cells below passage 10. PRO polypeptides testing positive in this assay are expected to be useful for therapeutically treating conditions where apoptosis of endothelial cells would be beneficial including, for example, the therapeutic treatment of tumors.

On day one, the cells were split [420,000 cells per gelatinized 6 cm dishes - (11 x 10^3cells/cm^2 Falcon, Primaria)] and grown in media containing serum (CS-C, Cell System) overnight or for 16 hours to 24 hours.

On day 2, the cells were washed 1x with 5 ml PBS ; 3 ml of 0% serum medium was added with VEGF (100 ng/ml); and 30 μl of the PRO test compound (final dilution 1%) or 0% serum medium (negative control) was added. The mixtures were incubated for 48 hours before harvesting.

The cells were then harvested for FACS analysis. The medium was aspirated and the cells washed once with PBS. 5 ml of 1 x trypsin was added to the cells in a T-175 flask, and the cells were allowed to stand until they were released from the plate (about 5-10 minutes). Trypsinization was stopped by adding 5 ml of growth media. The cells were spun at 1000 rpm for 5 minutes at 4°C. The media was aspirated and the cells were resuspended in 10 ml of 10% serum complemented medium (Cell Systems), 5 μl of Annexin-FITC (BioVison) added and chilled tubes were submitted for FACS. A positive result was determined to be enhanced apoptosis in the PRO polypeptide treated samples as compared to the negative control.

The following PRO polypeptides tested positive in this assay: PRO331.

EXAMPLE 87: Induction of c-fos in Cortical Neurons (Assay 83)

This assay is designed to determine whether PRO polypeptides show the ability to induce c-fos in cortical neurons. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of nervous system disorders and injuries where neuronal proliferation would be beneficial.

Cortical neurons are dissociated and plated in growth medium at 10,000 cells per well in 96 well plates. After aproximately 2 cellular divisions, the cells are treated for 30 minutes with the PRO polypeptide or nothing (negative control). The cells are then fixed for 5 minutes with cold methanol and stained with an antibody directed against phosphorylated CREB. mRNA levels are then calculated using chemiluminescence. A positive in the assay is any factor that results in at least a 2-fold increase in c-fos message as compared to the negative controls.

The following PRO polypeptides tested positive in this assay: PRO229 and PRO269.

EXAMPLE 88: Stimulation of Endothelial Tube Formation (Assay 85)

This assay is designed to determine whether PRO polypeptides show the ability to promote endothelial vacuole and lumen formation in the absence of exogenous growth factors. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of disorders where endothelial vacuole and/or lumen formation would be beneficial including, for example, where the stimulation of pinocytosis, ion pumping, vascular permeability and/or junctional formation would be beneficial.

HUVEC cells (passage <8 from primary) are mixed with type I rat tail collagen (final concentration 2.6 mg/ml) at a density of $6x10^5$ cells per ml and plated at 50 μl per well of M199 culture media supplemented with 1% FBS and 1 μM 6-FAM-FITC dye to stain the vacuoles while they are forming and in the presence of the PRO polypeptide. The cells are then incubated at 37°C/5% CO_2 for 48 hours, fixed with 3.7% formalin at room temperature for 10 minutes, washed 5 times with M199 medium and then stained with Rh-Phalloidin at 4°C overnight followed by nuclear staining with 4 μM DAPI. A positive result in the assay is when vacuoles are present in greater than 50% of the cells.

The following PRO polypeptides tested positive in this assay: PRO230.

EXAMPLE 89: Detection of Polypeptides That Affect Glucose and/or FFA Uptake in Skeletal Muscle (Assay 106)

This assay is designed to determine whether PRO polypeptides show the ability to affect glucose or FFA uptake by skeletal muscle cells. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of disorders where either the stimulation or inhibition of glucose uptake by skeletal muscle would be beneficial including, for example, diabetes or hyper- or hypo-insulinemia.

In a 96 well format, PRO polypeptides to be assayed are added to primary rat differentiated skeletal muscle, and allowed to incubate overnight. Then fresh media with the PRO polypeptide and +/- insulin are added to the wells. The sample media is then monitored to determine glucose and FFA uptake by the skeletal muscle cells. The insulin will stimulate glucose and FFA uptake by the skeletal muscle, and insulin in media without the PRO polypeptide is used as a positive control, and a limit for scoring. As the PRO polypeptide being tested may either stimulate or inhibit glucose and FFA uptake, results are scored as positive in the assay if greater than 1.5 times or less than 0.5 times the insulin control.

The following PRO polypeptides tested positive as either stimulators or inhibitors of glucose and/or FFA uptake in this assay: PRO187, PRO211, PRO221, PRO222, PRO224, PRO230, PRO239, PRO231, PRO245, PRO247, PRO258, PRO269, PRO328 and PRO533.

EXAMPLE 90: Rod Photoreceptor Cell Survival Assay (Assay 46)

This assay shows that certain polypeptides of the invention act to enhance the survival/proliferation of rod photoreceptor cells and, therefore, are useful for the therapeutic treatment of retinal disorders or injuries including, for example, treating sight loss in mammals due to retinitis pigmentosum, AMD, etc.

Sprague Dawley rat pups (postnatal day 7, mixed population: glia and netinal neural cell types) are

killed by decapitation following CO_2 anesthesia and the eyes removed under sterile conditions. The neural retina is dissected away from the pigment epithelium and other ocular tissue and then dissociated into a single cell suspension using 0.25% trypsin in Ca^{2+}, Mg^{2+}-free PBS. The retinas are incubated at 37°C in this solution for 7-10 minutes after which the trypsin is inactivated by adding 1 ml soybean trypsin inhibitor. The cells are plated at a density of approximately 10, 000 cells/ml into 96 well plates in DMEM/F12 supplemented with N_2. Cells for all experiments are grown at 37°C in a water saturated atmosphere of 5% CO_2. After 7-10 days in culture, the cells are stained using calcein AM or CellTracker Green CMFDA and then fixed using 4% paraformaldehyde. Rho 4D2 (ascities or IgG 1:100) monoclonal antibody directed towards the visual pigment rhodopsin is used to detect rod photoreceptor cells by indirect immunofluorescence. The results are calculated as % survival: total number of calcein - rhodopsin positive cells at 7-10 days in culture, divided by the total number of rhodopsin positive cells at time 7-10 days in culture. The total cells (fluorescent) are quantified at 20x objective magnification using a CCD camera and NIH image software for MacIntosh. Fields in the well are chosen at random.

The following polypeptides tested positive in this assay: PRO245.

Example 91: *In Vitro* Antitumor Assay (Assay 161)

The antiproliferative activity of various PRO polypeptides was determined in the investigational, disease-oriented *in vitro* anti-cancer drug discovery assay of the National Cancer Institute (NCI), using a sulforhodamine B (SRB) dye binding assay essentially as described by Skehan et al., J. Natl. Cancer Inst. 82:1107-1112 (1990). The 60 tumor cell lines employed in this study ("the NCI panel"), as well as conditions for their maintenance and culture *in vitro* have been described by Monks et al., J. Natl. Cancer Inst. 83:757-766 (1991). The purpose of this screen is to initially evaluate the cytotoxic and/or cytostatic activity of the test compounds against different types of tumors (Monks et al., *supra*; Boyd, Cancer: Princ. Pract. Oncol. Update 3(10):1-12 [1989]).

Cells from approximately 60 human tumor cell lines were harvested with trypsin/EDTA (Gibco), washed once, resuspended in IMEM and their viability was determined. The cell suspensions were added by pipet (100 μL volume) into separate 96-well microtiter plates. The cell density for the 6-day incubation was less than for the 2-day incubation to prevent overgrowth. Inoculates were allowed a preincubation period of 24 hours at 37°C for stabilization. Dilutions at twice the intended test concentration were added at time zero in 100 μL aliquots to the microtiter plate wells (1:2 dilution). Test compounds were evaluated at five half-log dilutions (1000 to 100,000-fold). Incubations took place for two days and six days in a 5% CO_2 atmosphere and 100% humidity.

After incubation, the medium was removed and the cells were fixed in 0.1 ml of 10% trichloroacetic acid at 40°C. The plates were rinsed five times with deionized water, dried, stained for 30 minutes with 0.1 ml of 0.4% sulforhodamine B dye (Sigma) dissolved in 1% acetic acid, rinsed four times with 1% acetic acid to remove unbound dye, dried, and the stain was extracted for five minutes with 0.1 ml of 10 mM Tris base [tris(hydroxymethyl)aminomethane], pH 10.5. The absorbance (OD) of sulforhodamine B at 492 nm was measured using a computer-interfaced, 96-well microtiter plate reader.

A test sample is considered positive if it shows at least 50% growth inhibitory effect at one or more concentrations. PRO polypeptides testing positive in this assay are shown in Table 7, where the abbreviations

are as follows:

NSCL = non-small cell lung carcinoma

CNS = central nervous system

Table 7

Test compound	Tumor Cell Line Type	Cell Line Designation
PRO211	NSCL	HOP62
PRO211	Leukemia	RPMI-8226
PRO211	Leukemia	HL-60 (TB)
PRO211	NSCL	NCI-H522
PRO211	CNS	SF-539
PRO211	Melanoma	LOX IMVI
PRO211	Breast	MDA-MB-435
PRO211	Leukemia	MOLT-4
PRO211	CNS	U251
PRO211	Breast	MCF7
PRO211	Leukemia	HT-60 (TB)
PRO211	Leukemia	MOLT-4
PRO211	NSCL	EKVX
PRO211	NSCL	NCI-H23

Table 7 (cont')

PRO211	NSCL	NCI-H322M
PRO211	NSCL	NCI-H460
PRO211	Colon	HCT-116
PRO211	Colon	HT29
PRO211	CNS	SF-268
PRO211	CNS	SF-295
PRO211	CNS	SNB-19
PRO211	CNS	U251
PRO211	Melanoma	LOX IMVI
PRO211	Melanoma	SK-MEL-5
PRO211	Melanoma	UACC-257
PRO211	Melanoma	UACC-62
PRO211	Ovarian	OVCAR-8
PRO211	Renal	RXF 393
PRO211	Breast	MCF7
PRO211	Breast	NCI/ADR-REHS 578T
PRO211	Breast	T-47D
PRO211	Leukemia	HL-60 (TB)
PRO211	Leukemia	SR
PRO211	NSCL	NCI-H23
PRO211	Colon	HCT-116
PRO211	Melanoma	LOX-IMVI
PRO211	Melanoma	SK-MEL-5
PRO211	Breast	T-47D
PRO228	Leukemia	MOLT-4
PRO228	NSCL	EKVX
PRO228	Colon	KM12
PRO228	Melanoma	UACC-62
PRO228	Ovarian	OVCAR-3
PRO228	Renal	TK10
PRO228	Renal	SN12C
PRO228	Breast	MCF7
PRO228	Leukemia	CCRF-CEM

PRO228	Leukemia	HL-60 (TB)
PRO228	Colon	COLO 205
PRO228	Colon	HCT-15
PRO228	Colon	KM12
PRO228	CNS	SF-268
PRO228	CNS	SNB-75
PRO228	Melanoma	LOX-IMVI
PRO228	Melanoma	SK-MEL2
PRO228	Melanoma	UACC-257
PRO228	Ovarian	IGROV1
PRO228	Ovarian	OVCAR-4
PRO228	Ovarian	OVCAR-5
PRO228	Ovarian	OVCAR-8
PRO228	Renal	786-0
PRO228	Renal	CAKI-1
PRO228	Renal	RXF 393
PRO228	Renal	TK-10
PRO228	Renal	UO-31
PRO228	Prostate	PC-3
PRO228	Prostate	DU-145
PRO228	Breast	MCF7
PRO228	Breast	NCI/ADR-REHS 578T

Table 7 (cont')

PRO228	Breast	MDA-MB-435MDA-N
PRO228	Breast	T-47D
PRO219	Leukemia	SR
PRO219	NSCL	NCI-H5222
PRO219	Breast	MCF7
PRO219	Leukemia	K-562; RPMI-8226
PRO219	NSCL	HOP-62; NCI-H322M
PRO219	NSCL	NCI -H460
PRO219	Colon	HT29; KM12; HCT-116
PRO219	CNS	SF-539; U251
PRO219	Prostate	DU-145
PRO219	Breast	MDA-N
PRO219	Ovarian	IGROV1
PRO219	NSCL	NCI-H226
PRO219	Leukemia	MOLT-4
PRO219	NSCL	A549/ATCC; EKVX; NCI-H23
PRO219	Colon	HCC-2998
PRO219	CNS	SF-295; SNB-19
PRO219	Melanoma	SK-MEL-2; SK-MEL-5
PRO219	Melanoma	UACC-257; UACC-62
PRO219	Ovarian	OCAR-4; SK-OV-3
PRO219	Renal	786-0; ACHN; CAKI-1; SN12C
PRO219	Renal	TK-10; UO-31
PRO219	Breast	NCI/ADR-RES; BT-549; T-47D
PRO219	Breast	MDA-MB-435
PRO221	Leukemia	CCRF-CEM
PRO221	Leukemia	MOLT-4
PRO221	NSCL	HOP-62
PRO221	Breast	MDA-N
PRO221	Leukemia	RPMI-8226; SR
PRO221	NSCL	NCI-H460
PRO221	Colon	HCC-2998
PRO221	Ovarian	IGROV1

PRO221	Renal	TK-10
PRO221	Breast	MCF7
PRO221	Leukemia	K-562
PRO221	Breast	MDA-MB-435
PRO224	Ovarian	OVCAR-4
PRO224	Renal	RXF 393
PRO224	Prostate	DU-145
PRO224	NSCL	HOP-62; NCI-H322M
PRO224	Melanoma	LOX IMVI
PRO224	Ovarian	OVCAR-8
PRO224	Leukemia	SR
PRO224	NSCL	NCI-H460
PRO224	CNS	SF-295
PRO224	Leukemia	RPMI-8226
PRO224	Breast	BT-549
PRO224	Leukemia	CCRF-CEM; LH-60 (TB)
PRO224	Colon	HCT-116
PRO224	Breast	MDA-MB-435
PRO224	Leukemia	HL-60 (TB)
PRO224	Colon	HCC-2998
PRO224	Prostate	PC-3
PRO224	CNS	U251

Table 7 (cont')

PRO224	Colon	HCT-15
PRO224	CNS	SF-539
PRO224	Renal	ACHN
PRO328	Leukemia	RPMI-8226
PRO328	NSCL	A549/ATCC; EKVX; HOP-62
PRO328	NSCL	NCI-H23; NCI-H322M
PRO328	Colon	HCT-15; KM12
PRO328	CNS	SF-295; SF-539; SNB-19; U251
PRO328	Melanoma	M14; UACC-257; UCAA-62
PRO328	Renal	786-0; ACHN
PRO328	Breast	MCF7
PRO328	Leukemia	SR
PRO328	Colon	NCI-H23
PRO328	Melanoma	SK-MEL-5
PRO328	Prostate	DU-145
PRO328	Melanoma	LOX IMVI
PRO328	Breast	MDA-MB-435
PRO328	Ovarian	OVCAR-3
PRO328	Breast	T-47D
PRO301	NSCL	NCI-H322M
PRO301	Leukemia	MOLT-4; SR
PRO301	NSCL	A549/ATCC; EKVX;
PRO301	NSCL	NCI-H23; NCI-460; NCI-H226
PRO301	Colon	COLO 205; HCC-2998;
PRO301	Colon	HCT-15; KM12; HT29;
PRO301	Colon	HCT-116
PRO301	CNS	SF-268; SF-295; SNB-19
PRO301	Melanoma	MALME-3M; SK-MEL-2;
PRO301	Melanoma	SK-MEL-5;UACC-257
PRO301	Melanoma	UACC-62
PRO301	Ovarian	IGROV1; OVCAR-4
PRO301	Ovarian	OVCAR-5
PRO301	Ovarian	OVCAR-8; SK0OV-3

PRO301	Renal	ACHN;CAKI-1; TK-10; UO-31
PRO301	Prostate	PC-3; DU-145
PRO301	Breast	NCI/ADR-RES; HS 578T
PRO301	Breast	MDA-MB-435;MDA-N; T-47D
PRO301	Melanoma	M14
PRO301	Leukemia	CCRF-CEM;HL-60(TB); K-562
PRO301	Leukemia	RPMI-8226
PRO301	Melanoma	LOX IMVI
PRO301	Renal	786-0; SN12C
PRO301	Breast	MCF7; MDA-MB-231/ATCC
PRO301	Breast	BT-549
PRO301	NSCL	HOP-62
PRO301	CNS	SF-539
PRO301	Ovarian	OVCAR-3
PRO326	NSCL	NCI-H322M
PRO326	CNS	SF295
PRO326	CNS	ST539
PRO326	CNS	U251

The results of these assays demonstrate that the positive testing PRO polypeptides are useful for inhibiting neoplastic growth in a number of different tumor cell types and may be used therapeutically therefor.. Antibodies against these PRO polypeptides are useful for affinity purification of these useful polypeptides. Nucleic acids encoding these PRO polypeptides are useful for the recombinant preparation of these polypeptides.

EXAMPLE 92: Gene Amplification

This example shows that certain PRO polypeptide-encoding genes are amplified in the genome of certain human lung, colon and/or breast cancers and/or cell lines. Amplification is associated with overexpression of the gene product, indicating that the polypeptides are useful targets for therapeutic intervention in certain cancers such as colon, lung, breast and other cancers and diagnostic determination of the presence of those cancers. Therapeutic agents may take the form of antagonists of the PRO polypeptide, for example, murine-human chimeric, humanized or human antibodies against a PRO polypeptide.

The starting material for the screen was genomic DNA isolated from a variety cancers. The DNA is quantitated precisely, *e.g.*, fluorometrically. As a negative control, DNA was isolated from the cells of ten normal healthy individuals which was pooled and used as assay controls for the gene copy in healthy individuals (not shown). The 5' nuclease assay (for example, TaqMan™) and real-time quantitative PCR (for example, ABI Prizm 7700 Sequence Detection System™ (Perkin Elmer, Applied Biosystems Division, Foster City, CA)), were used to find genes potentially amplified in certain cancers. The results were used to determine whether the DNA encoding the PRO polypeptide is over-represented in any of the primary lung or colon cancers or cancer cell lines or breast cancer cell lines that were screened. The primary lung cancers were obtained from individuals with tumors of the type and stage as indicated in Table 8. An explanation of the abbreviations used for the designation of the primary tumors listed in Table 8 and the primary tumors and cell lines referred to throughout this example are given below.

The results of the TaqMan™ are reported in delta (Δ) Ct units. One unit corresponds to 1 PCR cycle

or approximately a 2-fold amplification relative to normal, two units corresponds to 4-fold, 3 units to 8-fold amplification and so on. Quantitation was obtained using primers and a TaqMan™ fluorescent probe derived from the PRO polypeptide-encoding gene. Regions of the PRO polypeptide-encoding gene which are most likely to contain unique nucleic acid sequences and which are least likely to have spliced out introns are preferred for the primer and probe derivation, *e.g.*, 3'-untranslated regions. The sequences for the primers and probes (forward, reverse and probe) used for the PRO polypeptide gene amplification analysis were as follows:

PRO187 (DNA27864-1155)

27864.tm.p:

5'-GCAGATTTTGAGGACAGCCACCTCCA-3' (SEQ ID NO:381)

27864.tm.f:

5'-GGCCTTGCAGACAACCGT-3' (SEQ ID NO:382)

27864.tm.r:

5'-CAGACTGAGGGAGATCCGAGA-3' (SEQ ID NO:383)

27864.tm.p2:

5'-CAGCTGCCCTTCCCCAACCA-3' (SEQ ID NO:384)

27864.tm.f2:

5'-CATCAAGCGCCTCTACCA-3' (SEQ ID NO:385)

27864.tm.r2:

5'-CACAAACTCGAACTGCTTCTG-3' (SEQ ID NO:386)

PRO214 (DNA32286-1191):

32286.3utr-5:

5'-GGGCCATCACAGCTCCCT-3' (SEQ ID NO:387)

32286.3utr-3b:

5'-GGGATGTGGTGAACACAGAACA-3' (SEQ ID NO:388)

32286.3utr-probe:

5'-TGCCAGCTGCATGCTGCCAGTT-3' (SEQ ID NO:389)

PRO211 (DNA32292-1131):

32292.3utr-5:

5'-CAGAAGGATGTCCCGTGGAA-3' (SEQ ID NO:390)

32292.3utr-3:

5'-GCCGCTGTCCACTGCAG-3' (SEQ ID NO:391)

32292.3utr-probe.rc:

5'-GACGGCATCCTCAGGGCCACA-3' (SEQ ID NO:392)

PRO230 (DNA33223-1136):

33223.tm.p3:

5'-ATGTCCTCCATGCCCACGCG-3' (SEQ ID NO:393)

33223.tm.f3:

5'-GAGTGCGACATCGAGAGCTT-3' (SEQ ID NO:394)

33223.tm.r3:

5'-CCGCAGCCTCAGTGATGA-3' (SEQ ID NO:395)

33223.3utr-5:

5'-GAAGAGCACAGCTGCAGATCC-3' (SEQ ID NO:396)

33223.3utr-3:

5'-GAGGTGTCCTGGCTTTGGTAGT-3' (SEQ ID NO:397)

33223.3utr-probe:

5'-CCTCTGGCGCCCCCACTCAA-3' (SEQ ID NO:398)

PRO317 (DNA33461-1199):

33461.tm.f:

5'-CCAGGAGAGCTGGCGATG-3' (SEQ ID NO:399)

33461.tm.r:

5'-GCAAATTCAGGGCTCACTAGAGA-3' (SEQ ID NO:400)

33461.tm.p:

5'-CACAGAGCATTTGTCCATCAGCAGTTCAG-3' (SEQ ID NO:401)

PRO246 (DNA35639-1172):

35639.3utr-5:

5'-GGCAGAGACTTCCAGTCACTGA-3' (SEQ ID NO:402)

35639.3utr-3:

5'-GCCAAGGGTGGTGTTAGATAGG-3' (SEQ ID NO:403)

35639.3utr-probe:

5'-CAGGCCCCCTTGATCTGTACCCCA-3' (SEQ ID NO:404)

PRO533 (DNA49435-1219):

49435.tm.f:

5'-GGGACGTGCTTCTACAAGAACAG-3' (SEQ ID NO:405)

49435.tm.r:

5'-CAGGCTTACAATGTTATGATCAGACA-3' (SEQ ID NO:406)

49435.tm.p:

5'-TATTCAGAGTTTTCCATTGGCAGTGCCAGTT-3' (SEQ ID NO:407)

PRO343 (DNA43318-1217):

43318.tm.f1

5'-TCTACATCAGCCTCTCTGCGC-3' (SEQ ID NO:408)

43318.tm.p1

5'-CGATCTTCTCCACCCAGGAGCGG-3' (SEQ ID NO:409)

43318.tm.r1

5'-GGAGCTGCACCCCTTGC-3' (SEQ ID NO:237)

PRO232 (DNA34435-1140):

34435.3utr-5:

5'-GCCAGGCCTCACATTCGT-3' (SEQ ID NO:410)

DNA34435.3utr-probe:

5'-CTCCCTGAATGGCAGCCTGAGCA-3' (SEQ ID NO:411)

DNA34435.3utr-3:

5'-AGGTGTTTATTAAGGGCCTACGCT -3' (SEQ ID NO:412)

PRO269 (DNA38260-1180):

38260.tm.f:

5'-CAGAGCAGAGGGTGCCTTG-3' (SEQ ID NO:413

38260.tm.p:

5'-TGGCGGAGTCCCCTCTTGGCT-3' (SEQ ID NO:414)

38260.tm.r:

5'-CCCTGTTTCCCTATGCATCACT-3' (SEQ ID NO:415)

PRO304 (DNA39520-1217):

39520.tm.f:

5'-TCAACCCCTGACCCTTTCCTA-3' (SEQ ID NO:416)

39520.tm.p:

5'-GGCAGGGGACAAGCCATCTCTCCT-3' (SEQ ID NO:417)

39520.tm.r:

5'-GGGACTGAACTGCCAGCTTC -3' (SEQ ID NO:418)

PRO339 (DNA43466-1225):

43466.tm.f1:

5'-GGGCCCTAACCTCATTACCTTT-3' (SEQ ID NO:419)

43466.tm.p1:

5'-TGTCTGCCTCAGCCCCAGGAAGG-3' (SEQ ID NO:420)

43466.tm.r1:

5'-TCTGTCCACCATCTTGCCTTG -3' (SEQ ID NO:421)

The 5' nuclease assay reaction is a fluorescent PCR-based technique which makes use of the 5' exonuclease activity of Taq DNA polymerase enzyme to monitor amplification in real time. Two oligonucleotide primers (forward [.f] and reverse [.r]) are used to generate an amplicon typical of a PCR reaction. A third oligonucleotide, or probe (.p), is designed to detect nucleotide sequence located between the two PCR primers. The probe is non-extendible by Taq DNA polymerase enzyme, and is labeled with a reporter fluorescent dye and a quencher fluorescent dye. Any laser-induced emission from the reporter dye is quenched by the quenching dye when the two dyes are located close together as they are on the probe. During the amplification reaction, the Taq DNA polymerase enzyme cleaves the probe in a template-dependent manner. The resultant probe fragments disassociate in solution, and signal from the released reporter dye is free from the quenching effect of the second fluorophore. One molecule of reporter dye is liberated for each new molecule synthesized, and detection of the unquenched reporter dye provides the basis for quantitative interpretation of the data.

The 5' nuclease procedure is run on a real-time quantitative PCR device such as the ABI Prism 7700TM Sequence Detection. The system consists of a thermocycler, laser, charge-coupled device (CCD) camera and computer. The system amplifies samples in a 96-well format on a thermocycler. During amplification, laser-induced fluorescent signal is collected in real-time through fiber optics cables for all 96 wells, and detected at the CCD. The system includes software for running the instrument and for analyzing the data.

5' Nuclease assay data are initially expressed as Ct, or the threshold cycle. This is defined as the cycle at which the reporter signal accumulates above the background level of fluorescence. The ΔCt values are used as quantitative measurement of the relative number of starting copies of a particular target sequence in a nucleic acid sample when comparing cancer DNA results to normal human DNA results.

Table 8 describes the stage, T stage and N stage of various primary tumors which were used to screen the PRO polypeptide compounds of the invention.

Table 8
Primary Lung and Colon Tumor Profiles

Primary Tumor Stage	Stage	Other Stage	Dukes Stage	T Stage	N Stage
Human lung tumor AdenoCa (SRCC724) [LT1]	IIA			T1	N1
Human lung tumor SqCCa (SRCC725) [LT1a]	IIB			T3	N0
Human lung tumor AdenoCa (SRCC726) [LT2]	IB			T2	N0
Human lung tumor AdenoCa (SRCC727) [LT3]	IIIA			T1	N2
Human lung tumor AdenoCa (SRCC728) [LT4]	IB			T2	N0
Human lung tumor SqCCa (SRCC729) [LT6]	IB			T2	N0
Human lung tumor Aden/SqCCa (SRCC730) [LT7]	IA			T1	N0
Human lung tumor AdenoCa (SRCC731) [LT9]	IB			T2	N0
Human lung tumor SqCCa (SRCC732) [LT10]	IIB			T2	N1
Human lung tumor SqCCa (SRCC733) [LT11]	IIA			T1	N1
Human lung tumor AdenoCa (SRCC734) [LT12]	IV			T2	N0
Human lung tumor AdenoSqCCa (SRCC735)[LT13]	IB			T2	N0
Human lung tumor SqCCa (SRCC736) [LT15]	IB			T2	N0
Human lung tumor SqCCa (SRCC737) [LT16]	IB			T2	N0
Human lung tumor SqCCa (SRCC738) [LT17]	IIB			T2	N1
Human lung tumor SqCCa (SRCC739) [LT18]	IB			T2	N0
Human lung tumor SqCCa (SRCC740) [LT19]	IB			T2	N0
Human lung tumor LCCa (SRCC741) [LT21]	IIB			T3	N1
Human lung AdenoCa (SRCC811) [LT22]	1A			T1	N0
Human colon AdenoCa (SRCC742) [CT2]		M1	D	pT4	N0
Human colon AdenoCa (SRCC743) [CT3]			B	pT3	N0
Human colon AdenoCa (SRCC744) [CT8]			B	T3	N0
Human colon AdenoCa (SRCC745) [CT10]			A	pT2	N0
Human colon AdenoCa (SRCC746) [CT12]		MO, R1	B	T3	N0
Human colon AdenoCa (SRCC747) [CT14]		pMO, RO	B	pT3	pN0
Human colon AdenoCa (SRCC748) [CT15]		M1, R2	D	T4	N2
Human colon AdenoCa (SRCC749) [CT16]		pMO	B	pT3	pN0
Human colon AdenoCa (SRCC750) [CT17]			C1	pT3	pN1
Human colon AdenoCa (SRCC751) [CT1]		MO, R1	B	pT3	N0
Human colon AdenoCa (SRCC752) [CT4]			B	pT3	M0
Human colon AdenoCa (SRCC753) [CT5]		G2	C1	pT3	pN0
Human colon AdenoCa (SRCC754) [CT6]		pMO, RO	B	pT3	pN0
Human colon AdenoCa (SRCC755) [CT7]		G1	A	pT2	pN0
Human colon AdenoCa (SRCC756) [CT9]		G3	D	pT4	pN2
Human colon AdenoCa (SRCC757) [CT11]			B	T3	N0
Human colon AdenoCa (SRCC758) [CT18]		MO, RO	B	pT3	pN0

DNA Preparation:

DNA was prepared from cultured cell lines, primary tumors, normal human blood. The isolation was performed using purification kit, buffer set and protease and all from Quiagen, according to the manufacturer's instructions and the description below.

Cell culture lysis:

Cells were washed and trypsinized at a concentration of 7.5×10^8 per tip and pelleted by centrifuging at 1000 rpm for 5 minutes at 4°C, followed by washing again with 1/2 volume of PBS recentrifugation. The pellets were washed a third time, the suspended cells collected and washed 2x with PBS. The cells were then suspended into 10 ml PBS. Buffer C1 was equilibrated at 4°C. Qiagen protease #19155 was diluted into 6.25 ml cold ddH_20 to a final concentration of 20 mg/ml and equilibrated at 4°C. 10 ml of G2 Buffer was prepared by diluting Qiagen RNAse A stock (100 mg/ml) to a final concentration of 200 μg/ml.

Buffer C1 (10 ml, 4°C) and ddH2O (40 ml, 4°C) were then added to the 10 ml of cell suspension, mixed by inverting and incubated on ice for 10 minutes. The cell nuclei were pelleted by centrifuging in a Beckman swinging bucket rotor at 2500 rpm at 4°C for 15 minutes. The supernatant was discarded and the nuclei were suspended with a vortex into 2 ml Buffer C1 (at 4°C) and 6 ml ddH_2O, followed by a second 4°C centrifugation at 2500 rpm for 15 minutes. The nuclei were then resuspended into the residual buffer using 200 μl per tip. G2 buffer (10 ml) was added to the suspended nuclei while gentle vortexing was applied. Upon completion of buffer addition, vigorous vortexing was applied for 30 seconds. Quiagen protease (200 μl, prepared as indicated above) was added and incubated at 50°C for 60 minutes. The incubation and centrifugation was repeated until the lysates were clear (*e.g.*, incubating additional 30-60 minutes, pelleting at 3000 x g for 10 min., 4°C).

Solid human tumor sample preparation and lysis:

Tumor samples were weighed and placed into 50 ml conical tubes and held on ice. Processing was limited to no more than 250 mg tissue per preparation (1 tip/preparation). The protease solution was freshly prepared by diluting into 6.25 ml cold ddH_2O to a final concentration of 20 mg/ml and stored at 4°C. G2 buffer (20 ml) was prepared by diluting DNAse A to a final concentration of 200 mg/ml (from 100 mg/ml stock). The tumor tissue was homogenated in 19 ml G2 buffer for 60 seconds using the large tip of the polytron in a laminar-flow TC hood in order to avoid inhalation of aerosols, and held at room temperature. Between samples, the polytron was cleaned by spinning at 2 x 30 seconds each in 2L ddH_20, followed by G2 buffer (50 ml). If tissue was still present on the generator tip, the apparatus was disassembled and cleaned.

Quiagen protease (prepared as indicated above, 1.0 ml) was added, followed by vortexing and incubation at 50°C for 3 hours. The incubation and centrifugation was repeated until the lysates were clear (*e.g.*, incubating additional 30-60 minutes, pelleting at 3000 x g for 10 min., 4°C).

Human blood preparation and lysis:

Blood was drawn from healthy volunteers using standard infectious agent protocols and citrated into 10 ml samples per tip. Quiagen protease was freshly prepared by dilution into 6.25 ml cold ddH_2O to a final concentration of 20 mg/ml and stored at 4°C. G2 buffer was prepared by diluting RNAse A to a final concentration of 200 μg/ml from 100 mg/ml stock. The blood (10 ml) was placed into a 50 ml conical tube and 10 ml C1 buffer and 30 ml ddH_2O (both previously equilibrated to 4°C) were added, and the components mixed by inverting and held on ice for 10 minutes. The nuclei were pelleted with a Beckman swinging bucket rotor at 2500 rpm, 4°C for 15 minutes and the supernatant discarded. With a vortex, the nuclei were suspended into 2 ml C1 buffer (4°C) and 6 ml ddH_2O (4°C). Vortexing was repeated until the pellet was white. The nuclei were then suspended into the residual buffer using a 200 μl tip. G2 buffer (10 ml) were added to the suspended nuclei while gently vortexing, followed by vigorous vortexing for 30 seconds. Quiagen protease was added (200 μl) and incubated at 50°C for 60 minutes. The incubation and centrifugation was repeated until the lysates were clear (*e.g.*, incubating additional 30-60 minutes, pelleting at 3000 x g for 10 min., 4°C).

Purification of cleared lysates:

(1) Isolation of genomic DNA:

Genomic DNA was equilibrated (1 sample per maxi tip preparation) with 10 ml QBT buffer. QF elution buffer was equilibrated at 50°C. The samples were vortexed for 30 seconds, then loaded onto equilibrated tips and drained by gravity. The tips were washed with 2 x 15 ml QC buffer. The DNA was eluted into 30 ml

silanized, autoclaved 30 ml Corex tubes with 15 ml QF buffer (50°C). Isopropanol (10.5 ml) was added to each sample, the tubes covered with parafin and mixed by repeated inversion until the DNA precipitated. Samples were pelleted by centrifugation in the SS-34 rotor at 15,000 rpm for 10 minutes at 4°C. The pellet location was marked, the supernatant discarded, and 10 ml 70% ethanol (4°C) was added. Samples were pelleted again by centrifugation on the SS-34 rotor at 10,000 rpm for 10 minutes at 4°C. The pellet location was marked and the supernatant discarded. The tubes were then placed on their side in a drying rack and dried 10 minutes at 37°C, taking care not to overdry the samples.

After drying, the pellets were dissolved into 1.0 ml TE (pH 8.5) and placed at 50°C for 1-2 hours. Samples were held overnight at 4°C as dissolution continued. The DNA solution was then transferred to 1.5 ml tubes with a 26 gauge needle on a tuberculin syringe. The transfer was repeated 5x in order to shear the DNA. Samples were then placed at 50°C for 1-2 hours.

(2) Quantitation of genomic DNA and preparation for gene amplification assay:

The DNA levels in each tube were quantified by standard A_{260}, A_{280} spectrophotometry on a 1:20 dilution (5 μl DNA + 95 μl ddH_2O) using the 0.1 ml quartz cuvetts in the Beckman DU640 spectrophotometer. A_{260}/A_{280} ratios were in the range of 1.8-1.9. Each DNA samples was then diluted further to approximately 200 ng/ml in TE (pH 8.5). If the original material was highly concentrated (about 700 ng/μl), the material was placed at 50°C for several hours until resuspended.

Fluorometric DNA quantitation was then performed on the diluted material (20-600 ng/ml) using the manufacturer's guidelines as modified below. This was accomplished by allowing a Hoeffer DyNA Quant 200 fluorometer to warm-up for about 15 minutes. The Hoechst dye working solution (#H33258, 10 μl, prepared within 12 hours of use) was diluted into 100 ml 1 x TNE buffer. A 2 ml cuvette was filled with the fluorometer solution, placed into the machine, and the machine was zeroed. pGEM 3Zf(+) (2 μl, lot #360851026) was added to 2 ml of fluorometer solution and calibrated at 200 units. An additional 2 μl of pGEM 3Zf(+) DNA was then tested and the reading confirmed at 400 +/- 10 units. Each sample was then read at least in triplicate. When 3 samples were found to be within 10% of each other, their average was taken and this value was used as the quantification value.

The fluorometricly determined concentration was then used to dilute each sample to 10 ng/μl in ddH_2O. This was done simultaneously on all template samples for a single TaqMan plate assay, and with enough material to run 500-1000 assays. The samples were tested in triplicate with Taqman™ primers and probe both B-actin and GAPDH on a single plate with normal human DNA and no-template controls. The diluted samples were used provided that the CT value of normal human DNA subtracted from test DNA was +/- 1 Ct. The diluted, lot-qualified genomic DNA was stored in 1.0 ml aliquots at -80°C. Aliquots which were subsequently to be used in the gene amplification assay were stored at 4°C. Each 1 ml aliquot is enough for 8-9 plates or 64 tests.

Gene amplification assay:

The PRO polypeptide compounds of the invention were screened in the following primary tumors and the resulting ΔCt values greater than or equal to 1.0 are reported in Table 9 below.

Table 9

ΔCt values in lung and colon primary tumors and cell line models

Primary Tumors or Cell lines	PRO187	PRO533	PRO214	PRO343	PRO211	PRO230	PRO246	PRO317	PRO232	PRO269	PRO304	PRO339
LT7								1.52		1.04		1.08
LT13	2.74		1.85	2.71	1.88	3.42	1.63	1.90		1.27	1,29	1.04
	2.98		1.83	2.23	2.26	3.22	1.68	2.24				
	2.44					2.84		2.93				
						2.15						
						2.75						
						2.53						
						1.82						
LT3			1.57		1.97		1.06	1.86				1.17
LT4					1.17			1.18				
LT9					1.42			1.04		1.80		1.03
LT12	2.70		1.38	2.23	1.51	2.86	1.54	2.54	2.40	1.14	1.15	1.26
	2.90		1.49	1.50	1.27	2.96	2.47	1.74				
	2.27					2.92						
						1.25						
						2.68						
						2.28						
						1.34						
LT30	1.67					2.13						
						1.36						

Table 9 (Continued)

ΔCt values in lung and colon primary tumors and cell line models

Primary Tumors or Cell lines	PRO187	PRO533	PRO214	PRO343	PRO211	PRO230	PRO246	PRO317	PRO232	PRO269	PRO304	PRO339
LT21					1.26	1.09	1.50					
LT1-a		1.02			1.18			1.29				
LT6								1.93				
LT10					1.96		1.07	2.57				
LT11		1.09	1.67	1.00	2.05	1.32	3.43	2.20		1.14	1.51	1.39
			1.80		1.89	1.14	1.41	2.33				
						1.54		1.02				
LT15	3.75		1.77	3.62	2.44	4.32	2.11	2.06	1.86	1.36	1.34	
	3.92		1.58	1.30	2.16	4.47	1.56	2.76				
	3.49					3.64		1.63				
						2.94						
						3.56						
						3.32						
						2.68						
LT16	2.10	1.66		1.70	1.25	1.15		1.55			1.00	
						2.04		1.08				
						1.83		1.33				

Table 9 (Continued)

ΔCt values in lung and colon primary tumors and cell line models

Primary Tumors or Cell lines	PRO187	PRO533	PRO214	PRO343	PRO211	PRO230	PRO246	PRO317	PRO232	PRO269	PRO304	PRO339
LT17		1.32	1.93 1.87	1.15	1.85 2.30	1.26 1.39 1.30 1.33 1.30	2.68 1.69	2.29 2.03 1.10	1.35	1.42	1.68	1.63
LT18				1.17					1.04			
LT19	4.05 3.99	1.67	2.09 1.98	3.82	2.42 2.55	4.05 4.92 4.93 3.78 4.76	1.91 1.68 1.16	2.51 2.03	1.21	1.60	1.15	
HF-000840						1.58						
Calu-1						1.08						
SW900					1.86							
CT2	3.56		2.49	1.95 3.49	1.42			2.75 2.36				
CT3			2.06	1.15		1.34						

Table 9 (Continued)

ΔCt values in lung and colon primary tumors and cell line models

Primary Tumors or Cell lines	PRO187	PRO533	PRO214	PRO343	PRO211	PRO230	PRO246	PRO317	PRO232	PRO269	PRO304	PRO339
CT8	1.01		1.48	1.29 1.58								
CT10	1.81		1.84	1.88 1.49		1.00		1.88 1.55				
CT12			1.81	1.74		1.13						
CT14	1.82		2.48	2.33 1.72				1.36 1.24				
CT15			1.63	2.06 1.41				1.33 1.04				
CT16			1.95	1.78		1.40						
CT17			2.04	2.40		1.74						
CT1	1.24 1.34		1.22 1.46		1.27 1.14	1.25			2.41			
CT4			1.36 1.42	1.77	1.33 1.02	1.32	1.10	1.17	2.05			

Table 9 (Continued)

ΔCt values in lung and colon primary tumors and cell line models

Primary Tumors or Cell lines	PRO187	PRO533	PRO214	PRO343	PRO211	PRO230	PRO246	PRO317	PRO232	PRO269	PRO304	PRO339
CT5	2.96 2.99		1.56 2.76	2.68	1.76 1.64	2.27	1.33 2.39		1.59			
CT6	1.10		1.33		1.01 1.14							
CT7	1.40			1.66		1.39			1.00			
CT9	1.39		1.16				1.09	1.24	1.13			
CT11	2.22 2.26		2.05 1.85	1.55	2.01 1.83	1.75	1.48 1.12		1.92			
HF000539						1.57						
SW620						1.14						
HF000611						4.64						
HF000733						1.93 2.33						
HF000716						1.68 2.82						
CT18									1.29			

Summary

Because amplification of the various DNA's as described above occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against these polypeptides would be expected to be useful in cancer therapy.

EXAMPLE 94: Detection of PRO Polypeptides That Affect Glucose or FFA Uptake by Primary Rat Adipocytes (Assay 94)

This assay is designed to determine whether PRO polypeptides show the ability to affect glucose or FFA uptake by adipocyte cells. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of disorders where either the stimulation or inhibition of glucose uptake by adipocytes would be beneficial including, for example, obesity, diabetes or hyper- or hypo-insulinemia.

In a 96 well format, PRO polypeptides to be assayed are added to primary rat adipocytes, and allowed to incubate overnight. Samples are taken at 4 and 16 hours and assayed for glycerol, glucose and FFA uptake. After the 16 hour incubation, insulin is added to the media and allowed to incubate for 4 hours. At this time, a sample is taken and glycerol, glucose and FFA uptake is measured. Media containing insulin without the PRO polypeptide is used as a positive reference control. As the PRO polypeptide being tested may either stimulate or inhibit glucose and FFA uptake, results are scored as positive in the assay if greater than 1.5 times or less than 0.5 times the insulin control.

The following PRO polypeptides tested positive as stimulators of glucose and/or FFA uptake in this assay: PRO221, PRO235, PRO245, PRO295, PRO301 and PRO332.

The following PRO polypeptides tested positive as inhibitors of glucose and/or FFA uptake in this assay: PRO214, PRO219, PRO228, PRO222, PRO231 and PRO265.

EXAMPLE 95: Chondrocyte Re-differentiation Assay (Assay 110)

This assay shows that certain polypeptides of the invention act to induce redifferentiation of chondrocytes, therefore, are expected to be useful for the treatment of various bone and/or cartilage disorders such as, for example, sports injuries and arthritis. The assay is performed as follows. Porcine chondrocytes are isolated by overnight collagenase digestion of articulary cartilage of metacarpophalangeal joints of 4-6 month old female pigs. The isolated cells are then seeded at 25,000 cells/cm^2 in Ham F-12 containing 10% FBS and 4 μg/ml gentamycin. The culture media is changed every third day and the cells are then seeded in 96 well plates at 5,000 cells/well in 100μl of the same media without serum and 100 μl of the test PRO polypeptide, 5 nM staurosporin (positive control) or medium alone (negative control) is added to give a final volume of 200 μl/well. After 5 days of incubation at 37°C, a picture of each well is taken and the differentiation state of the chondrocytes is determined. A positive result in the assay occurs when the redifferentiation of the chondrocytes is determined to be more similar to the positive control than the negative control.

The following polypeptide tested positive in this assay: PRO214, PRO219, PRO229, PRO222, PRO224, PRO230, PRO257, PRO272 and PRO301.

EXAMPLE 96: Fetal Hemoglobin Induction in an Erythroblastic Cell Line (Assay 107)

This assay is useful for screening PRO polypeptides for the ability to induce the switch from adult hemoglobin to fetal hemoglobin in an erythroblastic cell line. Molecules testing positive in this assay are expected to be useful for therapeutically treating various mammalian hemoglobin-associated disorders such as the various thalassemias. The assay is performed as follows. Erythroblastic cells are plated in standard growth medium at 1000 cells/well in a 96 well format. PRO polypeptides are added to the growth medium at a concentration of 0.2% or 2% and the cells are incubated for 5 days at 37°C. As a positive control, cells are treated with 100μM hemin and as a negative control, the cells are untreated. After 5 days, cell lysates are prepared and analyzed for the expression of gamma globin (a fetal marker). A positive in the assay is a gamma globin level at least 2-fold above the negative control.

The following polypeptides tested positive in this assay: PRO221 and PRO245.

EXAMPLE 97: Mouse Kidney Mesangial Cell Proliferation Assay (Assay 92)

This assay shows that certain polypeptides of the invention act to induce proliferation of mammalian kidney mesangial cells and, therefore, are useful for treating kidney disorders associated with decreased mesangial cell function such as Berger disease or other nephropathies associated with Schönlein-Henoch purpura, celiac disease, dermatitis herpetiformis or Crohn disease. The assay is performed as follows. On day one, mouse kidney mesangial cells are plated on a 96 well plate in growth media (3:1 mixture of Dulbecco's modified Eagle's medium and Ham's F12 medium, 95% fetal bovine serum, 5% supplemented with 14 mM HEPES) and grown overnight. On day 2, PRO polypeptides are diluted at 2 concentrations(1% and 0.1%) in serum-free medium and added to the cells. Control samples are serum-free medium alone. On day 4, 20μl of the Cell Titer 96 Aqueous one solution reagent (Progema) was added to each well and the colormetric reaction was allowed to proceed for 2 hours. The absorbance (OD) is then measured at 490 nm. A positive in the assay is anything that gives an absorbance reading which is at least 15% above the control reading.

The following polypeptide tested positive in this assay: PRO227.

EXAMPLE 98: Proliferation of Rat Utricular Supporting Cells (Assay 54)

This assay shows that certain polypeptides of the invention act as potent mitogens for inner ear supporting cells which are auditory hair cell progenitors and, therefore, are useful for inducing the regeneration of auditory hair cells and treating hearing loss in mammals. The assay is performed as follows. Rat UEC-4 utricular epithelial cells are aliquoted into 96 well plates with a density of 3000 cells/well in 200 μl of serum-containing medium at 33°C. The cells are cultured overnight and are then switched to serum-free medium at 37°C. Various dilutions of PRO polypeptides (or nothing for a control) are then added to the cultures and the cells are incubated for 24 hours. After the 24 hour incubation, ^{3}H-thymidine (1 μCi/well) is added and the cells are then cultured for an additional 24 hours. The cultures are then washed to remove unincorporated radiolabel, the cells harvested and Cpm per well determined. Cpm of at least 30% or greater in the PRO polypeptide treated cultures as compared to the control cultures is considered a positive in the assay.

The following polypeptides tested positive in this assay: PRO310 and PRO346.

EXAMPLE 99: Chondrocyte Proliferation Assay (Assay 111)

This assay is designed to determine whether PRO polypeptides of the present invention show the ability to induce the proliferation and/or redifferentiation of chondrocytes in culture. PRO polypeptides testing positive in this assay would be expected to be useful for the therapeutic treatment of various bone and/or cartilage disorders such as, for example, sports injuries and arthritis.

Porcine chondrocytes are isolated by overnight collagenase digestion of articular cartilage of the metacarpophalangeal joint of 4-6 month old female pigs. The isolated cells are then seeded at 25,000 cells/cm^2 in Ham F-12 containing 10% FBS and 4 μg/ml gentamycin. The culture media is changed every third day and the cells are reseeded to 25,000 cells/cm^2 every five days. On day 12, the cells are seeded in 96 well plates at 5,000 cells/well in 100μl of the same media without serum and 100 μl of either serum-free medium (negative control), staurosporin (final concentration of 5 nM; positive control) or the test PRO polypeptide are added to give a final volume of 200 μl/well. After 5 days at 37°C, 20 μl of Alamar blue is added to each well and the plates are incubated for an additional 3 hours at 37°C. The fluorescence is then measured in each well (Ex:530 nm; Em: 590 nm). The fluorescence of a plate containing 200 μl of the serum-free medium is measured to obtain the background. A positive result in the assay is obtained when the fluorescence of the PRO polypeptide treated sample is more like that of the positive control than the negative control.

The following PRO polypeptides tested positive in this assay: PRO219, PRO222, PRO317, PRO257, PRO265, PRO287, PRO272 and PRO533.

EXAMPLE 100: Inhibition of Heart Neonatal Hypertrophy Induced by LIF+ET-1 (Assay 74)

This assay is designed to determine whether PRO polypeptides of the present invention show the ability to inhibit neonatal heart hypertrophy induced by LIF and endothelin-1 (ET-1). A test compound that provides a positive response in the present assay would be useful for the therapeutic treatment of cardiac insufficiency diseases or disorders characterized or associated with an undesired hypertrophy of the cardiac muscle.

Cardiac myocytes from 1-day old Harlan Sprague Dawley rats (180 μl at 7.5 x 10^4/ml, serum <0.1, freshly isolated) are introduced on day 1 to 96-well plates previously coated with DMEM/F12 + 4%FCS. Test PRO polypeptide samples or growth medium alone (negative control) are then added directly to the wells on day 2 in 20 μl volume. LIF + ET-1 are then added to the wells on day 3. The cells are stained after an additional 2 days in culture and are then scored visually the next day. A positive in the assay occurs when the PRO polypeptide treated myocytes are visually smaller on the average or less numerous than the untreated myocytes.

The following PRO polypeptides tested positive in this assay: PRO238.

EXAMPLE 101: Tissue Expression Distribution

Oligonucleotide probes were constructed from some of the PRO polypeptide-encoding nucleotide sequences shown in the accompanying figures for use in quantitative PCR amplification reactions. The oligonucleotide probes were chosen so as to give an approximately 200-600 base pair amplified fragment from the 3' end of its associated template in a standard PCR reaction. The oligonucleotide probes were employed in standard quantitative PCR amplification reactions with cDNA libraries isolated from different human adult and/or fetal tissue sources and analyzed by agarose gel electrophoresis so as to obtain a quantitative determination of the level of expression of the PRO polypeptide-encoding nucleic acid in the various tissues tested. Knowledge

of the expression pattern or the differential expression of the PRO polypeptide-encoding nucleic acid in various different human tissue types provides a diagnostic marker useful for tissue typing, with or without other tissue-specific markers, for determining the primary tissue source of a metastatic tumor, and the like. These assays provided the following results.

DNA Molecule	Tissues With Significant Expression	Tissues Lacking Significant Expression
DNA34436-1238	lung, placenta, brain	testis
DNA35557-1137	lung, kidney, brain	placenta
DNA35599-1168	kidney, brain	liver, placenta
DNA35668-1171	liver, lung, kidney	placenta, brain
DNA36992-1168	liver, lung, kidney, brain	placenta
DNA39423-1182	kidney, brain	liver
DNA40603-1232	liver	brain, kidney, lung
DNA40604-1187	liver	brain, kidney, lung
DNA41379-1236	lung, brain	liver
DNA33206-1165	heart, spleen, dendrocytes	substantia nigra, hippocampus, cartilage, prostate, HUVEC
DNA34431-1177	spleen, HUVEC, cartilage, heart, uterus	brain, colon tumor, prostate, THP-1 macrophages
DNA41225-1217	HUVEC, uterus, colon tumor, cartilage, prostate	spleen, brain, heart, IM-9 lymphoblasts

EXAMPLE 102: *In situ* Hybridization

In situ hybridization is a powerful and versatile technique for the detection and localization of nucleic acid sequences within cell or tissue preparations. It may be useful, for example, to identify sites of gene expression, analyze the tissue distribution of transcription, identify and localize viral infection, follow changes in specific mRNA synthesis and aid in chromosome mapping.

In situ hybridization was performed following an optimized version of the protocol by Lu and Gillett, Cell Vision 1:169-176 (1994), using PCR-generated ^{33}P-labeled riboprobes. Briefly, formalin-fixed, paraffin-embedded human tissues were sectioned, deparaffinized, deproteinated in proteinase K (20 g/ml) for 15 minutes at 37°C, and further processed for *in situ* hybridization as described by Lu and Gillett, *supra*. A [^{33}-P] UTP-labeled antisense riboprobe was generated from a PCR product and hybridized at 55°C overnight. The slides were dipped in Kodak NTB2 nuclear track emulsion and exposed for 4 weeks.

^{33}P-Riboprobe synthesis

6.0 μl (125 mCi) of ^{33}P-UTP (Amersham BF 1002, SA < 2000 Ci/mmol) were speed vac dried. To each tube containing dried ^{33}P-UTP, the following ingredients were added:

2.0 μl 5x transcription buffer

1.0 μl DTT (100 mM)

2.0 μl NTP mix (2.5 mM : 10 μ; each of 10 mM GTP, CTP & ATP + 10 μl H_2O)

1.0 μl UTP (50 μM)

1.0 μl Rnasin

1.0 μl DNA template (1μg)

1.0 μl H_2O

1.0 μl RNA polymerase (for PCR products T3 = AS, T7 = S, usually)

The tubes were incubated at 37°C for one hour. 1.0 μl RQ1 DNase were added, followed by incubation at 37°C for 15 minutes. 90 μl TE (10 mM Tris pH 7.6/1mM EDTA pH 8.0) were added, and the mixture was pipetted onto DE81 paper. The remaining solution was loaded in a Microcon-50 ultrafiltration unit, and spun using program 10 (6 minutes). The filtration unit was inverted over a second tube and spun using program 2 (3 minutes). After the final recovery spin, 100 μl TE were added. 1 μl of the final product was pipetted on DE81 paper and counted in 6 ml of Biofluor II.

The probe was run on a TBE/urea gel. 1-3 μl of the probe or 5 μl of RNA Mrk III were added to 3 μl of loading buffer. After heating on a 95°C heat block for three minutes, the gel was immediately placed on ice. The wells of gel were flushed, the sample loaded, and run at 180-250 volts for 45 minutes. The gel was wrapped in saran wrap and exposed to XAR film with an intensifying screen in -70°C freezer one hour to overnight.

^{33}P-Hybridization

A. Pretreatment of frozen sections

The slides were removed from the freezer, placed on aluminium trays and thawed at room temperature for 5 minutes. The trays were placed in 55°C incubator for five minutes to reduce condensation. The slides were fixed for 10 minutes in 4% paraformaldehyde on ice in the fume hood, and washed in 0.5 x SSC for 5

minutes, at room temperature (25 ml 20 x SSC + 975 ml SQ H_2O). After deproteination in 0.5 μg/ml proteinase K for 10 minutes at 37°C (12.5 μl of 10 mg/ml stock in 250 ml prewarmed RNase-free RNAse buffer), the sections were washed in 0.5 x SSC for 10 minutes at room temperature. The sections were dehydrated in 70%, 95%, 100% ethanol, 2 minutes each.

B. Pretreatment of paraffin-embedded sections

The slides were deparaffinized, placed in SQ H_2O, and rinsed twice in 2 x SSC at room temperature, for 5 minutes each time. The sections were deproteinated in 20 μg/ml proteinase K (500 μl of 10 mg/ml in 250 ml RNase-free RNase buffer; 37°C, 15 minutes) - human embryo, or 8 x proteinase K (100 μl in 250 ml Rnase buffer, 37°C, 30 minutes) - formalin tissues. Subsequent rinsing in 0.5 x SSC and dehydration were performed as described above.

C. Prehybridization

The slides were laid out in a plastic box lined with Box buffer (4 x SSC, 50% formamide) - saturated filter paper. The tissue was covered with 50 μl of hybridization buffer (3.75g Dextran Sulfate + 6 ml SQ H_2O), vortexed and heated in the microwave for 2 minutes with the cap loosened. After cooling on ice, 18.75 ml formamide, 3.75 ml 20 x SSC and 9 ml SQ H_2O were added, the tissue was vortexed well, and incubated at 42°C for 1-4 hours.

D. Hybridization

1.0 x 10^6 cpm probe and 1.0 μl tRNA (50 mg/ml stock) per slide were heated at 95°C for 3 minutes. The slides were cooled on ice, and 48 μl hybridization buffer were added per slide. After vortexing, 50 μl ^{33}P mix were added to 50 μl prehybridization on slide. The slides were incubated overnight at 55°C.

E. Washes

Washing was done 2 x 10 minutes with 2xSSC, EDTA at room temperature (400 ml 20 x SSC + 16 ml 0.25M EDTA, V_f=4L), followed by RNaseA treatment at 37°C for 30 minutes (500 μl of 10 mg/ml in 250 ml Rnase buffer = 20 μg/ml), The slides were washed 2 x 10 minutes with 2 x SSC, EDTA at room temperature. The stringency wash conditions were as follows: 2 hours at 55°C, 0.1 x SSC, EDTA (20 ml 20 x SSC + 16 ml EDTA, V_f=4L).

F. Oligonucleotides

In situ analysis was performed on a variety of DNA sequences disclosed herein. The oligonucleotides employed for these analyses are as follows.

(1) DNA33094-1131 (PRO217)

p1 5'-GGATTCTAATACGACTCACTATAGGGCTCAGAAAAGCGCAACAGAGAA-3' (SEQ ID NO:348)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGATGTCTTCCATGCCAACCTTC-3' (SEQ ID NO:349)

(2) DNA33223-1136 (PRO230)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGGCGATGTCCACTGGGGCTAC-3' (SEQ ID

NO:350)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACGAGGAAGATGGGCGGATGGT-3' (SEQ ID NO:351)

(3) DNA34435-1140 (PRO232)

p1 5'-GGATTCTAATACGACTCACTATAGGGCACCCACGCGTCCGGCTGCTT-3' (SEQ ID NO:352)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACGGGGGACACCACGGACCAGA-3' (SEQ ID NO:353)

(4) DNA35639-1172 (PRO246)

p1 5'-GGATTCTAATACGACTCACTATAGGGCTTGCTGCGGTTTTTGTTCCTG-3' (SEQ ID NO:354)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGCTGCCGATCCCACTGGTATT-3' (SEQ ID NO:355)

(5) DNA49435-1219 (PRO533)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGGATCCTGGCCGGCCTCTG-3' (SEQ ID NO:356)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGCCCGGGCATGGTCTCAGTTA-3' (SEQ ID NO:357)

(6) DNA35638-1141 (PRO245)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGGGAAGATGGCGAGGAGGAG-3' (SEQ ID NO:358)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACCAAGGCCACAAACGGAAATC-3' (SEQ ID NO:359)

(7) DNA33089-1132 (PRO221)

p1 5'-GGATTCTAATACGACTCACTATAGGGCTGTGCTTTCATTCTGCCAGTA-3' (SEQ ID NO:360)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGGGTACAATTAAGGGGTGGAT-3' (SEQ ID NO:361)

(8) DNA35918-1174 (PRO258)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCCGCCTCGCTCCTGCTCCTG-3' (SEQ ID NO:362)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGGATTGCCGCGACCCTCACAG-3' (SEQ ID NO:363)

(9) DNA32286-1191 (PRO214)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCCCTCCTGCCTTCCCTGTCC-3' (SEQ ID NO:364)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGTGGTGGCCGCGATTATCTGC-3' (SEQ ID NO:365)

(10) DNA33221-1133 (PRO224)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGCAGCGATGGCAGCGATGAGG-3' (SEQ ID NO:366)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACAGACGGGGCAGAGGGAGTG-3' (SEQ ID NO:367)

(11) DNA35557-1137 (PRO234)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCAGGAGGCGTGAGGAGAAAC-3' (SEQ ID NO:368)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAAAGACATGTCATCGGGAGTGG-3' (SEQ ID NO:369)

(12) DNA33100-1159 (PRO229)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCGGGTGGAGGTGGAACAGAAA-3' (SEQ ID NO:370)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACACAGACAGAGCCCCATACGC-3' (SEQ ID NO:371)

(13) DNA34431-1177 (PRO263)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCAGGGAAATCCGGATGTCTC-3' (SEQ ID NO:372)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGTAAGGGGATGCCACCGAGTA-3' (SEQ ID NO:373)

(14) DNA38268-1188 (PRO295)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCAGCTACCCGCAGGAGGAGG-3' (SEQ ID NO:374)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGATCCCAGGTGATGAGGTCCAGA-3' (SEQ ID NO:375)

G. Results

In situ analysis was performed on a variety of DNA sequences disclosed herein. The results from these analyses are as follows.

(1) DNA33094-1131 (PRO217)

Highly distinctive expression pattern, that does not indicate an obvious biological function. In the human embryo it was expressed in outer smooth muscle layer of the GI tract, respiratiry cartilage, branching respiratory epithelium, osteoblasts, tendons, gonad, in the optic nerve head and developing dermis. In the adult expression was observed in the epidermal pegs of the chimp tongue, the basal epithelial/myoepithelial cells of the prostate and urinary bladder. Also expressed in the alveolar lining cells of the adult lung, mesenchymal cells juxtaposed to erectile tissue in the penis and the cerebral cortex (probably glial cells). In the kidney, expression was only seen in disease, in cells surrounding thyroidized renal tubules.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals,

thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, myocardium, aorta, spleen, lymph node, gall bladder, pancreas, lung, skin, eye (inc. retina), prostate, bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

(a) Chimp Tissues: Salivary gland, stomach, thyroid, parathyroid, skin, thymus, ovary, lymph node.

(b) Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum, penis.

(2) DNA33223-1136 (PRO230)

Sections show an intense signal associated with arterial and venous vessels in the fetus. In arteries the signal appeared to be confined to smooth-muscle/pericytic cells. The signal is also seen in capillary vessels and in glomeruli. It is not clear whether or not endothelial cells are expressing this mRNA. Expression is also observed in epithelial cells in the fetal lens. Strong expression was also seen in cells within placental trophoblastic villi, these cells lie between the trophoblast and the fibroblast-like cells that express HGF - uncertain histogenesis. In the adult, there was no evidence of expression and the wall of the aorta and most vessels appear to be negative. However, expression was seen over vascular channels in the normal prostate and in the epithelium lining the gallbladder. Insurers expression was seen in the vessels of the soft-tissue sarcoma and a renal cell carcinoma. In summary, this is a molecule that shows relatively specific vascular expression in the fetus as well as in some adult organs. Expression was also observed in the fetal lens and the adult gallbladder.

In a secondary screen, vascular expression was observed, similar to that observed above, seen in fetal blocks. Expression is on vascular smooth muscle, rather than endothelium. Expression also seen in smooth muscle of the developing oesophagus, so as reported previously, this molecule is not vascular specific. Expression was examined in 4 lung and 4 breast carcinomas. Substantial expression was seen in vascular smooth muscle of at least 3/4 lung cancers and 2/4 breast cancers. In addition, in one breast carcinoma, expression was observed in peritumoral stromal cells of uncertain histogenesis (possibly myofibroblasts). No endothelial cell expression was observed in this study.

(3) DNA34435-1140 (PRO232)

Strong expression in prostatic epithelium and bladder epithelium, lower level of expression in bronchial epithelium. High background / low level expression seen in a number of sites, including among others, bone, blood, chondrosarcoma, adult heart and fetal liver. It is felt that this level of signal represents background, partly because signal at this level was seen over the blood. All other tissues negative.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine,

spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis, testis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, spleen, lymph node, pancreas, lung,

eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: adrenal

Rhesus Monkey Tissues: Cerebral cortex, hippocampus

In a secondary screen, expression was observed in the epithelium of the prostate, the superficial layers of the urethelium of the urinary bladder, the urethelium lining the renal pelvis and the urethelium of the ureter (1 out of 2 experiments). The urethra of a rhesus monkey was negative; it is unclear whether this represents a true lack of expression by the urethra, or if it is the result of a failure of the probe to cross react with rhesus tissue. The findings in the prostate and bladder are similar to those previously described using an isotopic detection technique. Expression of the mRNA for this antigen is NOT prostate epithelial specific. The antigen may serve as a useful marker for urethelial derived tissues. Expression in the superficial, post-mitotic cells, of the urinary tract epithelium also suggest that it is unlikely to represent a specific stem cell marker, as this would be expected to be expressed specifically in basal epithelium.

(4) DNA35639-1172 (PRO246)

Strongly expressed in fetal vascular endothelium, including tissues of the CNS. Lower level of expression in adult vasculature, including the CNS. Not obviously expressed at higher levels in tumor vascular endothelium. Signal also seen over bone matrix and adult spleen, not obviously cell associated, probably related to non-specific background at these sites.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis, testis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, spleen, lymph node, pancreas, lung, eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: adrenal

Rhesus Monkey Tissues: Cerebral cortex, hippocampus

(5) DNA49435-1219 (PRO533)

Moderate expression over cortical neurones in the fetal brain. Expression over the inner aspect of the fetal retina, possible expression in the developing lens. Expression over fetal skin, cartilage, small intestine, placental villi and umbilical cord. In adult tissues there is an extremely high level of expression over the gallbladder epithelium. Moderate expression over the adult kidney, gastric and colonic epithelia. Low-level expression was observed over many cell types in many tissues, this may be related to stickiness of the probe, these data should therefore be interpreted with a degree of caution.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis, testis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, spleen, lymph node, pancreas, lung, eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: adrenal

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum.

(6) DNA35638-1141 (PRO245)

Expression observed in the endothelium lining a subset of fetal and placental vessels. Endothelial expression was confined to these tissue blocks. Expression also observed over intermediate trophoblast cells of placenta. Expression also observed tumor vasculature but not in the vasculature of normal tissues of the same type. All other tissues negative.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined: Liver, kidney, adrenal, myocardium, aorta, spleen, lymph node, pancreas, lung, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), penis, eye, bladder, stomach, gastric carcinoma, colon, colonic carcinoma, thyroid (chimp), parathyroid (chimp) ovary (chimp) and chondrosarcoma. Acetominophen induced liver injury and hepatic cirrhosis

(7) DNA33089-1132 (PRO221)

Specific expression over fetal cerebral white and grey matter, as well as over neurones in the spinal cord. Probe appears to cross react with rat. Low level of expression over cerebellar neurones in adult rhesus brain. All other tissues negative.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined: Liver, kidney, adrenal, myocardium, aorta, spleen, lymph node, pancreas, lung, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), penis, eye, bladder, stomach, gastric carcinoma, colon, colonic carcinoma and chondrosarcoma. Acetominophen induced liver injury and hepatic cirrhosis

(8) DNA35918-1174 (PRO258)

Strong expression in the nervous system. In the rhesus monkey brain expression is observed in cortical, hippocampal and cerebellar neurones. Expression over spinal neurones in the fetal spinal cord, the developing brain and the inner aspects of the fetal retina. Expression over developing dorsal root and autonomic ganglia as well as enteric nerves. Expression observed over ganglion cells in the adult prostate. In the rat, there is strong expression over the developing hind brain and spinal cord. Strong expression over interstitial cells in the

placental villi. All other tissues were negative.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined: Liver, kidney, renal cell carcinoma, adrenal, aorta, spleen, lymph node, pancreas, lung, myocardium, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), bladder, prostate, stomach, gastric carcinoma, colon, colonic carcinoma, thyroid (chimp), parathyroid (chimp) ovary (chimp) and chondrosarcoma. Acetominophen induced liver injury and hepatic cirrhosis.

(9) DNA32286-1191 (PRO214)

Fetal tissue: Low level throughout mesenchyme. Moderate expression in placental stromal cells in membranous tissues and in thyroid. Low level expression in cortical neurones. Adult tissue: all negative.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined include: Liver, kidney, adrenal, myocardium, aorta, spleen, lymph node, pancreas, lung and skin.

(10) DNA33221-1133 (PRO224)

Expression limited to vascular endothelium in fetal spleen, adult spleen, fetal liver, adult thyroid and adult lymph node (chimp). Additional site of expression is the developing spinal ganglia. All other tissues negative.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, myocardium, aorta, spleen, lymph node, pancreas, lung, skin, eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: Salivary gland, stomach, thyroid, parathyroid, skin, thymus, ovary, lymph node.

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum, penis.

(11) DNA35557-1137 (PRO234)

Specific expression over developing motor neurones in ventral aspect of the fetal spinal cord (will develop into ventral horns of spinal cord). All other tissues negative. Possible role in growth, differentiation and/or development of spinal motor neurons.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined: Liver, kidney, adrenal, myocardium, aorta, spleen, lymph node,

pancreas, lung, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), penis, eye, bladder, stomach, gastric carcinoma, colon, colonic carcinoma and chondrosarcoma. Acetominophen induced liver injury and hepatic cirrhosis

(12) DNA33100-1159 (PRO229)

Striking expression in mononuclear phagocytes (macrophages) of fetal and adult spleen, liver, lymph node and adult thymus (in tingible body macrophages). The highest expression is in the spleen. All other tissues negative. Localisation and homology are entirely consistent with a role as a scavenger receptor for cells of the reticuloendothelial system. Expression also observed in placental mononuclear cells.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, myocardium, aorta, spleen, lymph node, gall bladder, pancreas, lung, skin, eye (inc. retina), prostate, bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: Salivary gland, stomach, thyroid, parathyroid, skin, thymus, ovary, lymph node.

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum, penis.

(13) DNA34431-1177 (PRO263)

Widepread expression in human fetal tissues and placenta over mononuclear cells, probably macrophages +/- lymphocytes. The cellular distribution follows a perivascular pattern in many tissues. Strong expression also seen in epithelial cells of the fetal adrenal cortex. All adult tissues were negative.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined: Liver, kidney, adrenal, spleen, lymph node, pancreas, lung, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), bladder, stomach, colon and colonic carcinoma. Acetominophen induced liver injury and hepatic cirrhosis.

A secondary screen evidenced expression over stromal mononuclear cells probably histiocytes.

(14) DNA38268-1188 (PRO295)

High expression over ganglion cells in human fetal spinal ganglia and over large neurones in the anterior horns of the developing spinal cord. In the adult there is expression in the chimp adrenal medulla (neural), neurones of the rhesus monkey brain (hippocampus [+++] and cerebral cortex) and neurones in ganglia in the normal adult human prostate (the only section that contains ganglion cells, ie expression in this cell type is presumed NOT to be confined to the prostate). All other tissues negative.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, great vessels, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body

wall, pelvis, testis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, spleen, lymph node, pancreas, lung, eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: adrenal

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum.

EXAMPLE 103: Isolation of cDNA clones Encoding Human PRO1868

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in Example 1 above. This consensus sequence is herein designated DNA49803. Based up an observed homology between the DNA49803 consensus sequence and an EST sequence contained within the Incyte EST clone no. 2994689, Incyte EST clone no. 2994689 was purchased and its insert obtained and sequenced. The sequence of that insert is shown in Figure 123 and is herein designated DNA77624-2515.

The entire nucleotide sequence of DNA77624-2515 is shown in Figure 123 (SEQ ID NO:422). Clone DNA77624-2515 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 51-53 and ending at the stop codon at nucleotide positions 981-983 (Figure 123). The predicted polypeptide precursor is 310 amino acids long (Figure 124). The full-length PRO1868 protein shown in Figure 124 has an estimated molecular weight of about 35,020 daltons and a pI of about 7.90. Analysis of the full-length PRO1868 sequence shown in Figure 124 (SEQ ID NO:423) evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 30, a transmembrane domain from about amino acid 243 to about amino acid 263, potential N-glycosylation sites from about amino acid 104 to about amino acid 107 and from about amino acid 192 to about amino acid 195, a cAMP- and cGMP-dependent protein kinase phosphorylation site from about amino acid 107 to about amino acid 110, casein kinase II phosphorylation sites from about amino acid 106 to about amino acid 109 and from about amino acid 296 to about amino acid 299, a tyrosine kinase phosphorylation site from about amino acid 69 to about amino acid 77 and potential N-myristolation sites from about amino acid 26 to about amino acid 31, from about amino acid 215 to about amino acid 220, from about amino acid 226 to about amino acid 231, from about amino acid 243 to about amino acid 248, from about amino acid 244 to about amino acid 249 and from about amino acid 262 to about amino acid 267. Clone DNA77624-2515 has been deposited with ATCC on December 22, 1998 and is assigned ATCC deposit no. 203553.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using a WU-BLAST2 sequence alignment analysis of the full-length sequence shown in Figure 124 (SEQ ID NO:423), evidenced significant homology between the PRO1868 amino acid sequence and the following Dayhoff sequences: HGS_RC75, P_W61379, A33_HUMAN, P_W14146, P_W14158, AMAL_DROME, P_R77437, I38346, NCM2_HUMAN and PTPD_HUMAN.

EXAMPLE 104: Identification of Receptor/Ligand Interactions

In this assay, various PRO polypeptides are tested for ability to bind to a panel of potential receptor molecules for the purpose of identifying receptor/ligand interactions. The identification of a ligand for a known

receptor, a receptor for a known ligand or a novel receptor/ligand pair is useful for a variety of indications including, for example, targeting bioactive molecules (linked to the ligand or receptor) to a cell known to express the receptor or ligand, use of the receptor or ligand as a reagent to detect the presence of the ligand or receptor in a composition suspected of containing the same, wherein the composition may comprise cells suspected of expressing the ligand or receptor, modulating the growth of or another biological or immunological activity of a cell known to express or respond to the receptor or ligand, modulating the immune response of cells or toward cells that express the receptor or ligand, allowing the preparaion of agonists, antagonists and/or antibodies directed against the receptor or ligand which will modulate the growth of or a biological or immunological activity of a cell expressing the receptor or ligand, and various other indications which will be readily apparent to the ordinarily skilled artisan.

The assay is performed as follows. A PRO polypeptide of the present invention suspected of being a ligand for a receptor is expressed as a fusion protein containing the Fc domain of human IgG (an immunoadhesin). Receptor-ligand binding is detected by allowing interaction of the immunoadhesin polypeptide with cells (e.g. Cos cells) expressing candidate PRO polypeptide receptors and visualization of bound immunoadhesin with fluorescent reagents directed toward the Fc fusion domain and examination by microscope. Cells expressing candidate receptors are produced by transient transfection, in parallel, of defined subsets of a library of cDNA expression vectors encoding PRO polypeptides that may function as receptor molecules. Cells are then incubated for 1 hour in the presence of the PRO polypeptide immunoadhesin being tested for possible receptor binding. The cells are then washed and fixed with paraformaldehyde. The cells are then incubated with fluorescent conjugated antibody directed against the Fc portion of the PRO polypeptide immunoadhesin (e.g. FITC conjugated goat anti-human-Fc antibody). The cells are then washed again and examined by microscope. A positive interaction is judged by the presence of fluorescent labeling of cells transfected with cDNA encoding a particular PRO polypeptide receptor or pool of receptors and an absence of similar fluorescent labeling of similarly prepared cells that have been transfected with other cDNA or pools of cDNA. If a defined pool of cDNA expression vectors is judged to be positive for interaction with a PRO polypeptide immunoadhesin, the individual cDNA species that comprise the pool are tested individually (the pool is "broken down") to determine the specific cDNA that encodes a receptor able to interact with the PRO polypeptide immunoadhesin.

In another embodiment of this assay, an epitope-tagged potential ligand PRO polypeptide (e.g. 8 histidine "His" tag) is allowed to interact with a panel of potential receptor PRO polypeptide molecules that have been expressed as fusions with the Fc domain of human IgG (immunoadhesins). Following a 1 hour co-incubation with the epitope tagged PRO polypeptide, the candidate receptors are each immunoprecipitated with protein A beads and the beads are washed. Potential ligand interaction is determined by western blot analysis of the immunoprecipitated complexes with antibody directed towards the epitope tag. An interaction is judged to occur if a band of the anticipated molecular weight of the epitope tagged protein is observed in the western blot analysis with a candidate receptor, but is not observed to occur with the other members of the panel of potential receptors.

Using these assays, the following receptor/ligand interactions have been herein identified: PRO245 binds to PRO1868.

Deposit of Material

The following materials have been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD, USA (ATCC):

Material	ATCC Dep. No.	Deposit Date
DNA32292-1131	ATCC 209258	September 16, 1997
DNA33094-1131	ATCC 209256	September 16, 1997
DNA33223-1136	ATCC 209264	September 16, 1997
DNA34435-1140	ATCC 209250	September 16, 1997
DNA27864-1155	ATCC 209375	October 16, 1997
DNA36350-1158	ATCC 209378	October 16, 1997
DNA32290-1164	ATCC 209384	October 16, 1997
DNA35639-1172	ATCC 209396	October 17, 1997
DNA33092-1202	ATCC 209420	October 28, 1997
DNA49435-1219	ATCC 209480	November 21, 1997
DNA35638-1141	ATCC 209265	September 16, 1997
DNA32298-1132	ATCC 209257	September 16, 1997
DNA33089-1132	ATCC 209262	September 16, 1997
DNA33786-1132	ATCC 209253	September 16, 1997
DNA35918-1174	ATCC 209402	October 17, 1997
DNA37150-1178	ATCC 209401	October 17, 1997
DNA38260-1180	ATCC 209397	October 17, 1997
DNA39969-1185	ATCC 209400	October 17, 1997
DNA32286-1191	ATCC 209385	October 16, 1997
DNA33461-1199	ATCC 209367	October 15, 1997
DNA40628-1216	ATCC 209432	November 7, 1997
DNA33221-1133	ATCC 209263	September 16, 1997
DNA33107-1135	ATCC 209251	September 16, 1997
DNA35557-1137	ATCC 209255	September 16, 1997
DNA34434-1139	ATCC 209252	September 16, 1997
DNA33100-1159	ATCC 209373	October 16, 1997
DNA35600-1162	ATCC 209370	October 16, 1997
DNA34436-1238	ATCC 209523	December 10, 1997
DNA33206-1165	ATCC 209372	October 16, 1997
DNA35558-1167	ATCC 209374	October 16, 1997
DNA35599-1168	ATCC 209373	October 16, 1997
DNA36992-1168	ATCC 209382	October 16, 1997
DNA34407-1169	ATCC 209383	October 16, 1997
DNA35841-1173	ATCC 209403	October 17, 1997
DNA33470-1175	ATCC 209398	October 17, 1997
DNA34431-1177	ATCC 209399	October 17, 1997
DNA39510-1181	ATCC 209392	October 17, 1997
DNA39423-1182	ATCC 209387	October 17, 1997
DNA40620-1183	ATCC 209388	October 17, 1997
DNA40604-1187	ATCC 209394	October 17, 1997
DNA38268-1188	ATCC 209421	October 28, 1997
DNA37151-1193	ATCC 209393	October 17, 1997
DNA35673-1201	ATCC 209418	October 28, 1997
DNA40370-1217	ATCC 209485	November 21, 1997
DNA42551-1217	ATCC 209483	November 21, 1997
DNA39520-1217	ATCC 209482	November 21, 1997
DNA41225-1217	ATCC 209491	November 21, 1997
DNA43318-1217	ATCC 209481	November 21, 1997
DNA40587-1231	ATCC 209438	November 7, 1997
DNA41338-1234	ATCC 209927	June 2, 1998
DNA40981-1234	ATCC 209439	November 7, 1997
DNA37140-1234	ATCC 209489	November 21, 1997

DNA40982-1235	ATCC 209433	November 7, 1997
DNA41379-1236	ATCC 209488	November 21, 1997
DNA44167-1243	ATCC 209434	November 7, 1997
DNA39427-1179	ATCC 209395	October 17, 1997
DNA40603-1232	ATCC 209486	November 21, 1997
DNA43466-1225	ATCC 209490	November 21, 1997
DNA43046-1225	ATCC 209484	November 21, 1997
DNA35668-1171	ATCC 209371	October 16, 1997
DNA77624-2515	ATCC 203553	December 22, 1998

These deposit were made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and the Regulations thereunder (Budapest Treaty). This assures maintenance of a viable culture of the deposit for 30 years from the date of deposit. The deposits will be made available by ATCC under the terms of the Budapest Treaty, and subject to an agreement between Genentech, Inc. and ATCC, which assures permanent and unrestricted availability of the progeny of the culture of the deposit to the public upon issuance of the pertinent U.S. patent or upon laying open to the public of any U.S. or foreign patent application, whichever comes first, and assures availability of the progeny to one determined by the U.S. Commissioner of Patents and Trademarks to be entitled thereto according to 35 USC § 122 and the Commissioner's rules pursuant thereto (including 37 CFR § 1.14 with particular reference to 886 OG 638).

The assignee of the present application has agreed that if a culture of the materials on deposit should die or be lost or destroyed when cultivated under suitable conditions, the materials will be promptly replaced on notification with another of the same. Availability of the deposited material is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any government in accordance with its patent laws.

The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. The present invention is not to be limited in scope by the construct deposited, since the deposited embodiment is intended as a single illustration of certain aspects of the invention and any constructs that are functionally equivalent are within the scope of this invention. The deposit of material herein does not constitute an admission that the written description herein contained is inadequate to enable the practice of any aspect of the invention, including the best mode thereof, nor is it to be construed as limiting the scope of the claims to the specific illustrations that it represents. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and fall within the scope of the appended claims.